



# aHUS: Facts, Controversies & Treatment Updates

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## DISCLOSURE STATEMENT

I, **Patrick Brophy** disclose the following relationships. Any real or apparent conflicts of interest related to the content of this presentation have been resolved.

<b>Affiliation / Financial Interest</b>	<b>Organization</b>
Advisor	Alexion – International Atypical Hemolytic Uremic Syndrome Advisory Board

The following presentation will not discuss unapproved or off-label, experimental or investigational use of medications.

# Objectives

- Review the diagnosis of aHUS
  - What its is
  - Genetics behind it
- Case presentations
  - Diagnostic dilemmas
- Treatment option
  - Old & New
- Homegrown perspectives



# aHUS

# FACTS



# aHUS?

**TTP**

**HIT**

**AMR**

**aHUS**

**STEC HUS**

**AKI**

**DIC**

**TMA**

**ITP**

**HELLP**

**MAHA**

**SIRS**

**AI**



# A Classification of TMA

(Thrombotic Microangiopathy)

<b>Typical / diarrheal</b>	<b>(HUS or TTP)</b>
<b>Complement defects</b>	<b>Atypical HUS</b>
<b>von Willebrand proteinase (ADAMSTS13) deficiency</b>	<b>Generally TTP</b>
<b>Cobalamin-C deficiency</b>	<b>TMA + multiorgan failure</b>
<b>Quinine-related</b>	<b>Abrupt TMA, exposure related</b>
<b>Post transplantation (calcineurin inhibitor related)</b>	<b>De-novo renal TMA May be renal "isolated"</b>
<b>Others: HIV, radiation, chemotherapy HELLP, antiphospholipid Ab syndrome, unclassified</b>	



# Typical HUS

**Triad of :**

**Microangiopathic hemolytic anemia**

**Thrombocytopenia**

**Acute kidney injury**

**Generally diarrhea-associated**

**Shiga toxin produced by *E coli* serotype O157:H7**

**Shigella, Salmonella, others also**

**Food borne disease: uncooked / unpasteurized  
products contaminated by animal wastes**

**Or other infections (respiratory):**

**Invasive *S. Pneumoniae* or viral infections**

# Typical HUS

**A severe condition:**

**acutely 2.5% mortality, often significant morbidity**

**Long term results (10-20 years after HUS\*)**

**63% Complete recovery**

**12% Recovery with proteinuria**

**6% Recovery with proteinuria and HTN**

**16% Recovery with low GFR  $\pm$  proteinuria or  
HTN**

**3% ESRD**

***\* Diarrheal or URI- related only, pediatric***





# Atypical HUS

## Clinically very severe

**15% died**

**25% ESRD**

**15% renal insufficiency**

**60% major sequelae**

**1/3 recover without significant renal disease**

**most (75%) of these had a single episode**

**few (25%) of these had recurrent aHUS**

***(a pediatric series)***



## Laboratory evaluation of suspected aHUS

<b>R/O STEC HUS</b>	Stool or rectal swab: culture for STEC; PCR for Stx Serum: Antibody testing
<b>R/O Streptococcus pneumoniae infection</b>	Blood cultures, pulmonary cultures
<b>R/O Thrombotic thrombocytopenia purpura</b>	Plasma ADAMTS-13 activity ± inhibitor
<b>R/O Other secondary causes of TMA</b>	Blood pressure, pancreatitis, malignancy, medication
<b>R/O Cobalamin metabolism abnormality (i.e. methyl-malonic aciduria)</b>	Homocysteine, Methionine, urine organic acid testing Consider genetic testing for mutation in MMACHC gene
<b>R/O Rheumatologic Disease</b>	Antinuclear antibody, lupus anticoagulant, anti-phospholipid antibodies
<b>R/O HIV</b>	HIV Serology
<b>R/O Pregnancy Associated (M) HELLP Syndrome</b>	Pregnancy test, liver enzymes.
<b>Rule in Complement Pathway Abnormality</b>	C3, Factor H, Factor I, Factor B serology Anti-factor H autoantibodies MCP - surface expression on leucocytes by FACS Gene mutation analysis for CFH, CFI, CFB, C3, CFHR5, MCP and CFHR1/3 Deletion

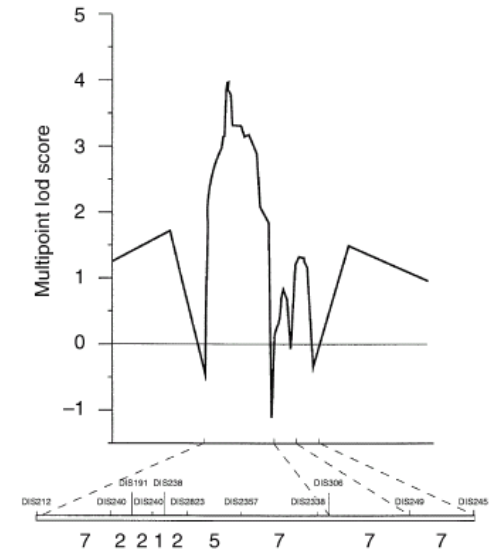
**Good Clinical History**

# Complement and Atypical HUS

Since the early 1970's alternative pathway complement activation (low C3), has been recognized in some cases of atypical HUS

1981: 1<sup>st</sup> case of HUS with factor H deficiency described

1998: Linkage analysis in 3 families with HUS provided clear association with *CFH*



# Atypical HUS

## COMPLEMENT DYSREGULATION

- complement components and pathways

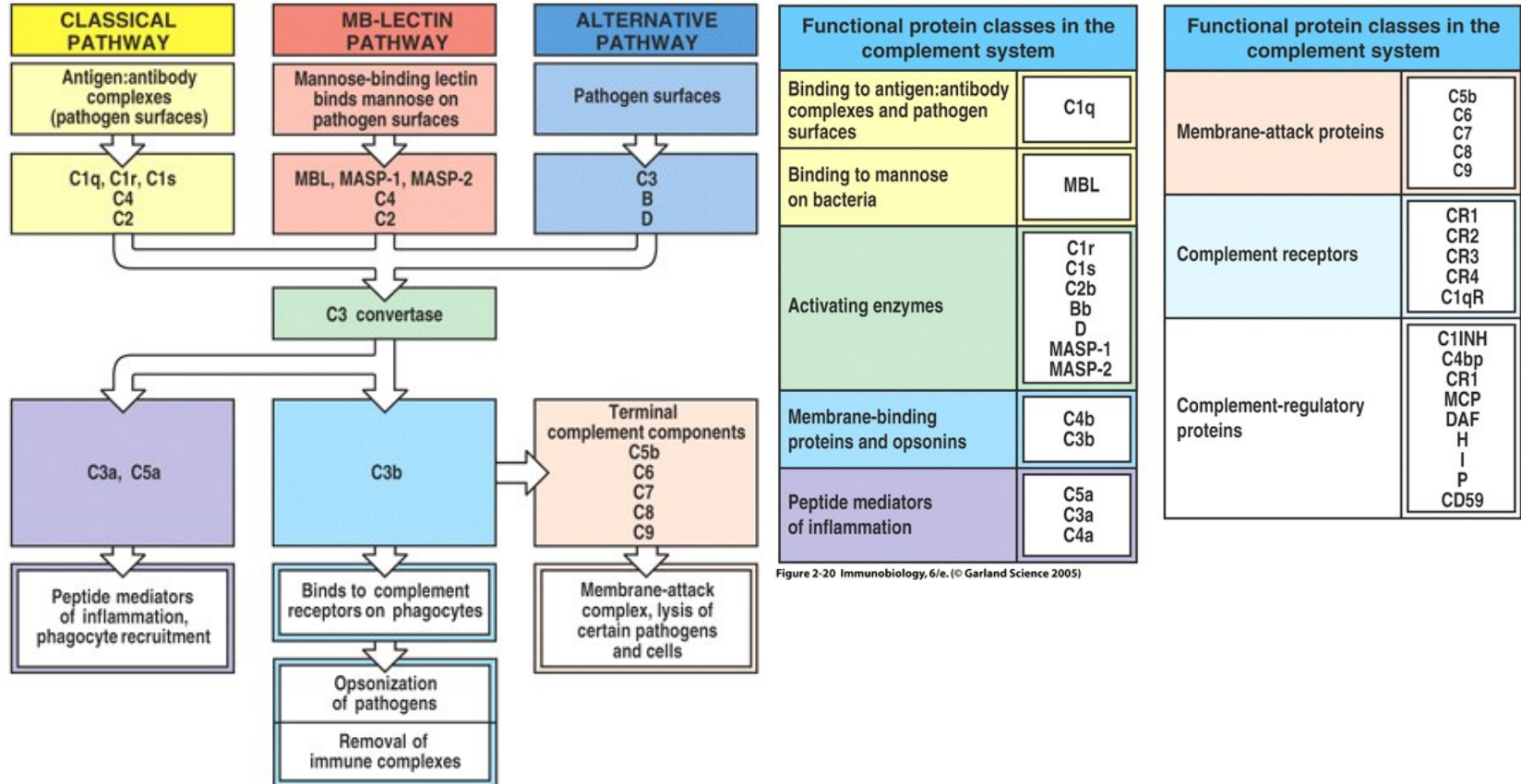
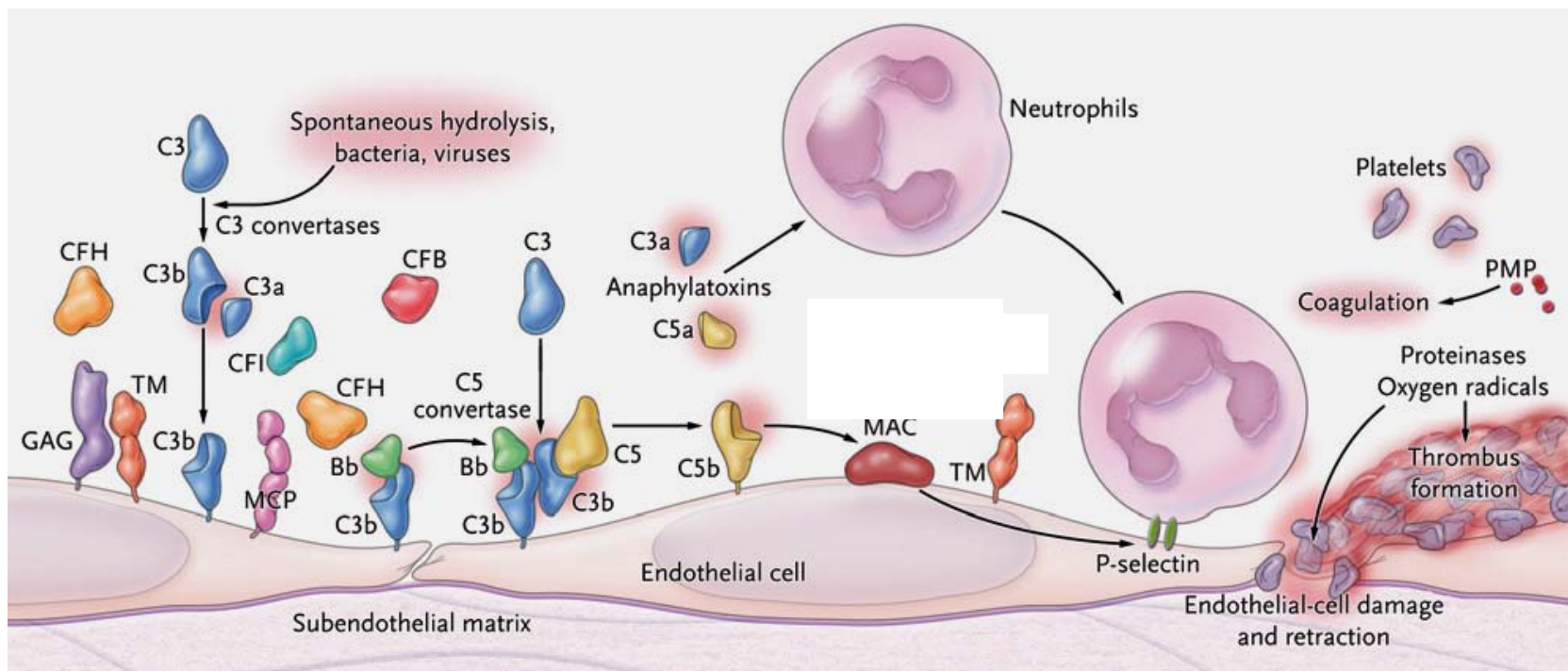


