

Bayesian Statistics for Genetics 10b Guided tour of software

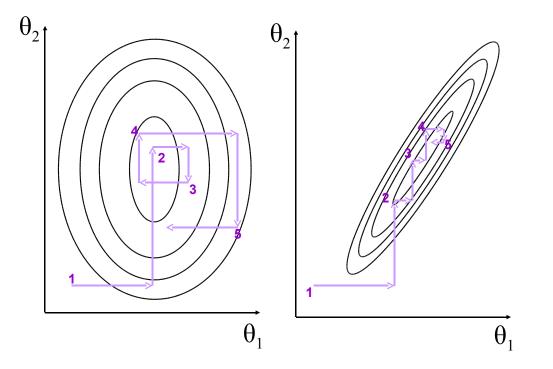
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Off-the-shelf MCMC

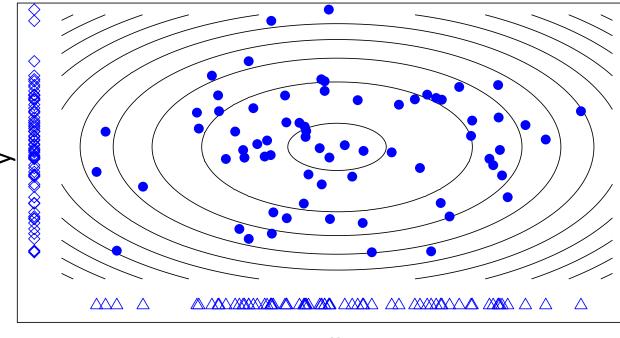
Recall the big picture of Bayesian computation;



We want a large sample from some distribution – i.e. the posterior. It **does not matter** if we get there by taking independent samples, or via some form of dependent sampling. (Gibbs Sampling, here)

Off-the-shelf MCMC

Once we have a big sample...



Sample (points) approximate distribution (contours)

Х

Any property of the actual posterior (contours) can be approximated by the *empirical* distribution of the samples (points)

Off-the-shelf MCMC

Markov Chain Monte Carlo (MCMC) is the general term for sampling methods that use Markov Chain processes to 'explore' the parameter space; the (many) random process values form our approximation of the posterior.

But in many settings this 'walking around' is mundane; once we specify the model and priors, the process of getting samples from the posterior can be done with no original thought – i.e. we can get a computer to do it.

Some example of this labor-saving approach;

- WinBUGS (next)
- ... or JAGS, OpenBUGS, NIMBLE and Stan
- INLA not a Monte Carlo method

The R Task Views on Genetics and Bayesian inference may also have specialized software; see also Bioconductor.

Bayes: WinBUGS

(τ) Y₁ Y₂ $Y_3 Y_4$ BUGS

Started in 1989, the **B**ayesian analysis **U**sing **G**ibbs 🔀 Sampling (BUGS) project has developed software where users specify only model and prior – everything else is internal. WinBUGS is the most comprehensive version.

- The model/prior syntax is very similar to R
- ... with some annoying wrinkles variance/precision, also column major ordering in matrices
- Can be 'called' from R see e.g. R2WinBUGS, but you still need to code the model

child cancers 'not caused Before we try it on GLMMs, a tiny GLM by Sellafield'



example
$$(n = 1, Y = 4);$$

$$Y|\theta \sim \text{Pois}(E \exp(\theta))$$

 $\theta \sim N(0, 1.797^2)$
 $E = 0.25$

One (sane) way to code this in the BUGS language;

```
model{
  lambda <- E*exp(theta) ...syntax follows R
  E <- 0.25
  theta \sim dnorm(m, tau)
  m < - 0
  tau <- 1/v
  v <- 1.797*1.797
```

```
Y \sim dpois(lambda) ... Poisson distribution, like R
                          ...constants could go in data
                          ... prior for \theta
                          tau = precision NOT variance!
                          ...finish the model
```

```
#data
list(Y=4)
#inits
list(theta=0)
```

```
Easiest way to input data
```

Same list format; or use gen.inits

Notes on all this; (not a substitute for reading the manual!)

