## REPORT

## MOLECULAR KNOTS

# Braiding a molecular knot with eight crossings 

Jonathan J. Danon, Anneke Krüger, David A. Leigh, * Jean-François Lemonnier, Alexander J. Stephens, Iñigo J. Vitorica-Yrezabal, Steffen L. Woltering


#### Abstract

Knots may ultimately prove just as versatile and useful at the nanoscale as at the macroscale. However, the lack of synthetic routes to all but the simplest molecular knots currently prevents systematic investigation of the influence of knotting at the molecular level. We found that it is possible to assemble four building blocks into three braided ligand strands. Octahedral iron(II) ions control the relative positions of the three strands at each crossing point in a circular triple helicate, while structural constraints on the ligands determine the braiding connections. This approach enables two-step assembly of a molecular $8_{19}$ knot featuring eight nonalternating crossings in a 192-atom closed loop $\sim 20$ nanometers in length. The resolved metal-free $8_{19}$ knot enantiomers have pronounced features in their circular dichroism spectra resulting solely from topological chirality.


Knots (1) form spontaneously in flexible polymer chains (2) and are found in circular DNA (3) and $\sim 1 \%$ of proteins in the Protein Data Bank (PDB) (4). To date, only three nontrivial knot topologies (trefoil, figure-eight, and pentafoil) (5-16) out of more than 6 billion (17, 18) known prime knots have been synthesized with small molecules. Most of the molecular knots isolated to date have been unanticipated reaction products ( $6,7,10,12,14$ ), but a few designed strategies for the construction of simple small-molecule knots have been successfully developed, including the use of linear (5) and cyclic (11) double helicates that twist two ligand strands about each other several times (13).
Every knot can be formed by braiding (19) strings and joining their ends together (Fig. 1) (20). However, the number of strings required for a particular knot depends on its topology. Twisting two strings about each other can generate only four of the 84 prime knots with up to nine crossings, whereas having three strings to braid makes nearly one-third of such knots accessible (21). Sauvage synthesized the first molecular trefoil knot (5), as well as a Solomon link (22), using linear double helicates in which two ligand strands are twisted about one another to generate the required number of crossings (three and four, respectively), followed by joining the termini of each strand along the length of the helicate (Fig. 1A). Recently, a Solomon link (23), pentafoil knots (11, 24) (five crossings), and a Star of David catenane (25) (six crossings) were prepared using double helicates that are cyclic, which reduces the distances needed to join the strands into closed loops (Fig. 1B). These circular helicates use Fe (II) ions to entwine two

[^0]ligand strands: Two bipyridine (bipy) chelating groups coordinated to the same metal ion are connected in the knot-forming reaction to form a strand that crosses that of the third bipy unit bound to the metal ion. However, the octahedral coordination geometry of these metal ions could, in principle, be used to position three bipycontaining strands with respect to each other. The key is to design ligands so that connections can only be made to bipy groups that are coordinated to adjacent metal ions. In this way the octahedral $\mathrm{Fe}(\mathrm{II})$ metal ions can control the relative position of three ligand strands at each crossing point (Fig. 1C)