Figure 2-20 Immunobiology, 6/e. (© Garland Science 2005)

Figure 2-19 Immunobiology, 6/e. (© Garland Science 2005)



# The Alternate Complement Pathway

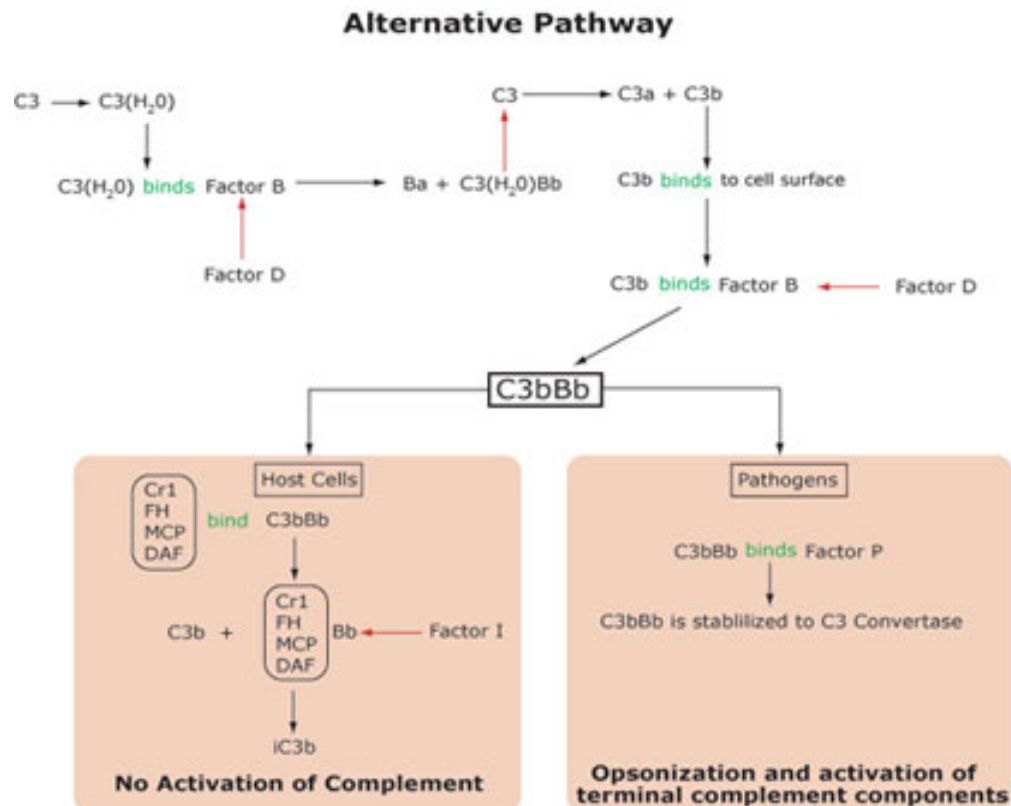




# Atypical HUS

## COMPLEMENT DYSREGULATION

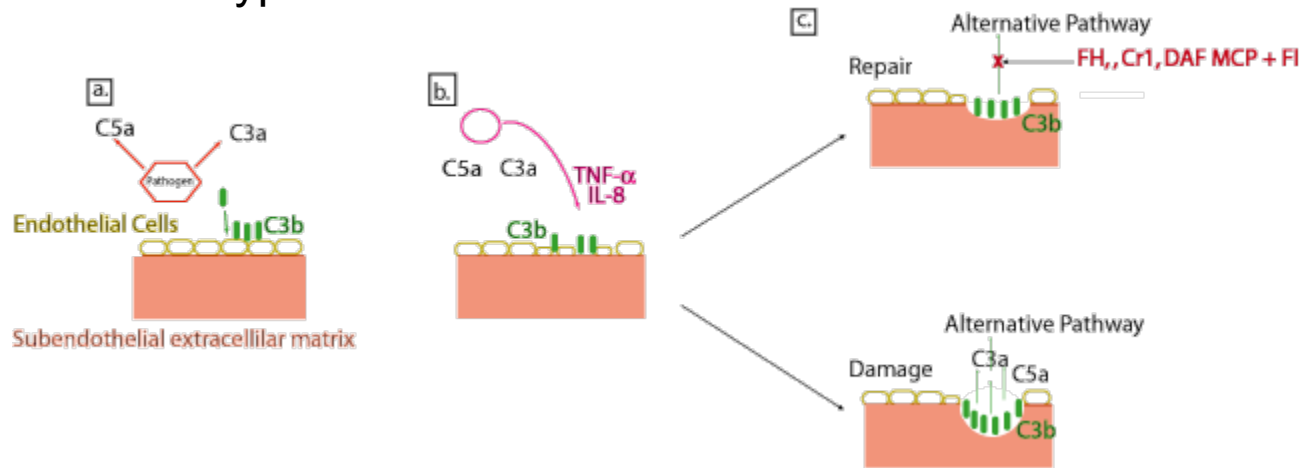
- complement regulation



# Atypical HUS

## COMPLEMENT DYSREGULATION

- pathogenesis of atypical HUS



- infection/inflammation increases rate of C3b formation
- activates complement cascade and C3a/C5a
- C3a/C5a attract leukocytes, producing TNF and IL-8
- cytokines cause endothelial damage and exposure of extracellular matrix
- ECM exposure amplifies deposition of C3b and complement activation
- lack of normal factor H, factor I, or MCP results in unchecked activation
- progressive tissue damage occurs

# Atypical HUS

## COMPLEMENT DYSREGULATION

- FH, FI, and MCP deficiency have incomplete penetrance
  - disease modifiers or other factors may have role
- environmental triggers
  - infections
    - preceded 70% of those with FH mutation
    - 60% of those with FI mutation
    - 100% of cases of HUS in MCP-mutants
  - pregnancy
    - trigger in 4% of FH-HUS
    - 40% of FI-HUS
- multiple-hits
  - one pedigree in which atypical HUS occurred only with inheritance of ALL:
    - MCP P131S mutation
    - MCP promoter polymorphism
    - dinucleotide insertion into FI gene
    - resulted in 50% expression level of each protein



# Atypical HUS

## COMPLEMENT DYSREGULATION

- outcomes of atypical HUS
  - overall 50% of patients develop ESRD
  - 25% mortality during acute illness
- end-stage renal disease
  - 70% with FH-deficiency HUS develop ESRD or die
  - >60% with FI-deficiency HUS develop ESRD
  - 86% with MCP-deficiency HUS remain dialysis-free
    - 70% had recurrence of HUS

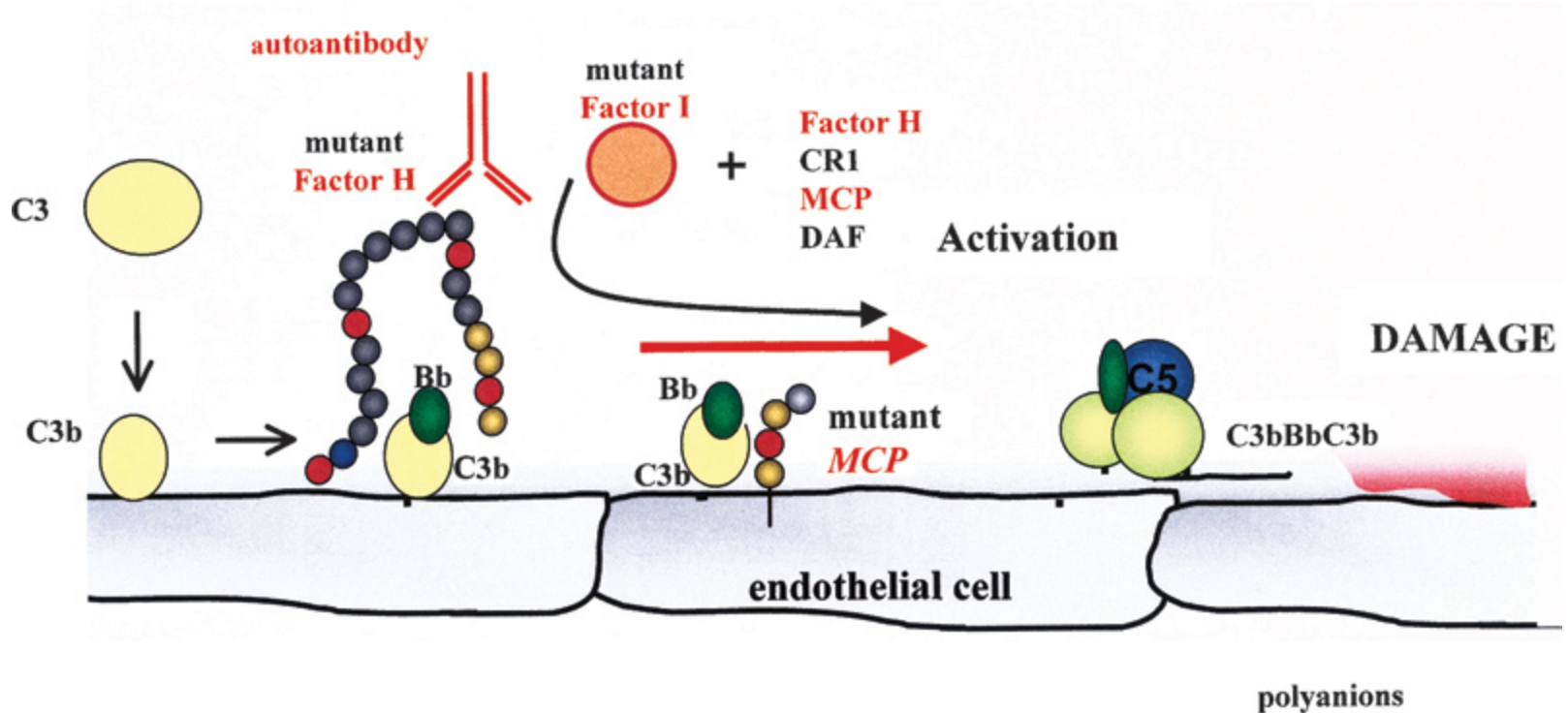


# SUMMARY

**Phase I**  
Activation

**Phase II**  
Amplification

**Phase III**  
Activation





# Genetic Abnormalities and Clinical Outcome in Patients with Atypical Hemolytic–Uremic Syndrome

Gene	Protein Affected	Main Effect	Frequency
<b><i>CFH</i></b>	Factor H	No binding to endothelium	20-30%
<b><i>CFHR1/3</i></b>	Factor HR1, R3	Anti-factor H Antibodies	6%
<b><i>MCP</i></b>	Membrane cofactor protein	No surface expression	10-15%
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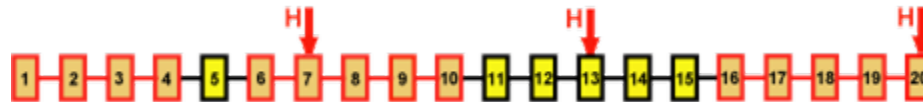
# Atypical HUS

