

27.3: Acid-Base Behavior of Amino Acids. Amino acids exist as a zwitterion: a dipolar ion having both a formal positive and formal negative charge (overall charge neutral).

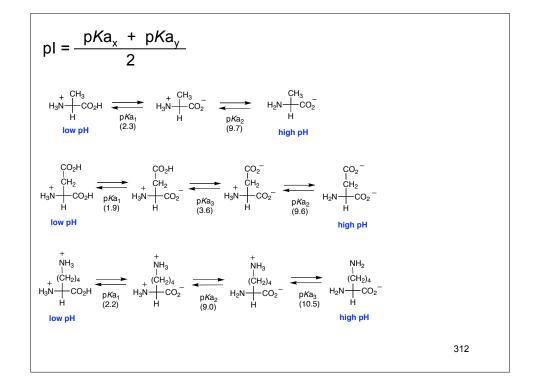


Amino acids are *amphoteric*: they can react as either an acid or a base. Ammonium ion acts as an acid, the carboxylate as a base.

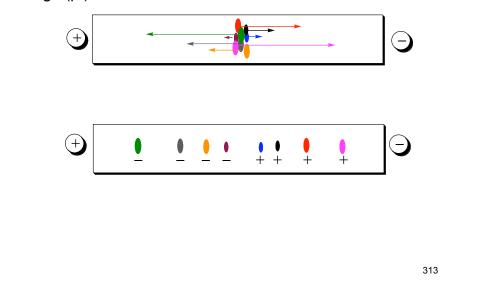
Isoelectric point (pI): The pH at which the amino acid exists largely in a neutral, zwitterionic form (influenced by the nature of the sidechain)

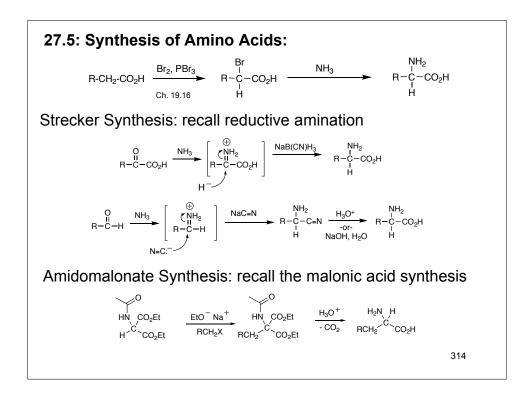
$$\begin{array}{c} \stackrel{+}{\overset{+}{H_{3}}}\stackrel{R}{\overset{+}{\longrightarrow}} CO_{2}H & \xrightarrow{H_{3}O^{+}} & \stackrel{+}{\overset{+}{\overset{+}{H_{3}}}}\stackrel{R}{\overset{+}{\overset{+}{\overset{+}{H_{3}}}} CO_{2}^{-} & \xrightarrow{HO^{-}} & \stackrel{R}{\overset{+}{\overset{+}{\underset{H}}} O^{-}_{2} & \stackrel{R}{\overset{+}{\underset{H}}} O^{-}_{2} \\ \hline \begin{array}{c} \stackrel{H}{\overset{+}{\underset{H}}} O^{+}_{2} & \stackrel{H}{\overset{+}{\underset{H}}} O^{+}_{2} & \stackrel{H}{\overset{+}{\underset{H}}} O^{-}_{2} & \stackrel{H}{\overset{H}} O^{-}_{2} & \stackrel{H}{\overset{H} O^{-}_{2} & \stackrel{H}{\overset{H}} O^{-}_{2} & \stackrel{H}{\overset{H} O^{-}_{2} & \stackrel{H}{\overset{H}} O^{-}_{2} & \stackrel{H}{\overset{H} O^{-}_{2} & \stackrel{H}{\overset{H}$$

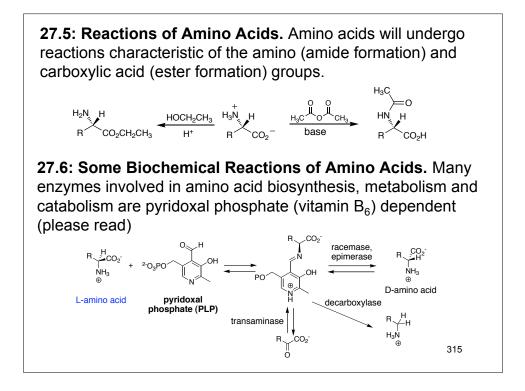
Table 27.2 (p. 1115) & 27.2 (p. 1116)