The $8_{19}$ knot [Alexander-Briggs notation, where
the main number denotes the number of crossings and the subscript differentiates the knot from others with the same number of crossings (20)] is one of 21 prime knots with eight crossings (20). Along with the $8_{20}$ and $8_{21}$ knots, the $8_{19}$ knot is the simplest nonalternating knot; it is absent from early knot tables (26) because it was not immediately recognized (27) [much less proven (28)] that not all knots can be written in a form that alternates crossings over-under around the closed loop. The $8_{19}$ knot is the $\mathrm{T}(3,4)$ torus knot, meaning that it can be represented by a closed loop that wraps around a torus (doughnut shape) three times longitudinally and four times meridionally without crossing itself (fig. S18) (20). Its most symmetrical form suggests a strategy to access it through a three-stranded braid using a four-fold repeat motif (Fig. 1C). Indeed, recent simulations showed an $8_{19}$ knot to be a favored structure in the theoretical assembly of four rigid helical building blocks with sticky ends (29). We envisaged employing a tetrameric circular triple helicate (30) within which each of four metal ions binds to bipy units from three different ligand strands. Rigid extensions from the bipy groups at both ends of each strand ensure that the terminal alkene groups are positioned such that they can only undergo olefin metathesis to groups coordinated to adjacent $\mathrm{Fe}(\mathrm{II})$ ions, producing the connections required to form an $8_{19}$ knot (Fig. 2).
Ligand strand $\mathbf{1}$ and $\mathrm{FeCl}_{2}$ were heated at $130^{\circ} \mathrm{C}$ in $N, N$-dimethylformamide (DMF) for 24 hours (Fig. 2, step 1). Subsequent treatment with methanolic $\mathrm{KPF}_{6}$ afforded a red precipitate, determined by electrospray ionization mass spectrometry (ESI-MS) to be a tetrameric complex $\left[\mathrm{Fe}_{4} \mathbf{1}_{4}\right]\left(\mathrm{PF}_{6}\right)_{8}$ (Fig. 2, step 2). Carrying out the assembly process in solvents other than DMF [e.g., ethylene glycol or dimethylsulfoxide (DMSO)] produced substantial quantities of a trimer, $\left[\mathrm{Fe}_{3} \mathbf{1}_{3}\right]\left(\mathrm{PF}_{6}\right)_{6}-$
 through connections a to $\mathrm{d}^{\prime}$, b to $\mathrm{e}^{\prime}$, c to $\mathrm{a}^{\prime}$, d to $\mathrm{b}^{\prime}$, and e to $\mathrm{c}^{\prime}$. (C) An $8_{19}$ knot can be synthesized from a cyclic tetrameric triple helicate. Four metal ions control eight crossing points for four building blocks assembled into a circular triple helix with connections made from a to $\mathrm{b}^{\prime}$, b to $\mathrm{c}^{\prime}, \mathrm{c}$ to $\mathrm{d}^{\prime}$, and d to $\mathrm{a}^{\prime}$. A feature of circular helicates is that the building blocks can be joined at different points along the braid to generate the desired number and sequence of crossings, rather than only at the braid ends.






Fig. 2. Synthesis of a molecular $\mathbf{8}_{\mathbf{1 9}} \mathbf{~ k n o t}$, 2. Reaction conditions: (1) $\mathbf{1}$ ( 1.0 equiv.), $\mathrm{FeCl}_{2}$ ( 1.0 equiv.), $N, N$-dimethylformamide (DMF), $130^{\circ} \mathrm{C}, 24$ hours; (2) $\mathrm{KPF}_{6}, \mathrm{MeOH}, 60 \%$ (two steps); (3) Hoveyda-Grubbs second-generation catalyst ( $25 \mathrm{~mol} \%$ per olefin metathesis reaction), $\mathrm{MeNO}_{2} / 1,2-$ dichloroethane $1: 1,60^{\circ} \mathrm{C}, 24$ hours; (4) $\mathrm{KPF}_{6}, \mathrm{H}_{2} \mathrm{O}, 62 \%$ (two steps); (5) $\mathrm{NaOH}_{(\mathrm{aq})}(1 \mathrm{M}) / \mathrm{MeCN} 1: 1,70^{\circ} \mathrm{C}, 30 \mathrm{~min}, 38 \%$.
presumed to be a linear triple helicate (25)-which could not easily be separated from the tetramer. The ${ }^{1} \mathrm{H}$ nuclear magnetic resonance (NMR) spectrum of $\left[\mathrm{Fe}_{4} \mathbf{1}_{4}\right]\left(\mathrm{PF}_{6}\right)_{8}$ (Fig. 3B) showed all four ligands to be in the same symmetric environment, which is possible only when ligands are arranged cyclically, with the diastereotopic splitting of protons $\mathrm{H}_{\mathrm{d}}$ and $\mathrm{H}_{\mathrm{e}}$ indicative of a helical conformation.