## COMPLEMENT DYSREGULATION

- factor H, factor I, or MCP deficiency accounts for 50% of atypical HUS

## FACTOR H

- 150kD plasma glycoprotein synthesized in liver
- 20 homologous units of 61 residues (short consensus repeats – SCRs)



- N-terminal domains SCR1 – SCR4 bind C3b
  - complement decay accelerating activity located here
- H = three heparin binding sites
  - tertiary structure through to be bent backwards
  - exposes C-terminal SCR20
- functions as co-factor for factor I-mediated degradation of C3b,Bb

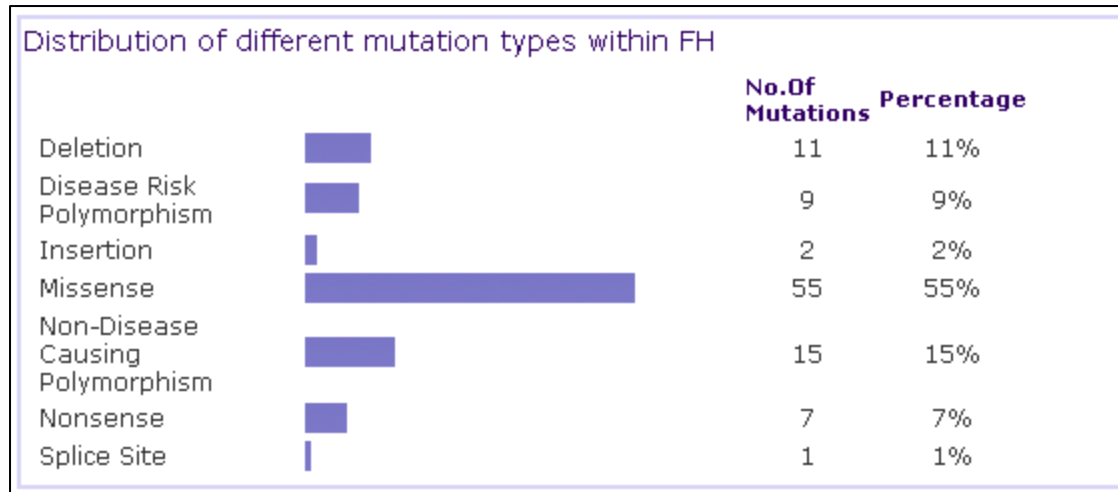


## FACTOR H DEFICIENCY

- thought to account for 10-22% of atypical HUS cases
- reported in both familial and sporadic forms
- usually presents in infancy or early childhood, but may present in adulthood
  
- one study of 16 FH-deficient patients
  - 6 with homozygous deficiency
    - 4 had membranoproliferative glomerulonephritis
    - 2 had atypical HUS
  - 10 had heterozygous deficiency
    - all developed atypical HUS
- homozygotes had low levels of FH, C3, FB and CH<sub>50</sub>
- heterozygotes had low to normal values
  
- some patients present with meningococcal infections
  - acquired C3 or terminal C' deficiencies
  
- some present with SLE, having combined FH and C2 deficiency

# Atypical HUS

## FACTOR H DEFICIENCY

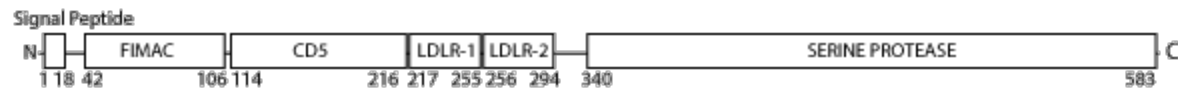


- 69 different FH mutations identified to date
- 3 patients have been described with atypical HUS and acquired anti-FH autoantibodies

# Atypical HUS

## FACTOR I

- 88kD plasma serine protease synthesized in liver



- N-terminal heavy chain
  - LDL-receptor domains x2
  - CD5 domain
  - FIMAC domain (factor I membrane attack complex)
- C-terminal catalytic domain
- functions to directly cleave C4b or C3b to inactivate complement
- efficient cleavage requires co-factors (C4bp, FH, MCP)

# Atypical HUS

## FACTOR I DEFICIENCY

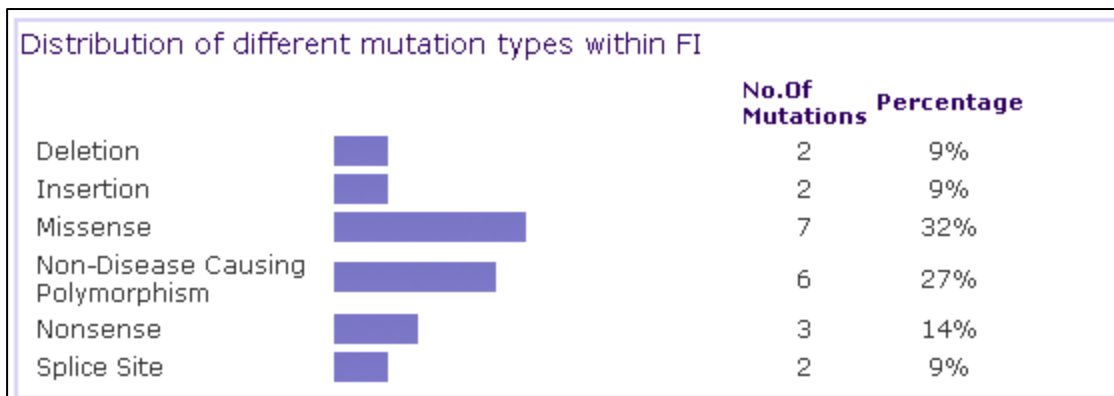
- reported only in sporadic forms of atypical HUS
- in one study, 2 out of 76 patients with atypical HUS had FI deficiency
  
- most reported cases involve heterozygous mutations
  - no increased susceptibility to infection
  
- homozygous FI deficiency associated with increased infection susceptibility
  - encapsulated organisms (meningococcus, pneumococcus, hemophilus)
  - acquired C3 deficiency due to uncontrolled consumption
  
- variable penetrance and expressivity
- C3 can be low to normal





# Atypical HUS

## FACTOR I DEFICIENCY



- 11 different FI mutations identified to date



# Atypical HUS

## MEMBRANE COFACTOR PROTEIN (MCP = CD46)

- ~65 kD transmembrane glycoprotein
- on leukocytes, platelets, endothelial & epithelial cells, fibroblasts, kidney



- extracellular domain
  - four SCR domains
  - alternative splice sites for O-glycosylation
  - multiple isoforms exist
- transmembrane domain
- cytoplasmic C-terminal anchor
  
- functions as cofactor for FI
- pathogen receptor for measles, adenovirus, HHV-6, Neisseria, and GAS

# Atypical HUS

## MCP DEFICIENCY

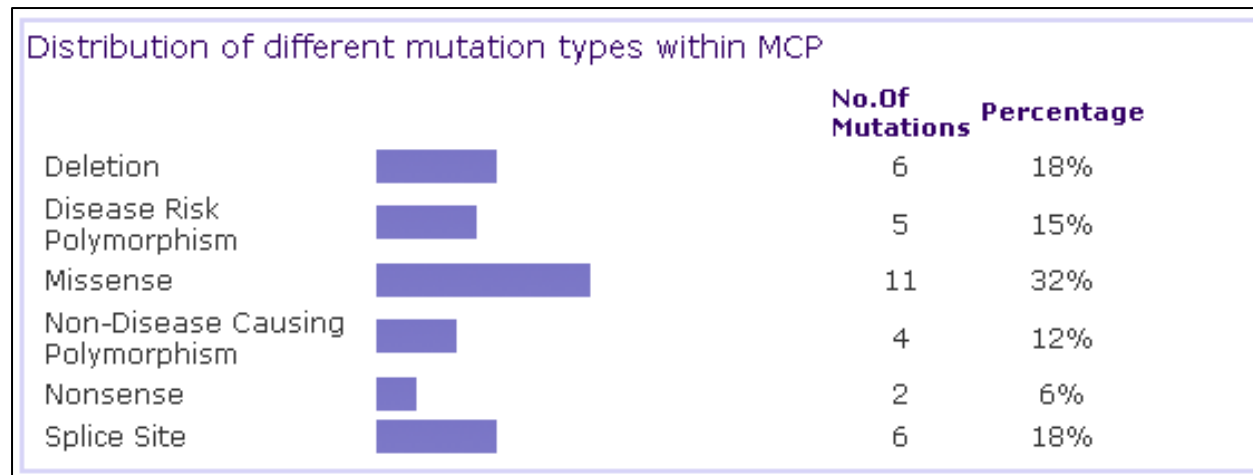
- reported only in familial forms of atypical HUS
- both homozygous and heterozygous types seen
- 80% of patients are heterozygotes

Patient	Gender	S/F	Age at Onset of Disease (yr)	Follow-Up (yr)	No. of Relapses	Renal Involvement (yr after the onset)	Comments
Homozygous							
1	F	S	27	4	2	CRF (CrCl 34 ml/min) (4)	Pierre-Robin syndrome, common variable immunodeficiency
2	M	S	5	35	10	HD (35)	
3	F	S	2	24	2	Proteinuria 2.66 g/24 h (23)	
Heterozygous							
4	M	S	10	9	1	HD (2)	Membranous glomerulopathy with proteinuria and HT at 6; transplant at 15; graft loss at 19
5	F	S	23	0.3	0	HD <sup>b</sup>	Onset 3 mo after second childbirth
6	F	S	0.7	11	0	HD <sup>b</sup>	Three transplants with early graft loss at 4, 6, and 10
7	F	S	5	8	4	None (8)	
8 <sup>c</sup>	F	F	3	18	4	HD (13)	Transplant at 17; lost at 21
9 <sup>c</sup>	F	F	5	13	6	CRF (ND)	Data on clinical progression are not available
Compound heterozygous							
10 <sup>c</sup>	M	F	1.5	3	4	None (3)	Initial presentation after <i>E. coli</i> O157:H7 infection
11 <sup>c</sup>	F	F	8	2	0	None (2)	



