# Applications of Flow Cytometry in Veterinary Medicine

Melinda J. Wilkerson, DVM, PhD Diplomate, ACVP Kansas State University College of Veterinary Medicine



## Overview

- 1. Instrumentation
  - A. Essential components
  - B. Basic principles of light scatter and fluorescence
- 2. Sample preparation/processing
- 3. Application of flow cytometry
  - A. Detection of surface antibody
  - B. Lineage determination of malignant neoplasms
  - C. Define an antibody panel
  - D. Identifying clonality (PARR test)
- 4. Case examples



# Flow cytometers

- "flow cyto meter" = measurement of cells in a fluid stream
  - Measure multiple characteristics of cells by light scatter and fluorescence using lasers
  - Quantify leukocytes and differentiate cell types
  - Fluorescent dye labeled antibodies to cluster of differentiation antigens identify subsets of cells
    - i.e. CD4 or CD8 lymphocytes
      - immunodeficiency syndromes
      - lineage of lymphoma and leukemia
    - DNA content, apoptosis, viability, metabolism, proliferation



## First Flow Cytometer, an "old tool"

#### 4. Electronics



K-STATE

#### **Basic Components**

- 1. Laser excitation optics
- 2. Fluidics
- **3. Collection Optics**
- **4. Electronics**

Marvin Van Dilla, Trujillo TT, Mullianey PF, Coulter JR (1969). Cell microfluorimetry: A method for rapid fluorescence measurement. Science 163:1213-1214.

## Analytical Flow Cytometer FACSCalibur ©BD





# Components of a bench top analytical flow cytometer





## Fluorescence colors

- First fluorochromes or fluorophores
  - Fluorescein (FITC)
  - Seaweeds/cyanobacteria
    - Phycobiliproteins (PE)
    - Allophycocyanin (APC)
  - Jelly fish Green
     fluorescent protein (GFP)
  - Propidium iodide (PI)





Aequoria victoria



## **Fluidics**

- Pressure lines and sheath fluid introduce sample stream into laser beam
- Sample pressure is higher than sheath fluid and introduces cells in single file

K-STATE



### Basic light scatter properties of cells





# Differential light scatter properties of cells define individual cells





Hematology Analyzers Advia 2120i use laser light scatter to differentiate cell populations





RBCS/platelets Lymphocytes Monocytes Neutrophils Eosinophils



# **Collection Optics**

#### Collection Optics

- Mirrors, Filters, Detectors
- Collect and filter wavelengths of light that come from the particle-laser beam interaction
  - -Light scatter
  - -Fluorescence



## Basic principles of fluorescence



#### **Fluorescence**

Laser excites fluorescently labeled antibodies to CD molecules



### **Collection Optics & Detectors**



K - STATE America. Hematology.Vol. 42. 2012.



## Large granular lymphocytosis



Heeb, Wilkerson, Chun. JAAHA. 2003:39:379-384



# When should immunophenotyping be ordered?

- 2006 Consensus recommendations to use FC IP to screen for malignancy in people:
  - Bicytopenia and pancytopenia
  - Elevated leukocyte concentration
  - Presence of atypical cells or blasts in blood, bone marrow, or body fluids
  - Plasmacytosis or monoclonal gammopathy
  - Organomegaly and tissue masses



## Key Role in Diagnosis and Classification

- Mature lymphoid neoplasms
- Plasma cell neoplasms
- Blastic malignancies
- Maturing myeloid and monocytic malignancies
- Monitoring treatment of a previously diagnosed hematolymphoid neoplasia
- Detection of minimal residual disease
  - 0.1 to 0.01% abnormal cells



## Immunophenotyping in Veterinary Medicine

- Immunophenotyping using monoclonal antibodies to cluster differentiation (CD) molecules better defines the lineage of hemic neoplasia than morphologic assessment
  - B lymphomas respond more readily to therapy
  - Leukemias can be differentiated into
    - Myeloid vs. Lymphocytic
    - Acute leukemias identified by CD34 expression
    - Chronic lymphocytic leukemia in dogs is principally a CD8+ cell lineage (CD4+ in cats)
      - W. Vernau and P. Moore, 1999. Vet. Immun. Immunopathol.