Tetrameric complex $\left[\mathrm{Fe}_{4} \mathbf{1}_{4}\right]\left(\mathrm{PF}_{6}\right)_{8}$ was treated with the Hoveyda-Grubbs second-generation catalyst (31) at $60^{\circ} \mathrm{C}$ in nitromethane/1,2-dichloroethane (1:1) (Fig. 2, step 3). After 24 hours, the reaction was quenched with ethyl vinyl ether and the product precipitated with aqueous $\mathrm{KPF}_{6}$ in an effort to confer anion uniformity and to remove catalyst residues (Fig. 2, step 4), furnishing $\left[\mathrm{Fe}_{4} \mathbf{2}\right]\left(\mathrm{PF}_{6}\right)_{8}$ in $62 \%$ yield over two steps. ESI-MS was consistent with the loss of four ethene molecules from $\left[\mathrm{Fe}_{4} \mathbf{1}_{4}\right]\left(\mathrm{PF}_{6}\right)_{8}$ (fig. S7), and ${ }^{1} \mathrm{H}$ NMR spec-
troscopy confirmed the loss of the terminal olefin signals from the precursor complex (Fig. 3C). In some experiments, ESI-MS indicated incomplete anion exchange and contamination of the product with a small amount of $\left[\mathrm{Fe}_{4} \mathbf{2}\right]\left(\mathrm{PF}_{6}\right)_{7} \mathrm{Cl}$ (fig. S13).

Slow evaporation of a saturated acetonitrile solution of the $\mathrm{Fe}_{4} 2$ knot complex produced red needles over several days suitable for singlecrystal x-ray diffraction. The solid-state structure confirms the topology and $D_{4}$ symmetry of the molecular $8_{19}$ knot (Fig. 4, movie S1, and supplementary materials). The organic ligand weaves a continuous 192-atom path passing each of the four coplanar iron centers three times to form a circular triple helicate closed loop with eight nonalternating crossings. Both $E$ - and $Z$-olefins are present in the chain, with some disorder, in approximately a $1: 1$ ratio. Each $\mathrm{Fe}(\mathrm{II})$ ion is bound to three bipy groups; the Fe-N distances are in
the range of 1.92 to $2.02 \AA$, as is typical (32) for $\mathrm{Fe}(\text { bipy })_{3}{ }^{2+}$ complexes. The metal ion octahedral coordination geometry is slightly distorted by the constraints of the tightly knotted ligand, with $\mathrm{N}-\mathrm{Fe}-\mathrm{N}$ angles within each bipyridyl moiety of $\sim 81^{\circ}$. The bipyridine groups define a cavity of diameter $3.30 \pm 0.02 \AA$ [versus $3.7 \pm 0.1 \AA$ for a related Solomon link (33)]. At the center, a chloride anion is bound by eight $\mathrm{CH} \cdots \mathrm{Cl}$ hydrogen bonds. Although the presence of the chloride ion is somewhat surprising given the anion exchange reaction with $\mathrm{KPF}_{6}$, it is reminiscent of the solid-state structures of anion template pentafoil knots $(11,24)$ and, as with those knots, we found the $\mathrm{Fe}_{4} 2$ knot complex to bind a single chloride anion strongly in solution (association constant $K_{\mathrm{a}}=2.6 \pm 0.1 \times 10^{8} \mathrm{M}^{-1}$ in $\mathrm{MeCN}, 298 \mathrm{~K}$; see supplementary materials).

Treatment of an acetonitrile solution of $\left[\mathrm{Fe}_{4} 2\right]\left(\mathrm{PF}_{6}\right)_{8}$ with $\mathrm{NaOH}(1 \mathrm{M})$ at $70^{\circ} \mathrm{C}$ for 30 min


Fig. 3. ${ }^{1} \mathrm{H}$ NMR spectra ( $\mathbf{6 0 0} \mathbf{~ M H z , 2 9 8 ~ K ) ~ o f ~ m o l e c u l a r ~} \mathbf{8}_{19} \mathbf{~ k n o t} 2$ and its complexes and precursors. (A) Ligand strand $\mathbf{1}$ (solvent $\mathrm{CDCl}_{3}$ ); (B) tetrameric circular helicate $\left[\mathrm{Fe}_{4} \mathbf{l}_{4}\right]\left(\mathrm{PF}_{6}\right)_{8}$ (solvent $\mathrm{CD}_{3} \mathrm{CN}$ ); (C) $8_{19}$ knot complex $\left[\mathrm{Fe}_{4} \mathbf{2}\right]\left(\mathrm{PF}_{6}\right)_{8}$ (solvent $\mathrm{CD}_{3} \mathrm{CN}$ ); ( $\mathbf{D}$ ) metal-free $8_{19}$ knot $\mathbf{2}$ (solvent $\mathrm{CDCl}_{3}$ ). The lettering corresponds to the proton labeling in Fig. 2. Full spectral assignments are given in the supplementary materials.