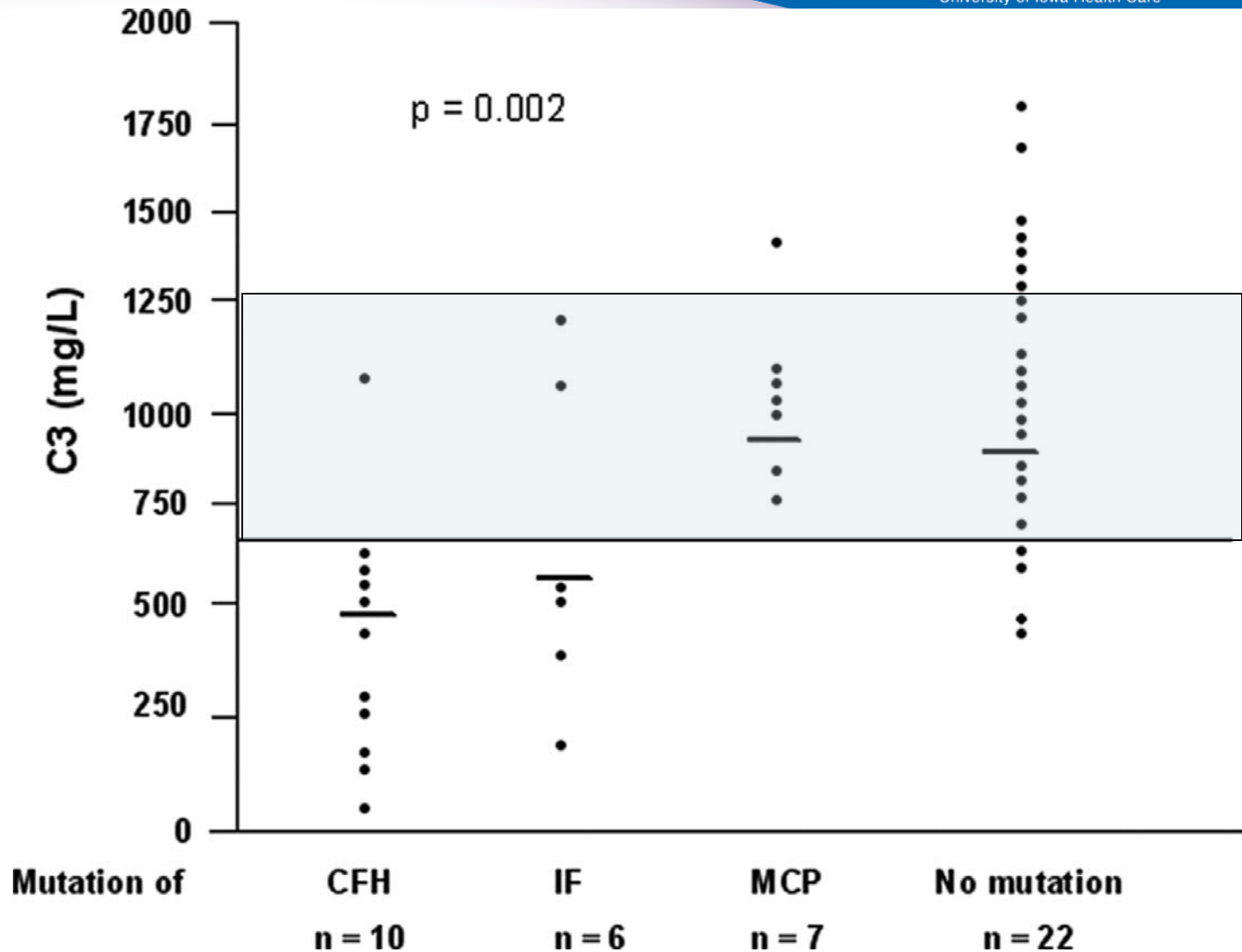
# Atypical HUS

## MCP DEFICIENCY



- 25 different MCP mutations identified to date

# C3 Levels By Mutation



# Complement and Atypical HUS

About 50%-60% of aHUS cases are associated with a mutation in a complement-related gene

Protein	Gene	Source	Location	% of aHUS
<b>Factor H</b>	<i>CFH</i>	Liver	circulates	~ 15-30%
<b>Factor I</b>	<i>CFI</i>	Liver	circulates	~ 5-10%
<b>Membrane Cofactor Protein</b>	<i>MCP</i>	Widespread	Membrane bound	~ 10-15%
<b>Factor B</b>	<i>CFB</i>	Liver, ?	circulates	<5%
<b>C3</b>	<i>C3</i>	Liver, ?	circulates	~ 5-10%
<b>Anti-FH-Ab</b>	<i>CFHR1/CFHR3</i>	Lymphocyte	circulates	~ 10%
<b>Unknown</b>				~ 40-50%



# **Recommended Initial Evaluation of HUS**

**Because infections trigger both typical and atypical HUS, initial evaluation should encompass both**

**Testing should include C3 level as well as classic evaluation (stool culture, LDH, smear, etc.)**

**ADAMSTS13 / auto-Ab analysis if TTP not ruled out**

**Save some plasma for later analysis**



# aHUS

# CONTROVERSIES





# Differentiating between aHUS, TTP & STEC HUS

- Thrombocytopenia
  - Platelet count  $<150,000$  Or  $>25\%$  Decrease from baseline
- Microangiopathic Hemolysis
  - Schistocytes *and/or* Elevated LDH *and/or* Decreased Haptoglobin *and/or* Decreased Hemoglobin
- Neurological Symptoms
  - Confusion/Seizures/Other abnormalities
- Renal Impairment
  - Increased creatinine/decreased GFR/Abnormal UA
- Gastrointestinal Symptoms
  - Diarrhea (+/-blood), N/V, gastroenteritis, Abdominal pain



# aHUS Cases



- 3 Cases- demonstrating the issues surrounding diagnosis and utilization of clinical criteria
- Case 1- 45 yr old woman: Initially diagnosed with TTP but accurate diagnosis is aHUS
- Case 2 – 10 yr old boy: Initially diagnosed with aHUS but accurate diagnosis was TTP
- Case 3 – 12 yr old girl undergoing kidney transplant as per UI protocol

Focus on clinical presentation and accurate diagnosis



# Case 1



- 45 yo Admit TMA management with atypical features (normal range platelet count ). History non-bloody diarrhea. Only Medication OCP.
- Initial Diagnosis TTP
- Initially presented with foot and ankle swelling and intermittent chest tightness. Lower extremity edema since one month prior (found to have a creatinine of 3.0 and an elevated blood pressure)



# Case 1



- Systemic symptoms progressed to nausea, vomiting, occasional low-grade fever and chills. Several days later followup noted to have a creatinine of 7.7, hemoglobin 7.2, and platelet count 227.
- Biopsy found thrombotic microangiopathy> Complement not done
- **Differential DIAGNOSIS: TTP vs. aHUS**
  - ADAMTS13 Activity levels at 58% (Post PEX) (N >67%). APL normal. ANA reported as abnormal. Hep screen negative. Haptoglobin low. Shistocytes 1-2+
  - Limitation of ADAMTS13 measured post PEX

**its critical to utilize ADAMTS13 activity to differentiate between aHUS and TTP**



- **Six PEX Treatments:** Fluid replacement: 500 mL normal saline, 500 mL of 5% albumin, 3000 mL of plasma
- PEX initiated because of **TTP diagnosis**
- **Continued Symptoms-** developed irreversible renal failure now on **Chronic Hemodialysis**
- **Subsequent follow up: complement levels obtained and...**

# Case 1 Labs

	HgB	Platelets	Hapto	LDH	C3
Day					
1	8	240	20	475	
2	8.4	258			
3	8.1	224	22	435	
30					55
40					58
50					68

PEX initiated at day 1.... ADAMTS level obtained after 1<sup>st</sup> treatment Level 58% (N >67%)



# Genetic Abnormalities and Clinical Outcome in Patients with Atypical Hemolytic–Uremic Syndrome: Need to assess Complement in patients with TMA

Gene	Protein Affected	Main Effect	Frequency	Response to Plasma Therapy
<b>CFH</b>	Factor H	No binding to endothelium	20-30%	60% Remission
<b>CFHR1/3</b>	Factor HR1, R3	Anti-factor H Antibodies	6%	70-80% Remission
<b>MCP</b>	Membrane cofactor protein	No surface expression	10-15%	No definitive indication for Therapy
<b>CFI</b>	Factor I	Low level or low cofactor activity	4-10%	30-40% Remission
<b>CFB</b>	Factor B	C3 Convertase stabilization	1-2%	30% Remission
<b>C3</b>	Complement C3	Resistance to C3b inactivation	5-10%	40-50% Remission
<b>THBD</b>	Thrombomodulin	Reduced C3b inactivation	5%	60% Remission





## Case 2



- 10 yr old male- mild fever, nausea, vomiting and some confusion (negative past history)
- HgB 7 mg/dl, plt 100, LDH 4715, Haptoglobin low, 8% schistocytes, Acute kidney injury creat- 1.4 mg/dl
- Patient referred as an aHUS patient and ADAMTS13 activity level used to differentiate from TTP
- Complement studies- initially normal
- ADAMTS 13 activity level sent-given the variable presentation of aHUS





# Table 1.

Table 1. Course of ADAMTS13 activity and anti-ADAMTS13 immunoglobulin G titer

Time After Admission	Day 1	Month 1	Month 2	Month 3	Month 6	Month 12
ADAMTS13 activity, %	<5	<5	41	60	140	150
Anti-ADAMTS13 immunoglobulin G titer, IU/mL	84	22	13	12	neg	neg

neg, negative.

Table 1. Course of ADAMTS13 activity and anti-ADAMTS13 immunoglobulin G titer

**Successful treatment with rituximab for acute refractory thrombotic thrombocytopenic purpura related to acquired ADAMTS13 deficiency: A pediatric report and literature review.**

Harambat, Jerome; Lamireau, Delphine; Delmas, Yahsou; Ryman, Anne; Llanas, Brigitte; Brissaud, Olivier; MD, PhD

Pediatric Critical Care Medicine. 12(2):e90-e93, March 2011.

DOI:

10.1097/PCC.0b013e3181e89f8f

## ADAMTS13 to differentiate between aHUS and TTP



## Case 2

- PEX Initiated with FFP replacement– no initial response of hematological parameters
- Condition worsened and patient developed TAMOF
- Based on ADAMTS 13 levels (<5%) and presence of strong inhibitory antibody patient started on immunosuppression
- Good Response- clinical parameters began to improve: platelets, LDH etc