- This should look familiar, from the models we have been writing out. In particular ' \sim ' is used to denote distributions of data *and* parameters
- All 'nodes' appear once on the LHS; hard work is done on RHS
- No formulae allowed when specifying distributions
- Data nodes *must* have distributions. Non-data nodes *must* have priors – it's easy to forget these
- Write out regressions 'by hand'; beta0 + beta1*x1 + ...
- This language can't do everything; BUGS does not allow e.g.
 Y <- U + V

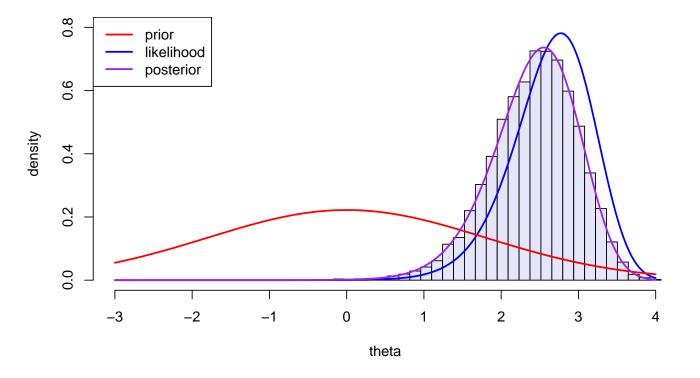
U~dnorm(meanu,tauu); V~dt(meanv,tauv,k)

#data

```
list(Y=...)
```

Bayes: WinBUGS

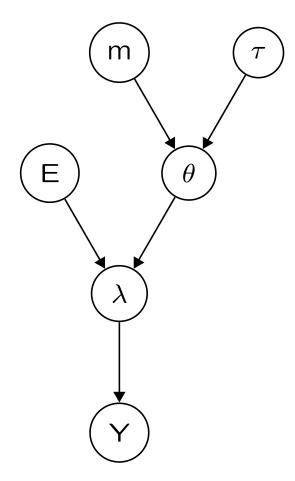
From 10,000 iterations, how do we do? (Note 'MC error' estimates Monte Carlo error in the posterior mean)



Histogram of WinBUGS output

nodemeansdMC error2.5%median97.5%theta2.4220.56080.0052461.2292.4663.388

Under the hood, here's how WinBUGS 'thinks';



- It's a DAG; arrows represent stochastic relationships (not causality)
- Some texts use square nodes for observed variables (Y, here)
- To do a Gibbs update, we need to know/work out the distribution of a node conditional on only its parents, children, and its children's other parents*.

* This set is a node's 'Markov blanket'. The idea saves a lot of effort, and is particularly useful when fitting random effects models.

WinBUGS: HWE example

A multinomial example, with a default prior;

$$\begin{array}{rcl} \mathbf{Y} & \sim & \mathsf{Multinomial}(n, \boldsymbol{\theta}) \\ \text{where } \boldsymbol{\theta} & = & (p^2, 2p(1-p), (1-p)^2) \\ p & \sim & \mathsf{Beta}(0.5, 0.5). \end{array}$$

And a typical way to code it in "the BUGS language";

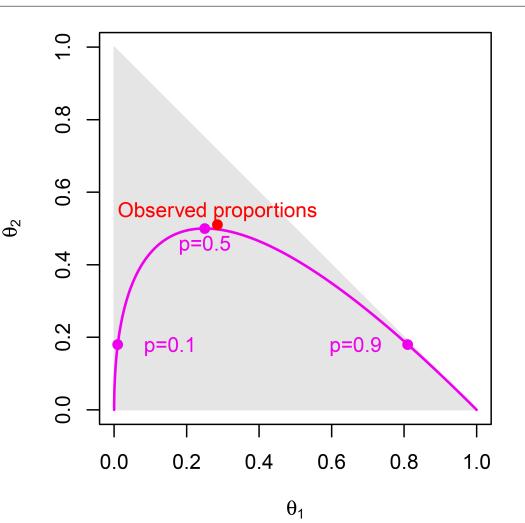
```
model{
    y[1:3] ~ dmulti(theta[], n)
    theta[1] <- p*p
    theta[2] <- 2*p*(1-p)
    theta[3] <- (1-p)*(1-p)
    p ~ dbeta(0.5, 0.5)
}</pre>
```

WinBUGS: HWE example

We have n = 186, and Y = (53,95,38).

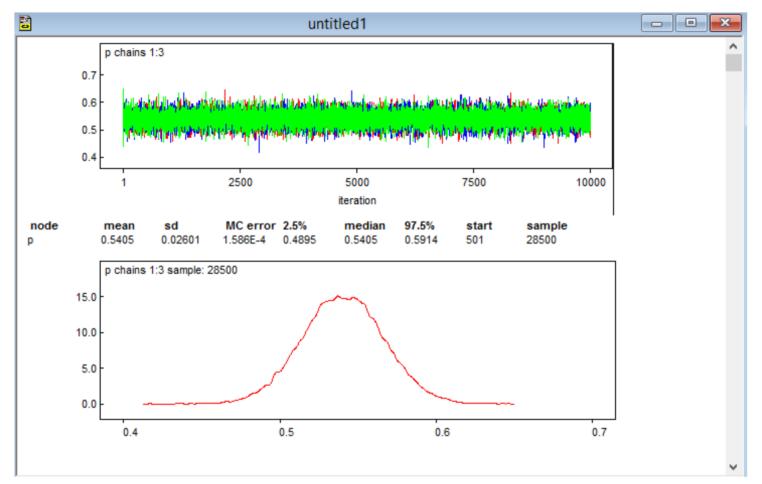
We will run 3 chains, starting at p = 0.5, 0.1 and 0.9.