Fig. 4. X-ray crystal structure of molecular $8_{19}$ knot complex $\left[\mathrm{Fe}_{4} 2\right]\left(\mathrm{PF}_{6}\right)_{7} \mathrm{Cl}$ and circular dichroism (CD) spectra of each mirror image form of metal-free $\mathbf{8}_{\mathbf{1 9}} \mathbf{k n o t}$ ligand 2. (A) X-ray structure viewed perpendicular to the plane of the four Fe(II) ions. (B) Side view, in the plane of the four Fe(II) ions. C atoms are light gray (turquoise in one repeat unit of the knot); N , blue; O , red; Fe, purple. $\mathrm{PF}_{6}{ }^{-}$anions and hydrogen atoms omitted for clarity. The iron(II) centers in each molecular knot have the same coordination stereochemistry ( $\Lambda$ or $\Delta$ ). Both enantiomers are present in each unit cell; the all- $\Lambda$ enantiomer is shown here. (C) CD spectra of $8_{19}$ knot enantiomers (+)-2 (blue) and (-)-2 (red, $90 \%$ ee) in $\mathrm{CH}_{2} \mathrm{Cl}_{2}\left(1.76 \times 10^{-4} \mathrm{M}\right)$ at 298 K .
afforded the demetallated $8_{19}$ knot $\mathbf{2}$ in $38 \%$ yield (Fig. 2, step 5). Matrix-assisted laser desorption/ ionization mass spectrometry (MALDI-MS) gave
an isotope distribution for the $[\mathbf{2} \cdot \mathrm{Na}]^{+}$molecular ion that matched the theoretical pattern and values (fig. S10). The ${ }^{1}$ H NMR spectrum of 2 (Fig. 3D)
is relatively broad in comparison to both the metallated knot $\left[\mathrm{Fe}_{4} 2\right]\left(\mathrm{PF}_{6}\right)_{8}$ (Fig. 3C) and building block 1 (Fig. 3A). Molecular modeling and
variable temperature ${ }^{1} \mathrm{H}$ NMR (fig. S16) suggest that $\mathbf{2}$ does not exist in a single well-defined conformation and that the broad features of the ${ }^{1} \mathrm{H}$ NMR spectrum are likely a consequence of reptation [the thermal motion of long, entangled polymer chains (34)] as the 192-atom-long knot backbone's motion is restricted when it passes through tight loops and traverses crossings. Furthermore, most of the signals in the ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{2}$ (Fig. 3D) are shielded with respect to that of building block $\mathbf{1}$ (Fig. 3A); this reflects the compact structure of the knot bringing each region of the loop into close proximity with aromatic rings during reptation.

The $8_{19}$ knot is intrinsically chiral by virtue of its topology. One enantiomer, (+)-2 (sign determined by optical rotation), could be isolated by chiral high-performance liquid chromatography (see supplementary materials) and a sample of the other enantiomer obtained in highly enriched form [(-)-2, 90\% enantiomeric excess (ee)] (35). The mirror-image $8_{19}$ knots have circular dichroism (CD) spectra of equal and opposite shape and sign (Fig. 4C) $(35,36)$. Despite the knots possessing no elements of Euclidean chirality, several features of the CD spectra (particularly the maxima and minima at $221,250,294,327$, and 351 nm ) show that the chiral environment is well expressed despite the dynamics of the closed loop and the absence of well-defined conformations, presumably also a result of the tightness of the knotting.

The braiding of ligand strands should enable the synthesis of many more molecular knots [and links $(25,37)$ ] of greater complexity than has previously been possible (fig. S19). The inherent chirality of many knots, and the tightness of the knotting possible through this synthetic approach, may prove useful when investigating
applications of knotting in molecular tools (24, 33, 38) and materials (39, 40).