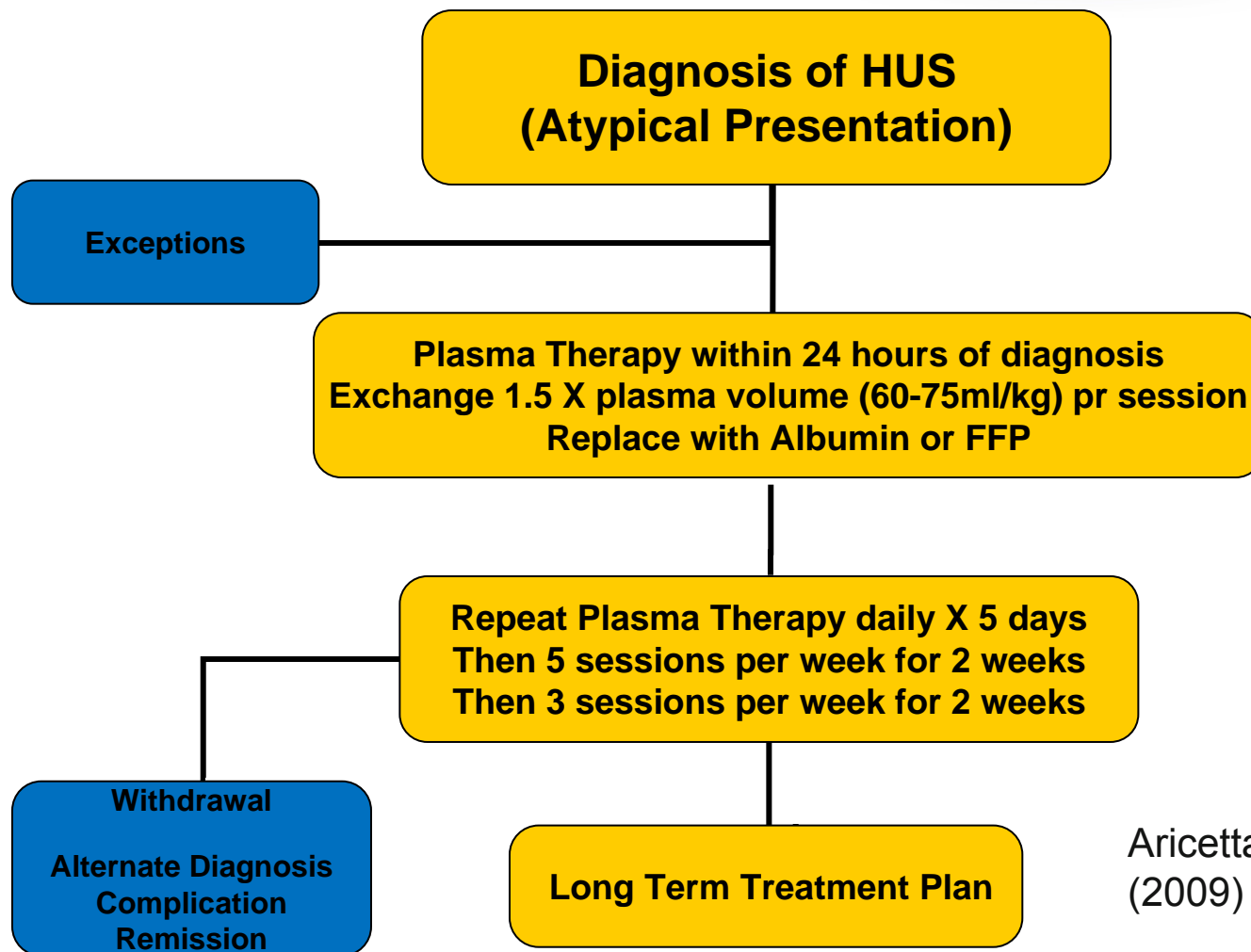
# Case 3

- Teenage patient- second transplant, initial diagnosis missed
- 28 Plasma Exchanges
  - Platelet 144,000 mm<sup>3</sup> to 337,000 mm<sup>3</sup> after thirteen Exchanges over 5 weeks
  - Hgb 5.5g/dl to 11.1 g/dl
  - Renal Function did not recover
    - Started on Chronic HD.....what else?





# Plasma Therapy- standard approach



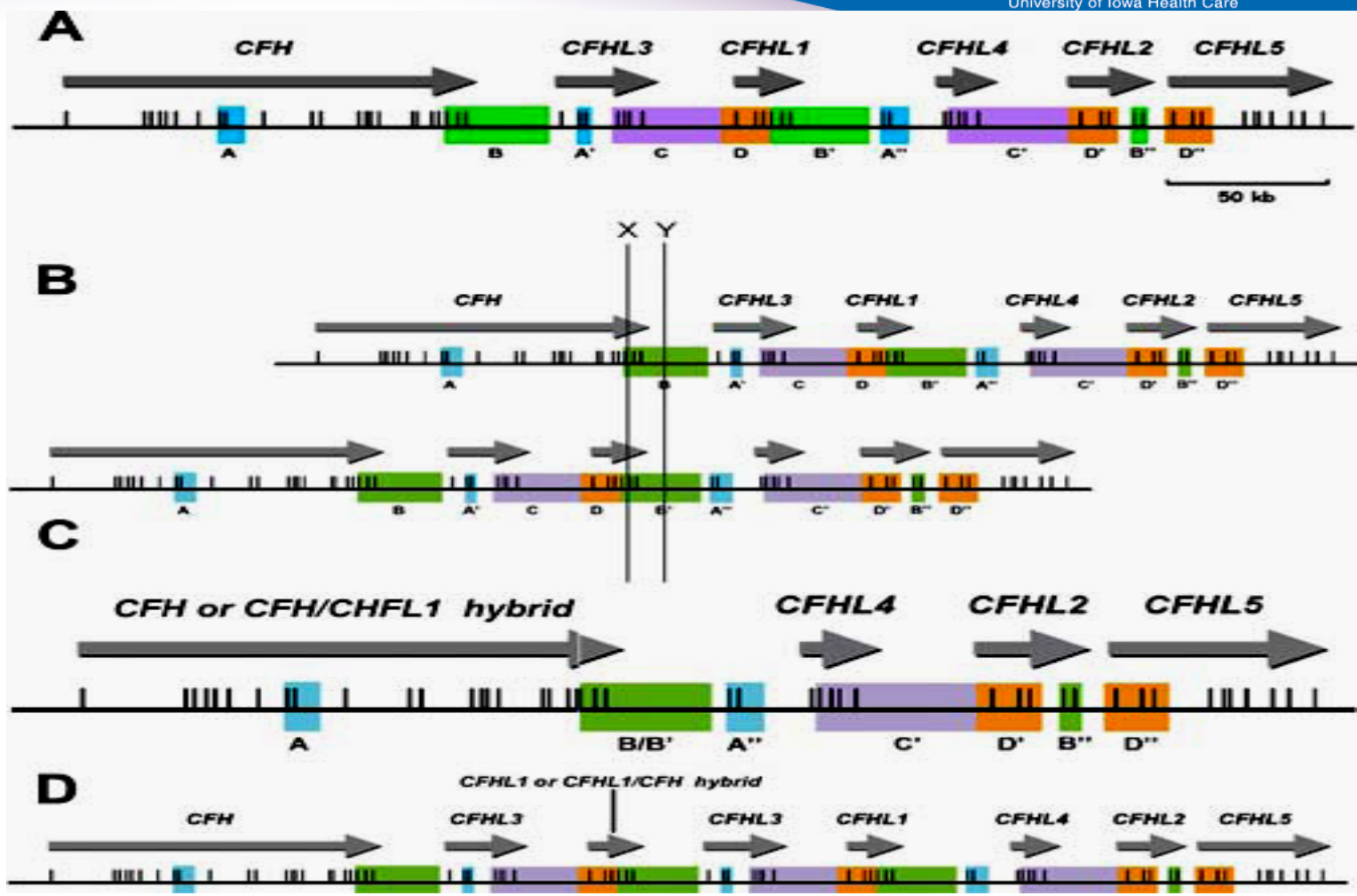
Aricetta, Ped Neph,  
(2009) 24:687–696



# Case 3

- No mutations in *CFH*, *CFI*, *CFHR5*, *MCP*, *CFB*, *C3* or *THBD*
- only 30-50% aHUS patients have an identifiable mutation
- Copy Number studies (MPLA)
  - Hybrid *CFH/CFHR1* gene
    - Fusion protein comprised of the first 18 short consensus repeats (SCRs) of *CFH* and the last two SCRs of *CFHR1*

To be continued.....





## Genetic Abnormalities and Clinical Outcome in Patients with Atypical Hemolytic–Uremic Syndrome

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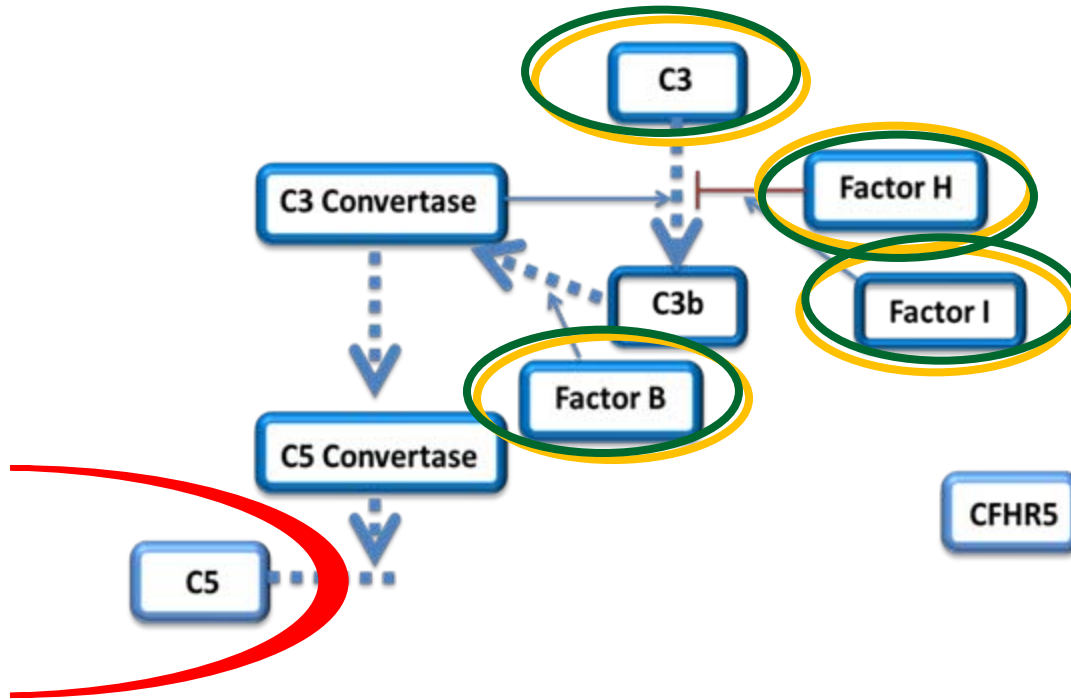
# aHUS