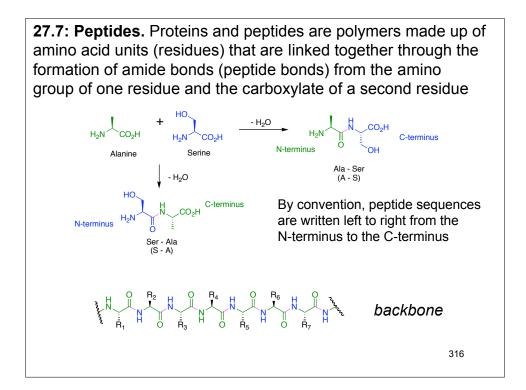


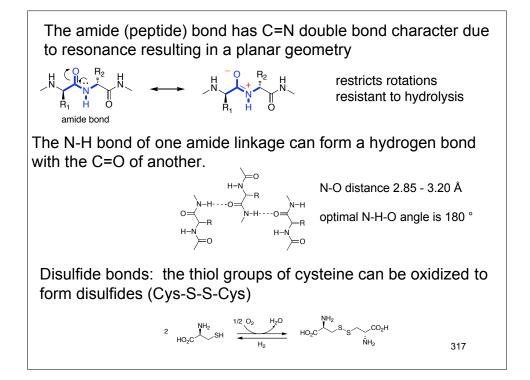
Electrophoresis: separation of polar compounds based on their mobility through a solid support. The separation is based on charge (pl) or molecular mass.

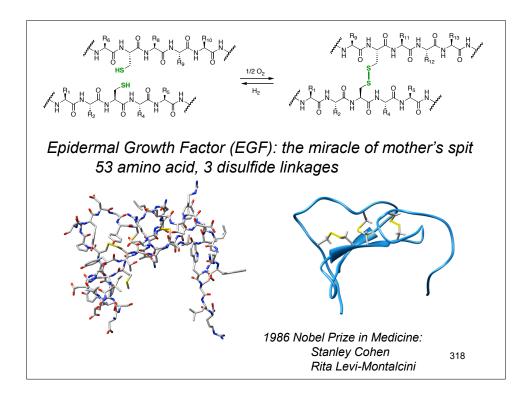


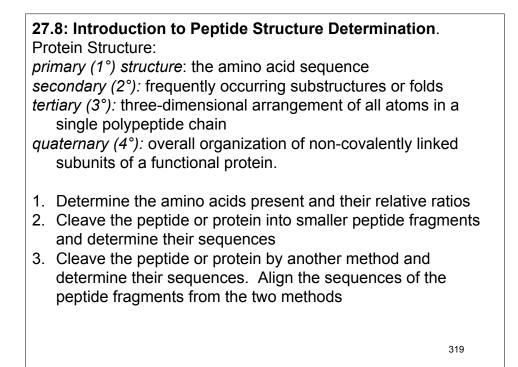


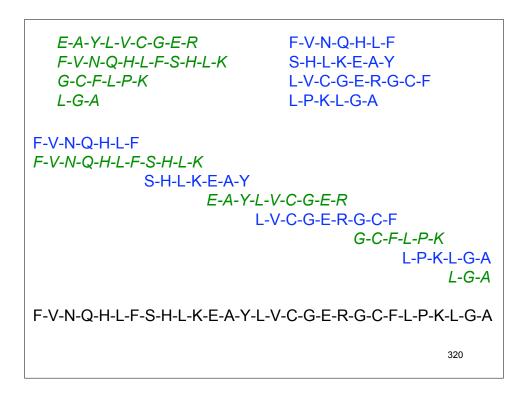


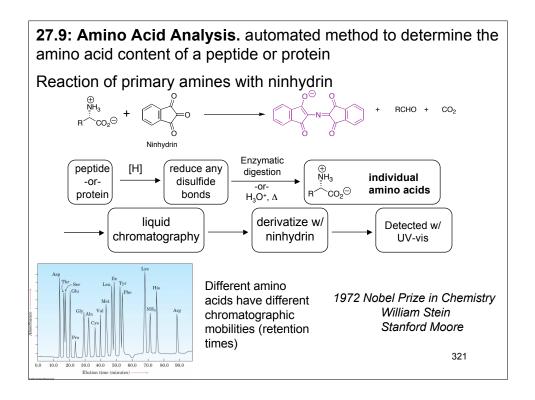


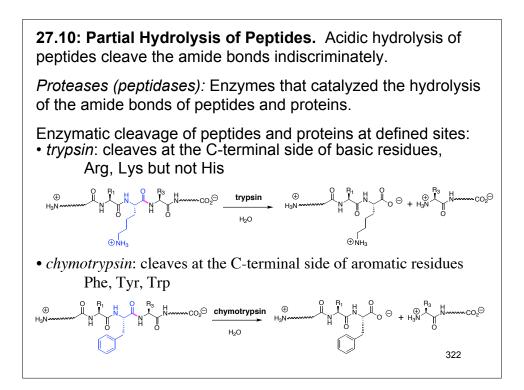










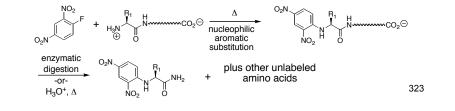


Trypsin and chymotrypsin are endopeptidases

Carboxypeptidase: Cleaves the amide bond of the C-terminal amino acid (exopeptidase)

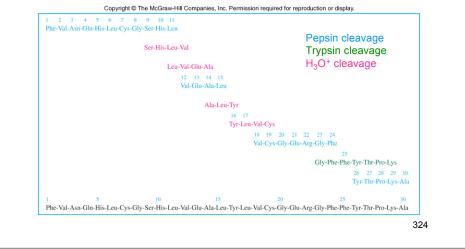
27.11: End Group Analysis. The C-terminal AA is identified by treating with peptide with carboxypeptidase, then analyzing by liquid chormatography (AA Analysis).