### Cell lineage differentiation defined by Cluster of Differentiation molecules



## Selecting an Antibody Panel

<b>CD</b> molecule	T cell	B cell	Others
CD45			Pan leukocyte
CD34			Stem cell
CD3 or CD5	T cells		
CD4	T helper		
CD8	T cytotoxic		
CD79a		Pro B cell to	
(cytoplasmic)		Plasma cell	
slgM, CD22		Immat./Mat. B	
CD21		Mature B	
CD14			Monocytes
CD11b			Granulocytes
CD11c/CD1a			Dendritic cells



# Sample preparation and processing

- Blood or bone marrow in EDTA anticoagulated collection tubes
- Tissue samples (lymph nodes)
  - Needle biopsy of multiple areas
  - Best to have 500,000 to 1 million cells/mL
  - Add sample to a tube of 1 2 mL of saline
  - Send overnight carrier on cool packs



## Scatter Plot of blood leukocytes







**Possible Myelomonocytic** 

K-STATE



### Guidelines for lymphoid malignancy

- In blood, lymphocyte concentration above reference range for lab and one of the following
  - 80% of lymphocytes with a single phenotype or
  - 60% of lymphocytes with a single phenotype and a positive clonality assay (PARR) or
  - Presence of lymphocytes with an aberrant phenotype for peripheral blood
- In tissue
  - Expansion of homogenous population of single phenotype
  - Presence of aberrant phenotype



# Immunophenotyping of Fluids

- Cerebral spinal fluid
- Dog with ataxia
- Epidural lesion at T4
- Total nucleated cell count = 3,175/μL
- Protein = 145 mg/dL
- Cytology
  - Pleocytocysis
  - Monocytic or lymphocytic?





# Diagnosis of CSF case

- Imunophenotyping panel – CD45 (pan leukocyte)
  - CD3+ (T cell marker)
  - CD21 (mature B cell)
  - CD14 (monocyte)
- Necropsy
  - Multicentric lymphoma
  - Lymph nodes and Liver
  - Invasion of spinal nerves

99% cells 87% cells 3% cells 18% cells



#### Adult Dog with mediastinal mass

Cells from needle aspirate labeled with fluorescent antibodies to CD4 and CD8

- Thymic lymphoma?
- B cell lymphoma?
- Thymoma?





CD4+CD8+



Is this lesion polyclonal or monoclonal? What would You expect with PARR? See Lana et al., 2006 JVIM and Lara-Garcia et al., 2008 VCP.

# Antibody specificity

- Dot plot analysis
  - Complexity vs. size
    - Place gates on cells based on size
    - Green = Small cells
    - Pink = Large cells
  - Size vs.
     Fluorescence

K-STATE

 Cells stained for CD markers are detected to the right of the vertical line

#### **Normal Lymph Node**



## Lymphoma cases

- 5 yr old Labrador
- Large lymphoblastic cells
   CD3 negative
   CD21+ IgM+



#### **B cell lineage**





**Fluorescence** 

**Fluorescence** 

# Case 2

**K-STATE** 

#### 9 year old mix breed dog •Large & small cells •CD3+ CD4+ •Few small cells (resident) •CD8+ •CD21+CD79a+



#### CD4 CD3 Size SC-H 103 10<sup>2</sup> CD4 FITC 103 10 10<sup>2</sup> CD3 FITC **CD21** CD8 Size 10<sup>2</sup> CD21 FITC 103 101 10<sup>2</sup> CD8B FITC 10 103 800 100 800 100 Control CD79a 600 H-: Size F 00 103 100 104 101 10<sup>2</sup> FL2-H 10 10 10

**Fluorescence** 

## Case 3 7 year old Pointer

- Large cells
  - CD34+ CD21+ IgM+
     CD79a wk+
- Small cells
  - CD3+

K-STATE

- Significance of CD34?
- Which cells are neoplastic?