# TREATMENT



## The Genetics of Atypical Hemolytic Uremic Syndrome

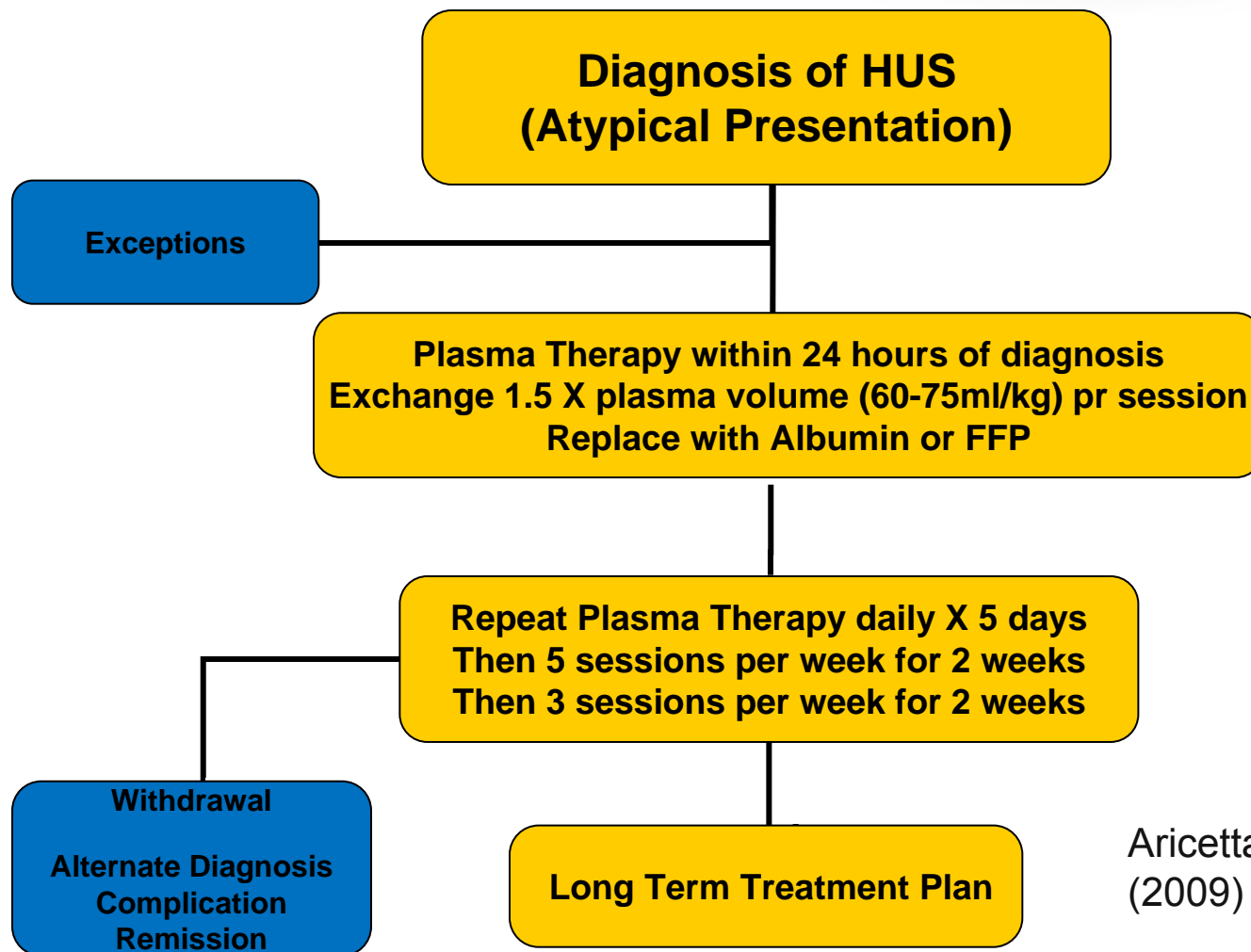
Gene	Role of Mutation	European Cohort <sup>1</sup>			US Cohort <sup>2</sup>
		Frequency	Risk of ESRD at 3 years	Risk of transplant loss within 1 year	Frequency
<b>CFH</b>	Mutation results in a quantitative deficiency of protein or altered binding to C3b*	23%	77%	71%	27%
<b>MCP</b>	Mutant proteins have low C3b binding capacity and therefore decreased cofactor activity	7%	6%	0%	5%
<b>CFI</b>	Mutations induce a default of secretion of the protein or disrupt its cofactor activity altering degradation of C3b/C4b	4%	60%	67%	8%
<b>C3</b>	Mutations interfere with binding of C3 to MCP and regulation by MCP or increased binding to CFB resulting in increased C3 convertase formation.	8%	67%	43%	2%
<b>CFB</b>	Mutated proteins binds excessively to C3b and stabilizing the C3 convertase making it resistant to decay by CFH, enhancing formation of C5b-9 complexes and deposition of C3-fragments onto endothelial cell surfaces	1%	-	-	4%
<b>THBD</b>	Mutated proteins are less effective at moderating CFI-mediated inactivation of C3b	5%	54%	0%	3%
<b>CFHR1/3</b>	Associated with CFH Antibodies	6%	-		
<b>CFHR5</b>	Unknown	Not Reported			3%
<b>Fusion Proteins</b>	Results in non-functional CFH	Not Reported			Not Reported
<b>CFH Antibody</b>	Anti-CFH IgG bind to CFH and inhibit CFH binding to C3b and cell surfaces	3%	63%		Not Reported
<b>Unknown</b>		52%	50%		54%



1. Remove abnormal proteins
2. Replace deficient proteins
3. Block terminal complement effects.



# Plasma Therapy- standard approach



Aricetta, Ped Neph,  
(2009) 24:687–696



# Plasmatherapy

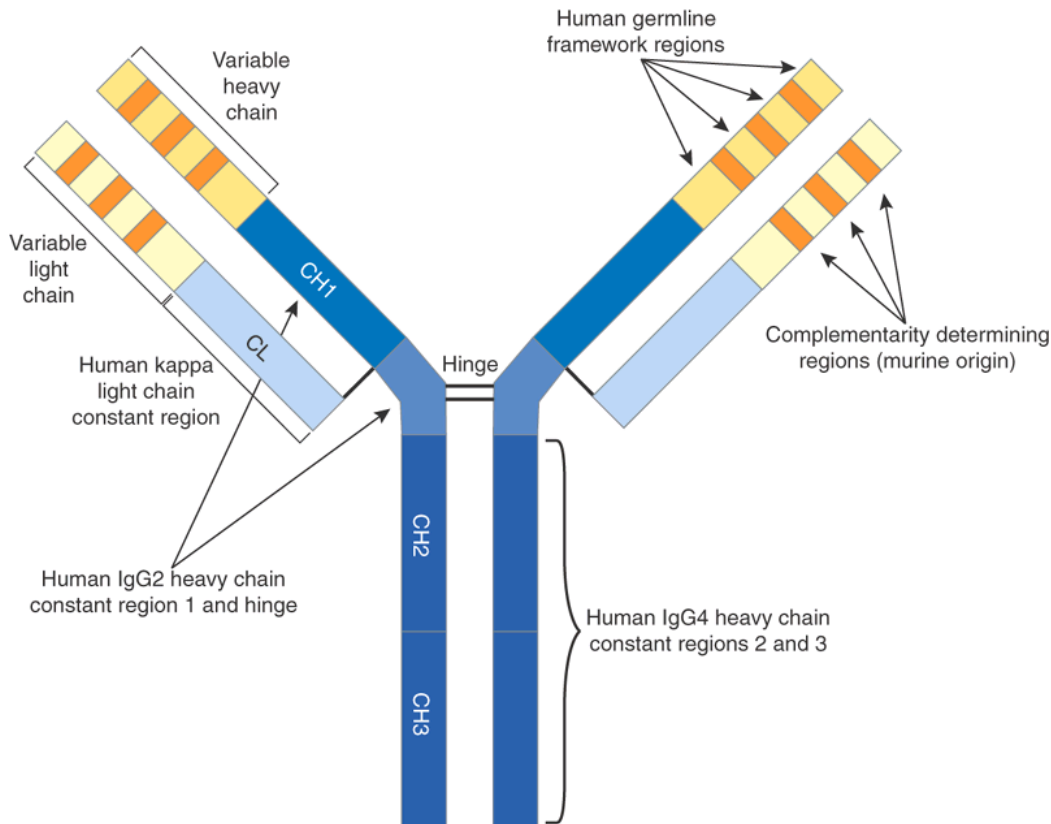
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CFH Mutation	37%	Death or ESRD
CFI	75%	Death or ESRD
C3	43%	Death or ESRD
Thrombomodulin	13%	Death or ESRD



# Eculizumab



- Recombinant, humanized, monoclonal antibody directed against C5 – specifically preventing its cleavage by the C5 convertase.
- Prevents the generation of the terminal complement complex C5b-9.
- The single most expensive drug in the world. (\$409,500/yr – *Forbes Magazine*)



# Eculizumab and aHUS

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# Terminal Complement Blockade

Eculizumab and Atypical Hemolytic Uremic Syndrome		
<b>Patients Resistant to Plasma Therapy - 26 Week Results</b>		
Change in Platelet Count	+ 96 ± 21 X 10 <sup>9</sup>	P = <.0001
TMA Event Free Status	15/17 patients (88%)	95% CI 64-100
Change in Renal Function		
One stage improvement	11/17 patients (65%)	
Less than one stage	4/17 patients (23%)	
Removal from dialysis	5/7 patients (71%)	
Quality of Life Improvement	EuroQol 5D: 0.33 ± 0.09	P = <0001
<b>Patients on Chronic Plasma Therapy - 12 week Results</b>		
TMA Event Free Status	13/15 patients (87%)	95% CI 60-98
TMA Interventions	0/17 patients (0%)	
<b>Pediatric Patients Exposed to Eculizumab - Retrospective Review of 4 Week Data</b>		
Hematologic Parameters	Normalization in 8/19 patients (42%)	
Change in Renal Function	≥ 15ml/min/1.73m <sup>2</sup> improvement in 7/19 patients (78%)	
Removal from dialysis	4/8 patients (50%)	

**FDA Approval September 2011**



# Liver-Kidney Transplant- Option

## 1

- Goal: correct genetic defect
  - Liver protein replacement strategy
- Initial attempts - high mortality
  - Early failure of transplant liver
- 7/8 with extensive PTx have retained their liver
  - 8th died of hepatic encephalopathy





# Liver Transplant

- CFH, CFI, CFB and C3
- 20 Cases (Presse Med. 2012; 41: e115–e135)
- 4 procedures in 2002 – All Fatal
- With preconditioning - 86% patient survival reported
- Protection against kidney transplant rejection
- No reports of aHUS recurrence



**Table 6. Guidelines to surgery and perioperative treatment of patients receiving a combined liver and kidney transplant or a liver transplant alone<sup>a</sup>**

Dialysis<sup>b</sup>

better before plasma exchange in all cases

mandatory before plasma exchange in cases with evidence of complement activation (e.g., angioedema) during dialysis

Plasma exchange<sup>b</sup>

a minimum of 1.5 Vol of FFP is exchanged within 4 to 6 h of surgery

exchange must be repeated if surgery is delayed

Plasma infusion

10 to 20 ml/kg body wt FFP is infused intraoperatively after native hepatic explant

additional plasma may be given as clinical need dictates

Surgery

split or whole liver transplantation is indicated

adequate liver mass must be provided (minimum 2% liver to recipient mass ratio)

auxiliary liver transplantation is not recommended

living-related donation is not recommended

Monitoring

posttransplantation liver function should be judged by coagulation profile

in cases of inadequate liver function, plasma exchange in conjunction with standard care is indicated<sup>c</sup>

Posttransplantation anticoagulation<sup>d</sup>

low molecular weight heparin at prophylactic dosages (e.g., enoxaparin 0.5 mg/kg twice daily)

aspirin (2 mg/kg per d up to 80 mg/d)

to be continued for 3 mo

Immunosuppression

per standard practice of each center

mTOR inhibitors are not encouraged

**Eculizumab**



# Liver Transplant

- PEX with FFP 4-6 hours before transfer to OR
- Eculizumab
- Intraoperative transplant immune suppression
- Hepatectomy → Whole-liver orthotopic transplant
- Intra-operative FFP after liver perfusion
- Kidney transplant
- Eculizumab
- Heparin/ASA



# Impediments to Care

- Consideration of the diagnosis
- Availability of diagnostic studies
- Decision to PEX or Ecu acutely may be based on logistics
  - Local Expertise
  - Formulary concerns
  - Hospital financial concerns