N-labeling: The peptide is first treated with 1-fluoro-2,4-dinitro benzene (Sanger's reagent), which selectively reacts with the N-terminal amino group. The peptide is then hydrolyzed to their amino acids and the N-terminal amino acid identified as its N-(2,4-dinitrophenyl) derivative (DNP).



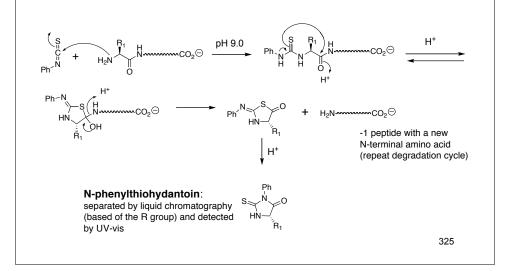
27.12: Insulin. (please read) Insulin has two peptide chains (the A chain has 21 amino acids and the B chain has 30 amino acids) held together by two disulfide linkages

Pepsin: cleaves at the C-terminal side of Phe, Tyr, Leu; but not at Val or Ala



27.13: The Edman Degradation and Automated Peptide

Sequencing. Chemical method for the sequential cleavage and identification of the amino acids of a peptide, one at a time starting from the N-terminus. *Reagent:* Ph-N=C=S, phenylisothiocyanate (PITC)



Peptide sequencing by Edman degradation:

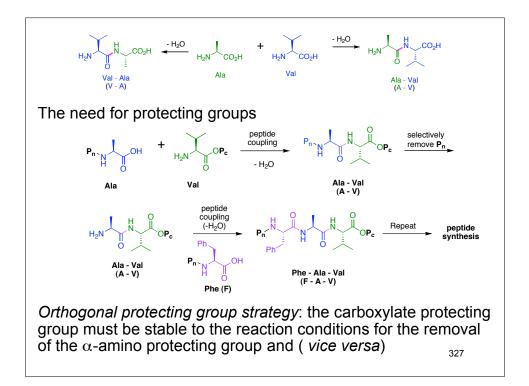
- Cycle the pH to control the cleavage of the N-terminal amino acid by PITC.
- Monitor the appearance of the new *N*-phenylthiohydantoin for each cycle.
- Good for peptides up to ~ 25 amino acids long.
- Longer peptides and proteins must be cut into smaller fragments before Edman sequencing.

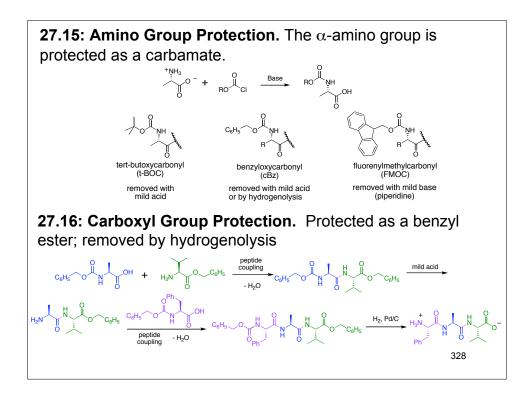
Tandem mass spectrometry has largely replaced Edman degradation for peptide sequencing

27.14: The Strategy for Peptide Synthesis:

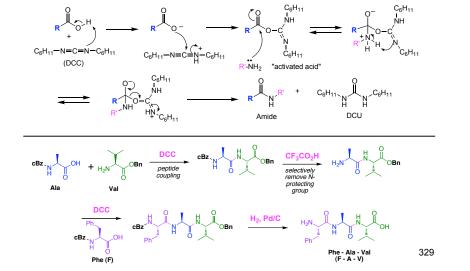
Chemical synthesis of peptide:

- 1. Solution phase synthesis
- 2. Solid-phase synthesis





27.17: Peptide Bond Formation. Amide formation from the reaction of an amine with a carboxylic acid is slow. Amide bond formation (peptide coupling) can be accelerated if the carboxylic acid is activated. *Reagent: dicyclohexylcarbodiimide (DCC)*



- In order to practically synthesize peptides and proteins, time consuming purifications steps must be avoided until the very end of the synthesis.
- Large excesses of reagents are used to drive reactions forward and accelerate the rate of reactions.
- How are the excess reagents and by-products from the reaction, which will interfere with subsequent coupling steps, removed without a purification step?

27.18: Solid-Phase Peptide Synthesis: The Merrifield Method. Peptides and proteins up to ~ 100 residues long are synthesized on a solid, insoluble, polymer support. Purification is conveniently accomplished after each step by a simple wash and filtration.