# Significance of CD34 expression in human lymphoma

 Schmidt et al., Aberrant Antigen Expression Detected by Multiparameter Three Color Flow Cytometry in Intermediate and High Grade B-Cell Lymphomas. 1999. Leukemia and Lymphoma

15% of the B cell lymphomas were CD34+



#### Immunophenotypic markers of prognosis in canine Iymphoproliferative disorders

- Study at CSU of 96 dogs with lymphocytosis including stage V lymphoma. Williams et. al. 2008. JVIM
  - CD34+ phenotype in the blood had shorter survival time (average of 16 days) compared to lymphocytosis that expressed mature B and T cell antigens (300 – 500 days)
  - CD21+ lymphocytosis composed of <u>large</u> cells had shorter survival times compared to small cell CD21+ lymphocytosis
- Rao, et. al., JVIM. 2011
  - Lack of MHC II expression in B cell lymphomas had a poor outcome (Rao, et al., JVIM. 2011.)
- Avery et. Al. Abstract, ACVP Proceedings 2012
  - T cell lymphomas not expressing CD5 had a better prognosis



## Case 4: 2-year old male Golden retriever with peripheral lymphadenopathy

#### **Laboratory Data Abnormalities**

Hypercalcemia [19.4 mg/dL] Thrombocytopenia 36,000/μL Lymphopenia 800/ μL Ref. Range [9.7-12.1 mg/dL] Ref. Range [164,000-10,000/μL] Ref. Range [1,500-5,000/μL]



## Aspirate of prescapular lymph node



90% of cells are immature lymphocytes

## **Bone Marrow Aspirate**



Immature Iymphocytes replace normal bone marrow

## High power image of Bone Marrow



Diagnosis = Stage V lymphoma Additional Tests: Immunophenotyping using flow cytometry to determine cell lineage

> Majority of lymphocytes in lymph node aspirate express T cell antigens (CD5, CD3, CD4) Cell lineage = ?



## Lymphoma vs reactive lymph node?

- Lymphoma = neoplasia of lymphocytes
  - Clonal expansion of T or B cells
    - Single receptor specificity
    - Immature morphology
- Reactive lymph node
  - Antigenic stimulation causes expansion of multiple clones of T and B cells (polyclonal)
    - Multiple receptor specificities
    - Mature morphology and presence of plasma cells



## **Reactive lymph node**



# How do you distinguish reactive lymph node enlargement from lymphoma?



 Grossly bilateral
 lymphadenopathy suggests neoplasia



# PCR for antigen receptor (PARR) rearrangements to identify clonality







### Important information on the PARR test

- 1. Sensitivity & Specificity: 75% & 92%
- Detects 1 neoplastic lymphocytes in 100 heterogeneous nonneoplastic cells
- Detects neoplastic lymphocytes in PB
   2.5 times more than microscopic evaluation
- 4. Not prognostic for disease-free interval or duration of survival
- 5. Confirmed B-cell lymphosarcoma in aqueous humor sample
- False negatives primers not specific, somatic recombination, NK cells
- 7. False positive -pseudoclonality (amount and quality of DNA)

- Burnett et al., 2003 Vet. Path.
- Keller et al., 2004. Vet Clin. Path.
- Lana et al., 2006. JVIM
- Pate et al., 2011.
   JAVMA
- Werner et al., 2005 Vet. Path.

### Case 5 Mixture of B and T cells Large B lymphocytes are clonal



T cell Rich B cell Lymphoma



## Case 6

7 yr Labrador cross

- Large & small cells
   CD3+, CD34+
- Small cells

   CD4+, CD8+ bright
- Large cells

- CD21+/CD3+, CD79a+



K-STATE

C IgH Igh TCR $\gamma$ 

B cell lineage



# Case 7: Aberrant T cell and monocyte antigen expression in a B cell clone



K-STATE

B cell clonality

## Reports of aberrant CD expression and gene rearrangements

- Gelain et al., 2008. Aberrant phenotypes and quantitative antigen expression in different subtypes of canine lymphoma by flow cytometry.
  - B decreased CD79 and expression of CD34
  - T decreased CD45 expression
  - CD3 or CD5 expression without CD4 or CD8
  - CD79 and not CD21 for B cells
- Wilkerson et al., 2005. Lineage differentiation of canine lymphoma/leukemias and aberrant expression of CD molecules.
  - CD8, CD14, and CD21
- Kyoda et . al. 1997. Prognostic significance of immunoglobulin heavy chain gene rearrangement in patients with acute myelogenous leukemia.