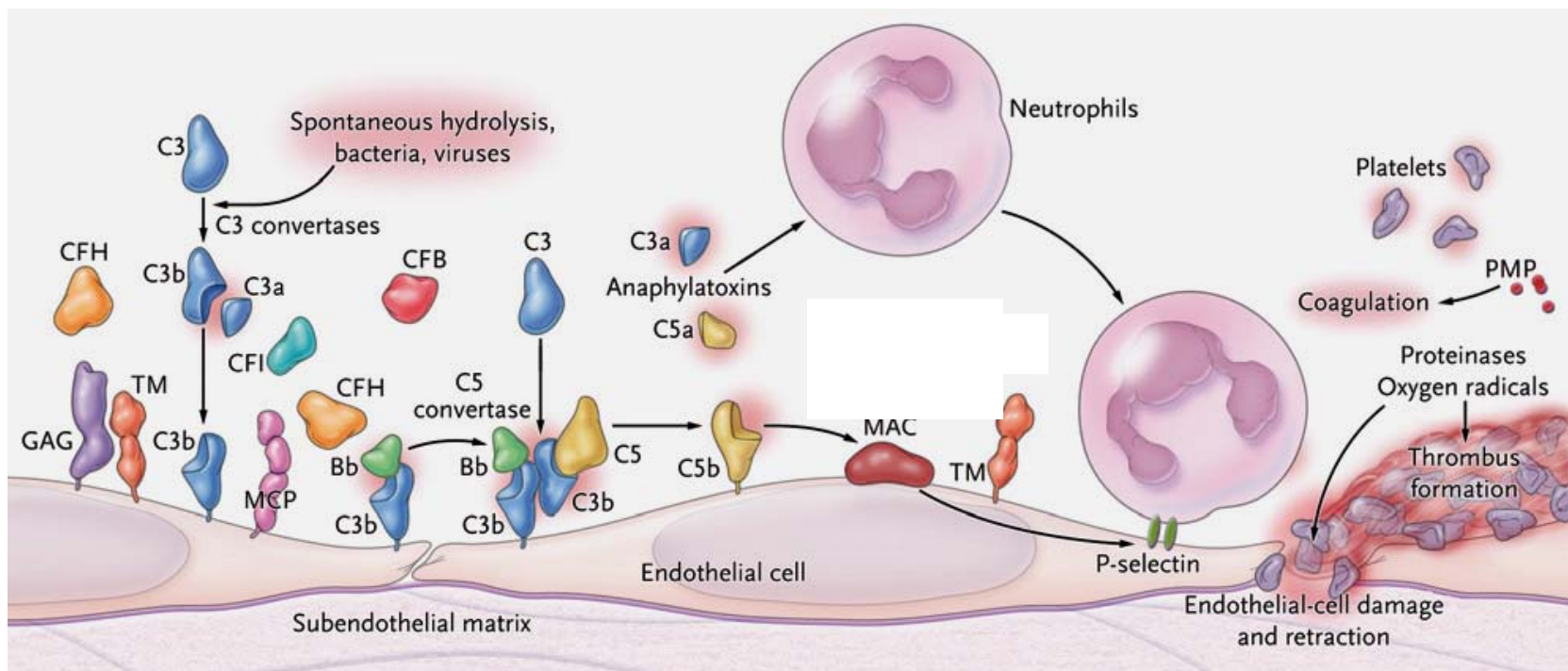


# New Kidney + Plasma Therapy & Eculizumab

## Option 2

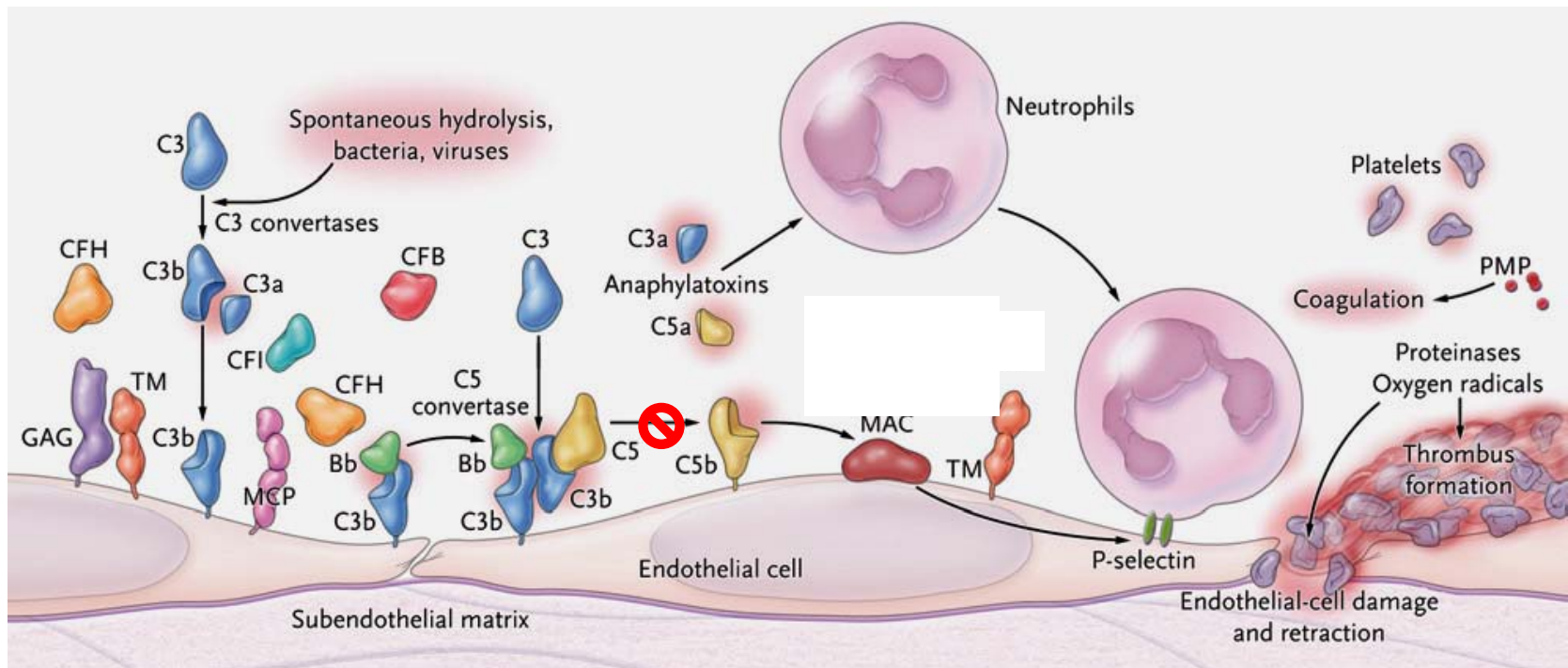


# The Alternate Complement Pathway





# The Alternate Complement Pathway





# Case 3

- On dialysis for 15 months
- Altruistic donor comes forward
- Rare Renal Disease Team approval
- Renal Transplant Protocol Approved  
– Eculizumab and Plasma Therapy
- Transplant 10/07/10

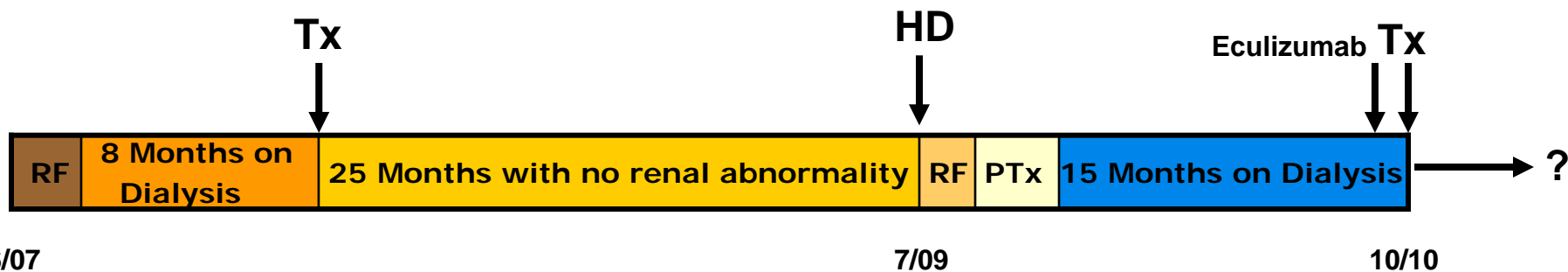






# Concerns

- Genetic Testing
- Efficacy
- Life time risk of infection
- Surveillance
- Length of Therapy
- Payment
- On going research





# Patient Update

- Creatinine 0.9, 16 months s/p transplant
- No bacterial infections
- Returned to school full time
- 1/13 H&H 12.2/36 → 3/10 6.3/18
  - WBC, platelets normal
  - Borderline low C3
  - What else do you want to know?



- Hemolytic Workup Negative
- Bone Marrow Biopsy
  - Mildly hypocellular bone marrow (50-70%) showing normal granulopoiesis, normal megakaryocytes and markedly decreased erythropoiesis with maturation arrest and giant normoblasts with viral inclusions
  - Now what do you want to know?



- Parvovirus B19 found in bone marrow
- Cellcept discontinued
- 1 Unit PRBC given 3/10
- 2<sup>nd</sup> Unit given 3/24



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## Pre-transplant Evaluation

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- Donor evaluation for pathogenic aHUS mutations
- Immunize against meningococcus (as well as hemophilus and pneumococci if not current)
- Verify titer if on dialysis or if immune suppressed at the time of vaccination – consider re-immunization and antibacterial prophylaxis as necessary.

### Zero minus 1 week (prior to transplant)

- Measure C3\*, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate C3Nef, CFI and/or CFI levels.  
*Labs must be drawn before plasma therapy and before eculizumab*
- Plasma exchange or Infusion – **1st dose** - (1.5 volumes of Albumin)
- Administer 900 mg eculizumab – **1st dose**

### Zero minus (24-48 hours prior to transplant)

- Measure C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate C3Nef, CFI and/or CFI levels.  
*Labs must be drawn before plasma therapy and before eculizumab*
- Plasma exchange or infusion – **2<sup>nd</sup> dose** – (1.5 volumes of FFP)

### Zero hour (0-24 hours before transplant)

- Administer 900mg eculizumab – **2<sup>nd</sup> dose**

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## Post-transplant

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- Measure C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate C3Nef, CFI and/or CFI levels.  
*Labs must be drawn before plasma therapy and before eculizumab*
- Administer 900mg eculizumab q7 days times two (**3rd and 4<sup>th</sup> dose**)
- Administer 900mg eculizumab q 14 days



# Kidney Transplant

Univ of Iowa OTC ver 1.4 Jan 2012

## Protocol for the prophylactic use of eculizumab for renal transplant in a patient with atypical HUS (1)

### Pre-transplant Preparation

#### Donor

- Unrelated Living Donor: (Preferred option). Usual pre-donation transplant workup.
- Related Living Donor: (Less desirable option [2]). Donor should be evaluated for pathogenic aHUS mutations<sup>†</sup>.
- Deceased Donor: (Less desirable option). Usual donor workup. Will require adjustment of the recipient protocol based on type of genetic mutation in recipient, anticipated cold ischemic time, prior history of transplant and current PRA.

#### Recipient

- Immunize recipient against meningococcus – meningococcal conjugate vaccine (if 2 through 55 years of age) - 2 doses 2 months apart (immunize against hemophilus and pneumococci if not currently immunized).
- Verify titer 1 month after 2<sup>nd</sup> dose if on dialysis or if immune suppressed at the time of vaccination – consider re-immunization and antibacterial prophylaxis as necessary.
- If unable to immunize at least two weeks in advance of first dose of eculizumab, must use anti-meningococcal antibiotic coverage for the first 14 days post immunization.