In WinBUGS, input these by highlighting two list objects:



Data list(y=c(53,95,38),n=186) # Initial values
 list(p=0.5)
 list(p=0.1)
 list(p=0.9)

WinBUGS unlovely but functional in-house output;

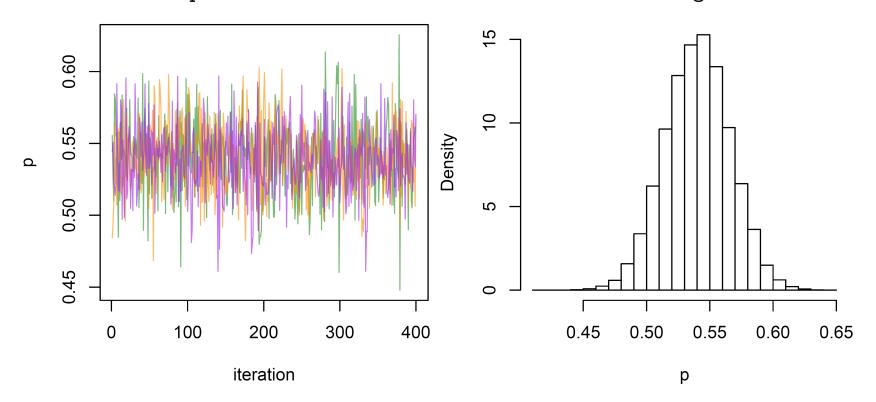


The posterior has 95% support for $p \in (0.49, 0.59)$, the posterior mean = posterior median = 0.54. Use code to get the chain(s).

Apart from coming up with the model, everything can be automated, using R's R2WinBUGS package;

- Model code now in a separate file (hweprog.txt)
- Specify the data and initial values as R structures
- Tell R where to find WinBUGS
- The output is stored in hweout, an R object no need to go via coda
- When debugging, pointy-clicky WinBUGS is still useful
- See next slide for less-clunky graphics

> print(hweout, digits=3) Inference for Bugs model at "hweprog.txt", fit using WinBUGS, 3 chains, each with 10000 iterations (first 500 discarded) n.sims = 28500 iterations saved 2.5% 50% 97.5% Rhat sd n.eff mean 0.540 0.026 0.490 0.541 0.590 1.001 28000.000 For each parameter, n.eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor (at convergence, Rhat=1).



- As well as the Markov blanket idea, WinBUGS uses what it knows about conjugacy to substitute closed form integrals in the calculations, where it can. (e.g. using inverse-gamma priors on Normal variances)
- Otherwise, it chooses from a hierarchy of sampling methods
 though these are not cutting-edge
- Because of its generality, and the complexity of turning a model into a sampling scheme, don't expect too much help from the error messages
- Even when the MCMC is working correctly, it is possible you may be fitting a ridiculous, unhelpful model. WinBUGS' authors assume you take responsibility for that

Also, while Gibbs-style sampling works well in many situations, for some problems it's not a good choice. If unsure, check the literature to see what's been tried already.

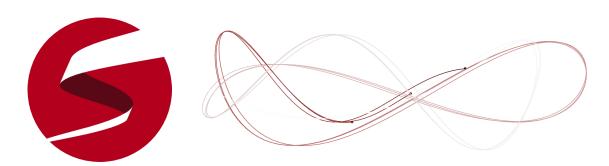
WinBUGS is no longer updated, but it's pointy-clicky interface remains a good place to get started. The BUGS language, describing models, is now used in JAGS, NIMBLE and OpenBUGS. Here's rjags using the **exact** same model file we just saw;

```
> library("rjags")
> jags1 <- jags.model("hweprog.txt", data=list(y=c(53,95,38),n=186) )</pre>
> update(jags1, 10000)
> summary( coda.samples(jags1, "p", n.iter=10000) )
Iterations = 11001:21000
Thinning interval = 1
Number of chains = 1
Sample size per chain = 10000
1. Empirical mean and standard deviation for each variable,
   plus standard error of the mean:
        Mean
                     SD
                            Naive SE
                                         Time-series SE
   0.5398583 0.0258055 0.0002581
                                              0.0003308
2. Quantiles for each variable:
 2.5%
          25%
                50%
                        75%
                               97.5%
0.4890 0.5225 0.5398 0.5576 0.5895
```

JAGS uses C, so is easier to extend than WinBUGS.