## Conclusions

- 1. Flow cytometry Immunophenotyping useful tool in diagnosis/prognosis of canine lymphoproliferative and hematopoietic neoplasias
  - Providing the sample can be dispersed in suspension
  - Correlates with cytomorphologic features
- 2. Broad immunophenotyping panels and multi-color analysis improves diagnostic capabilities.
- 3. Lineage infidelity is common at early stages of hematopoietic differentiation. Aberrant expression of CD molecules occurs in canine lymphomas/leukemias and could be used to screen for neoplasic disorders, minimal residual disease or monitor relapse.
- 4. PARR done when cytology, histopathology, and immunophenotyping is ambiguous.



### **Select Citations**

- Avery, A. 2009. Molecular diagnostics of hematologic malignancies. Top Companion Anim Med 24: 144-150.
- Burnett, R. C., W. Vernau, J. F. Modiano, C. S. Olver, P. F. Moore, and A. C. Avery. 2003. Diagnosis of canine lymphoid neoplasia using clonal rearrangements of antigen receptor genes. Vet Pathol 40: 32-41.
- Gelain, M. E., M. Mazzilli, F. Riondato, L. Marconato, and S. Comazzi. 2008. Aberrant phenotypes and quantitative antigen expression in different subtypes of canine lymphoma by flow cytometry. Vet Immunol Immunopathol 121: 179-188.
- Keller, R. L., A. C. Avery, R. C. Burnett, J. A. Walton, and C. S. Olver. 2004. Detection of neoplastic lymphocytes in peripheral blood of dogs with lymphoma by polymerase chain reaction for antigen receptor gene rearrangement. Vet Clin Pathol 33: 145-149.
- Lana, S., S. Plaza, K. Hampe, R. Burnett, and A. C. Avery. 2006. Diagnosis of mediastinal masses in dogs by flow cytometry. J Vet Intern Med 20: 1161-1165.
- Lara-Garcia, A., M. Wellman, M. J. Burkhard, C. Machado-Parrula, V. E. Valli, P. C. Stromberg, and C. G. Couto. **2008.** Cervical thymoma originating in ectopic thymic tissue in a cat. Vet Clin Pathol 37: 397-402.
- Pate, D. O., B. C. Gilger, S. E. Suter, and A. B. Clode. 2011. Diagnosis of intraocular lymphosarcoma in a dog by use of a polymerase chain reaction assay for antigen receptor rearrangement. J Am Vet Med Assoc 238: 625-630.
- **Kyoda, K., S. Nakamura, S. Matano, S. Ohtake, and T. Matsuda. 1997.** Prognostic significance of immunoglobulin heavy chain gene rearrangement in patients with acute myelogenous leukemia. Leukemia : official journal of the Leukemia Society of America, Leukemia Research Fund, U.K 11: 803-806.
- Werner, J. A., J. C. Woo, W. Vernau, P. S. Graham, R. A. Grahn, L. A. Lyons, and P. F. Moore. 2005. Characterization of feline immunoglobulin heavy chain variable region genes for the molecular diagnosis of B-cell neoplasia. Vet Pathol 42: 596-607.
- Wilkerson, M. J., K. Dolce, T. Koopman, W. Shuman, R. Chun, L. Garrett, L. Barber, and A. Avery. 2005. Lineage differentiation of canine lymphoma/leukemias and aberrant expression of CD molecules. Vet Immunol Immunopathol 106: 179-196.
- Williams, M. J., A. C. Avery, S. E. Lana, K. R. Hillers, A. M. Bachand, and P. R. Avery. 2008. Canine lymphoproliferative disease characterized by lymphocytosis: immunophenotypic markers of prognosis. J Vet Intern Med 22: 596-601.