#### Zero minus 4-7 days (prior to transplant)(for living donor recipients only)

- Measure C3<sup>\*\*</sup>, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate CFH, CFI and/or autoantibody levels (C3Nef, FHAA, FBAA)
- One red top and one pink top for immediate isolation of serum and plasma for storage<sup>®</sup>  
*Labs must be drawn before eculizumab*
- Administer 900 mg eculizumab– 1<sup>st</sup> dose
- One red top and one pink top for immediate isolation of serum and plasma for storage<sup>®</sup>  
*Labs must be drawn 60 minutes after eculizumab*

#### Zero minus 1 day (prior to transplant) (for all recipients)

- Measure C3<sup>\*\*</sup>, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate CFH, CFI, CFB and/or autoantibody levels (C3Nef, FHAA, FBAA)
- One red top and one pink top for immediate isolation of serum and plasma for storage<sup>®</sup>  
*Labs must be drawn before plasma therapy and before eculizumab*
- Plasmapheresis (PE) – 1st dose<sup>§</sup> - (1 volume of FFP)
- Administer 1200 mg eculizumab after plasma exchange– 2<sup>nd</sup> dose for LD recipients (1<sup>st</sup> dose for DD recipients)

#### Zero hour (immediately prior to OR - day of transplant for all recipients)

- Measure PT, PTT, C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate CFB, CFI and/or autoantibody levels (C3Nef, FHAA, FBAA)
- One red top and one pink top for immediate isolation of serum and plasma for storage<sup>®</sup>  
*Labs must be drawn 60 minutes after eculizumab*

### Post-transplant

#### Post-Op Day Zero (upon discharge from OR)

- Measure C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate CFB, CFI and/or autoantibody levels (C3Nef, FHAA, FBAA)
- One red top and one pink top for immediate isolation of serum and plasma for storage<sup>®</sup>
- Administer 1200 mg eculizumab for DD recipients only (2<sup>nd</sup> dose for DD recipients)

#### Daily

- Monitor Hb, platelets, renal function, haptoglobin and LDH

#### Day 7, Day 14, and Day 21

- Measure C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet

count and if appropriate CFH, CFI and/or autoantibody levels (C3Nef, FHAA, FBAA)  
*Labs must be drawn before eculizumab*

- Administer 900-1200 mg eculizumab on day 7, 14, and 21 (3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup> dose)

#### Day 35 Postop

- Measure C3, C4, AH50, CH50, C5 Functional Level, CBC, LDH, haptoglobin, platelet count and if appropriate CFH, CFI and/or autoantibody levels (C3Nef, FHAA, FBAA)  
*Labs must be drawn before eculizumab*
- Administer 900-1200 mg eculizumab q 14 days starting with the 6<sup>th</sup> dose.

The following measures to be considered if ongoing hemolysis is noted: Plasmapheresis with FFP or albumin. Additional doses of eculizumab may be necessary if apheresis is performed, or if breakthrough hemolysis occurs

#### Notes:

<sup>†</sup> Mutation screening of *CFH*, *MCP (CD46)*, *CFI*, *CFB*, *C3*, *THBD* and *MLPA* for copy number analysis of *CFHR1* and *CFHR3* performed at the Molecular Otolaryngology & Renal Research Laboratory, Iowa City

<sup>®</sup>Contact Amy Weaver (335-6623), or Richard Smith ([Richard-smith@uiowa.edu](mailto:Richard-smith@uiowa.edu)), MORL, Iowa City.

<sup>§</sup>Consider additional plasmapheresis pre-transplant if autoantibody present or if CFH mutation is expected to result in a gain of function or dominant negative effect.

#### Eculizumab - drug Interactions.

A number of therapeutic antibodies are used in the management of kidney transplant recipients. Antibodies, such as rituximab [3] and alemtuzumab[4], that exert its action primarily by complement dependent cytotoxicity (CDC) may be inhibited by concurrent administration of eculizumab. Thymoglobulin depletes peripheral T cells by a combination of CDC, antibody dependent cellular cytotoxicity (ADCC) and activation induced apoptosis [5]. If thymoglobulin is used with eculizumab monitor its efficacy by measurement of absolute lymphocyte or CD3 count. Basiliximab competitively inhibits IL-2 binding to activated T lymphocytes and does not require complement for its action [6].

#### \*\*Abbreviations:

C3, complement component 3; C4, complement component 4; AH50, alternate complement pathway hemolytic assay; CH50, total complement pathway hemolytic assay; C5, complement component 5; CBC, complete blood count; C3Nef, complement component 3 nephritic factor; CFI, complement factor I level; CFH, complement factor H level; FHAA, factor H autoantibody; FBAA, factor B autoantibody; q, every

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# Molecular Otolaryngology & Renal Research Laboratory

- University of Iowa
  - Dr. Richard Smith
- CLIA Testing
- Research Testing – Clinician/Patient driven
  - IRB Mechanism

<http://www.healthcare.uiowa.edu/labs/morl/>



# MORL

(Molecular Otolaryngology & Renal Research Laboratory)

- Not for profit Lab
- Diagnostics section
  - CLIA certified
  - aHUS testing
  - screen all KNOWN genes for ANY possible variant.
  - will detect novel mutations and previously reported mutations
- Discovery section
  - Genetics of rare renal diseases including atypical HUS and Dense Deposit Disease (one section)
  - New assays in development





# Specialized Testing

- Adam TS 13 mutations-  
ELISA
  - TTP analysis
- sMAC testing
  - only lab in US- some with DDD and all with C3 GN-
- Only ~50% DDD patients successful in transplantation those seem to have sMAC (eculizumab)



# aHUS

- 260 cases processed and diagnosed in lab
- Large genetics repository- goal to develop more fully a complete registry
- Other tests – recent German cases: may have complement abnormalities underlying as well—EHEC may help ID those with underlying genetic predisposition

# Characterization of an American aHUS Cohort

**Table 1:** Clinical outcomes of aHUS patients (n=70)

Clinical Outcome	Patients
Renal Failure	57* (81%)
No Relapse	13 (19%)
Renal Transplant	13 (19%)
Transplant Failures	7 (54% of all transplants)

\*4 patients deceased

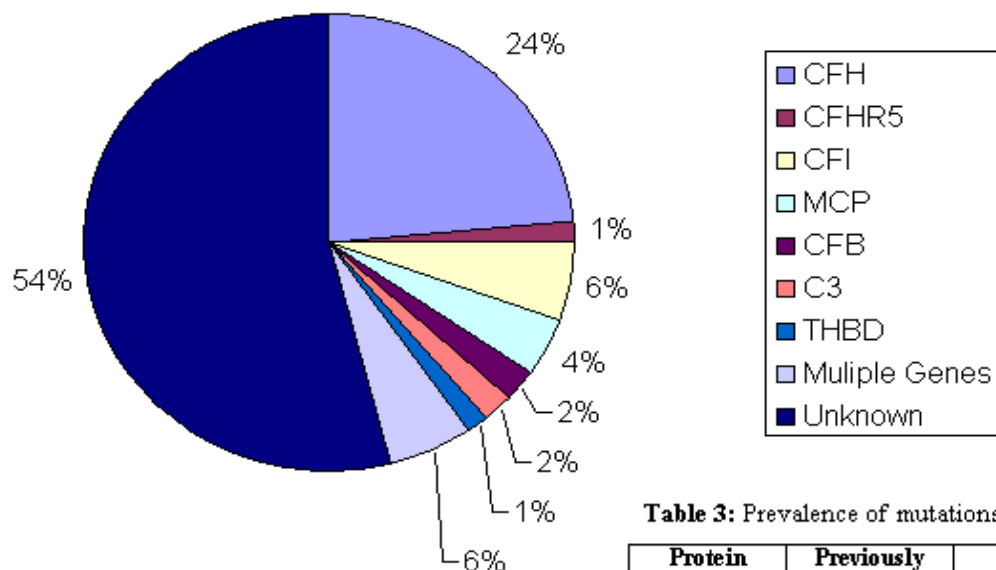


**Table 2:** Clinical outcome for each mutated protein

Mutated Protein	Renal Failure	Failed Transplant*	No Relapse	Unknown Clinical Outcome	Total
CFH	16	4	2	16	34
CFHR5	-	-	1	1	2
CFI	4	-	-	4	8
MCP	1	-	1	4	6
CFB	2	-	-	1	3
C3	-	-	-	3	3
THBD	1	-	-	1	2
CFH/CFI/MCP	1	-	-	-	1
CFH/CFHR5	-	-	-	1	1
CFH/CFB	1	-	-	1	2
CFH/CFI	1	-	-	-	1
CFI/CFB	1	1	-	-	1
THBD/CFHR5	-	-	-	1	1
THBD/CFI/CFI	1	-	-	-	1
No Mutations	28	2	9	41	78
<b>Total</b>	<b>57</b>	<b>7</b>	<b>13</b>	<b>74</b>	<b>144</b>

\*These patients are included in the number of patients in renal failure (Column 2)

# Prevalence of Mutations in American Cohort

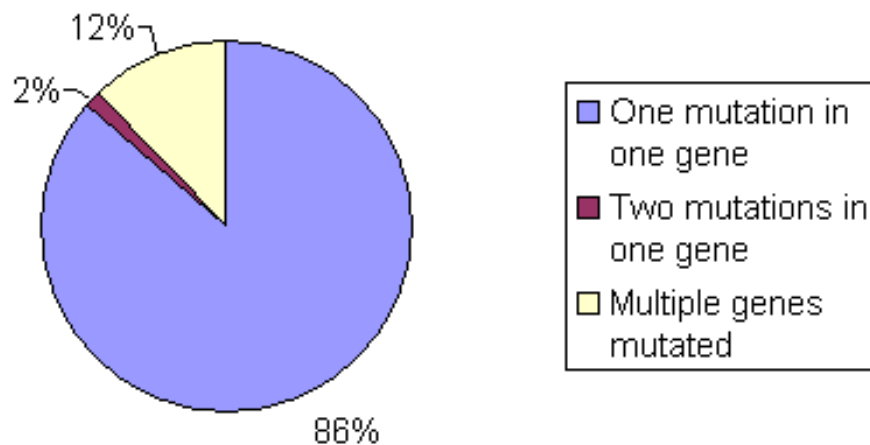


**Table 3:** Prevalence of mutations found in American aHUS cohort (n=144)

Protein	Previously Reported Mutations	Novel Mutations	Total Mutations	Number of Patients	Percentage in Cohort	Reported Frequencies
CFH	9	9	18	39	27%	20-30%
CFHR5	0	3	3	4	3%	-
CFI	2	7	9	12	8%	4-10%
MCP	2	5	7	7	5%	10-15%
CFB	0	6	6	6	4%	1-2%
C3	1	2	3	3	2%	5-10%
THBD	2	2	4	4	3%	5%
<b>Total</b>	16	34	50	66*	46%	-
<b>Unknown</b>	-	-	-	78	54%	-

\*Correction for patients with either a mutation in more than one gene or multiple mutations in a single gene

# American aHUS Patients Carrying Multiple Complement Gene Variants



Category	Number	Patient	Genes
Patients who have more than one mutation in the same gene	1	1679	<i>CFI</i> (p.G119R and p.G287R)
Patients who have mutations in more than one gene	8*	aHUS-12	<i>CFH, CFI, MCP</i>
		770	<i>CFH, CFB</i>
		1964	<i>CFH, CFB</i>
		1939	<i>CFH, CFI</i>
		aHUS-07	<i>CFI, CFB</i>
		1679	<i>CFI, THBD</i>
		511	<i>CFH/CFHR5</i>
		1525	<i>CFHR5/THBD</i>



# Summary of Results

- Percentage of Mutations found in American patients
  - Similar
    - Overall mutations found ~50%
    - CFH Mutations ~30%
    - CFI ~10%
    - THBD ~5%
  - Increased
    - CFB ~4% compared to ~2% (doubled)
  - Decreased
    - CD46 ~5% compared to ~10% - Likely due to the level of severity in our cohort (half)
    - C3 ~2% compared to ~10% (one quarter)
- Important to screen all known susceptibility genes
  - 12% of patients that were mutation positive carried more than one mutation
- Likely that rare variants are important in disease
- Thrombomodulin- unsure of the significance of the variants at this point
- Ethnic controls are important



# University of Iowa Rare Renal Disease Clinic

- Research: Dr Smith and his research team
- <http://www.healthcare.uiowa.edu/labs/morl/>
- Clinical- held in the Pediatric Specialty Clinics
- Dr. Richard Smith
- Dr. Carla Nester
- Dr. Christie Thomas
- Dr. Alan Reed
- Dr. Pat Brophy
- Monica-  
keleher@uiowa.edu



# Thank You

- Carla Nester
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- Gary Pien
- Brad Dixon
- Rare Renal team  
University of Iowa