Stan

Stan is similar to BUGS, WinBUGS, JAGS etc – but new & improved;



- Coded in C++, for faster updating, it runs the No U-Turn Sampler – cleverer than WinBUGS' routines
- The rstan package lets you run chains from R, just like we did with R2WinBUGS
- Some modeling limitations no discrete parameters but becoming popular; works well with some models where WinBUGS would struggle
- Basically the same modeling language as WinBUGS but Stan allows R-style vectorization
- Requires declarations (like C++) unlike WinBUGS, or R so models require a bit more typing...

Stan: HWE example

A Stan model for the HWE example

```
data {
    int y[3];
}
parameters {
   real<lower=0,upper=1> p;
}
transformed parameters {
   simplex[3] theta;
   theta[1] = p*p;
   theta[2] = 2*p*(1-p);
   theta[3] = (1-p)*(1-p);
}
model {
   p<sup>~</sup>beta(0.5, 0.5);
   y<sup>multinomial(theta);</sup>
}
```

- More typing than BUGS!
- But experienced programmers will be used to this overhead

Stan: HWE example

With the model stored in HWEexample.stan (a text file) the rest follows as before;

```
> library("rstan")
> stan1 <- stan(file = "HWEexample.stan", data = list(y=c(53,95,38)),</pre>
+ iter = 10000, chains = 1)
> print(stan1)
Inference for Stan model: HWEexample.
1 chains, each with iter=10000; warmup=5000; thin=1;
post-warmup draws per chain=5000, total post-warmup draws=5000.
                                                  75% 97.5% n_eff
                       sd
                            2.5%
                                     25%
                                            50%
          mean se_mean
                                                 0.56 0.60
          0.54 0.00 0.03 0.48 0.52 0.54
                                                              5000
р
theta[1] 0.29 0.00 0.03 0.23 0.27 0.29 0.31 0.36
                                                              5000
theta[2] 0.49 0.00 0.01 0.48 0.49 0.50 0.50
                                                              4541
                                                        0.50
theta[3] 0.21 0.00 0.03 0.16 0.19 0.21
                                                 0.23
                                                        0.27
                                                              5000
       -192.17 0.02 0.87 -194.71 -192.44 -191.81 -191.57 -191.49
lp__
                                                              2762
```

Samples were drawn using NUTS(diag_e) at Tue Jul 26 14:13:31 2016.

- Iterations in the stan1 object can be used for other summaries, graphs, etc
- lp₋₋ is the log likelihood, used in (some) measures of model fit

INLA

We've already seen various examples of Bayesian analysis using Integrated Nested Laplace Approximation (INLA). For a (wide) class of models known as Gaussian Markov Random Fields, it gives a very accurate approximation of the posterior by 'adding up' a series of Normals.

- This approximation is not stochastic it is not a Monte Carlo method
- Even with high-dimensional parameters, where MCMC works less well/badly, INLA can be practical
- INLA is so fast that e.g. 'leave-one-out' & bootstrap methods are practical and can scale to GWAS-size analyses
- Fits most regression models but not everything, unlike MCMC
- Non-standard posterior summaries require more work than manipulating MCMC's posterior sample

INLA

The inla package in R has syntax modeled on R's glm() function. And with some data reshaping, our HWE example is a GLM;

```
> y <- c(53,95,38) # 2,1,0 copies of allele with frequency "p"
> n <- 186
> longdata <- data.frame(y=rep(2:0, y), ni=rep(2, n) )</pre>
> # non-Bayesian estimate of log(p)/(1-log(p)) i.e. log odds
> glm1 <- glm( cbind(y,ni-y) ~ 1, data=longdata, family="binomial" )</pre>
> expit <- function(x) {\exp(x)/(1+\exp(x))}
> expit(coef(glm1))
(Intercept)
  0.5403226
> expit(confint(glm1))
    2.5 % 97.5 %
0.4895317 0.5905604
> inla1 <- inla( y~1, family="binomial", data=longdata, Ntrials=rep(2,n) )</pre>
> summary(inla1)$fixed
                      sd 0.025quant 0.5quant 0.975quant mode kld
             mean
(Intercept) 0.1616 0.104
                            -0.0422 0.1615
                                             0.3661 0.1612 0
> expit(summary(inla1)$fixed[,3:5]) # posterior of "p"
0.025quant 0.5quant 0.975quant
0.4894516 0.5402875 0.5905163
```

For non-default priors, see the examples on the course site.