## REFERENCES AND NOTES

1. N. C. H. Lim, S. E. Jackson, J. Phys. Condens. Matter 27, 354101 (2015).
2. M. D. Frank-Kamenetskii, A. V. Lukashin, A. V. Vologodskii, Nature 258, 398-402 (1975).
3. S. A. Wasserman, N. R. Cozzarelli, Science 232, 951-960 (1986).
4. J. I. Sułkowska, E. J. Rawdon, K. C. Millett, J. N. Onuchic, A. Stasiak, Proc. Natl. Acad. Sci. U.S.A. 109, E1715-E1723 (2012).
5. C. O. Dietrich-Buchecker, J.-P. Sauvage, Angew. Chem. Int. Ed. Engl. 28, 189-192 (1989).
6. O. Safarowsky, M. Nieger, R. Fröhlich, F. Vögtle, Angew. Chem. Int. Ed. 39, 1616-1618 (2000).
7. M. Feigel, R. Ladberg, S. Engels, R. Herbst-Irmer, R. Fröhlich, Angew. Chem. Int. Ed. 45, 5698-5702 (2006).
8. J. Guo, P. C. Mayers, G. A. Breault, C. A. Hunter, Nat. Chem. 2, 218-222 (2010).
9. P. E. Barran et al., Angew. Chem. Int. Ed. 50, 12280-12284 (2011).
10. N. Ponnuswamy, F. B. L. Cougnon, J. M. Clough, G. D. Pantoş, J. K. M. Sanders, Science 338, 783-785 (2012)
11. J.-F. Ayme et al., Nat. Chem. 4, 15-20 (2011)
12. T. Prakasam et al., Angew. Chem. Int. Ed. 52, 9956-9960 (2013).
13. J.-F. Ayme, J. E. Beves, C. J. Campbell, D. A. Leigh, Chem. Soc. Rev. 42, 1700-1712 (2013)
14. N. Ponnuswamy, F. B. L. Cougnon, G. D. Pantoş, J. K. M. Sanders, J. Am. Chem. Soc. 136, 8243-8251 (2014)
15. J.-F. Ayme et al., J. Am. Chem. Soc. 136, 13142-13145 (2014).
16. G. Gil-Ramírez et al., J. Am. Chem. Soc. 138, 13159-13162 (2016).
17. J. Hoste, in Handbook of Knot Theory, W. Menasco, M. Thistlethwaite, Eds. (Elsevier, 2005), pp. 209-232.
18. E. E. Fenlon, Eur. J. Org. Chem. 2008, 5023-5035 (2008)
19. T. Ciengshin, R. Sha, N. C. Seeman, Angew. Chem. Int. Ed. 50, 4419-4422 (2011)
20. C. C. Adams, The Knot Book (Freeman, 1994).
21. V. F. R. Jones, Ann. Math. 126, 335-388 (1987).
22. J.-F. Nierengarten, C. O. Dietrich-Buchecker, J.-P. Sauvage J. Am. Chem. Soc. 116, 375-376 (1994).
23. J. E. Beves, C. J. Campbell, D. A. Leigh, R. G. Pritchard Angew. Chem. Int. Ed. 52, 6464-6467 (2013).
24. V. Marcos et al., Science 352, 1555-1559 (2016).
25. D. A. Leigh, R. G. Pritchard, A. J. Stephens, Nat. Chem. 6, 978-982 (2014).
26. P. G. Tait, Trans. R. Soc. Edinb. 32, 327-342 (1883-1884).
27. C. N. Little, Trans. R. Soc. Edinb. 35, 663-664 (1889).
28. K. Murasugi, Osaka Math. J. 12, 277-303 (1960).
29. G. Polles, D. Marenduzzo, E. Orlandini, C. Micheletti, Nat. Commun. 6, 6423 (2015).
30. B. Hasenknopf et al., J. Am. Chem. Soc. 119, 10956-10962 (1997).
31. S. B. Garber, J. S. Kingsbury, B. L. Gray, A. H. Hoveyda, J. Am. Chem. Soc. 122, 8168-8179 (2000).
32. M. E. García Posse et al., Inorg. Chem. 23, 948-952 (1984).
33. J.-F. Ayme et al., J. Am. Chem. Soc. 137, 9812-9815 (2015)
34. P. G. de Gennes, J. Chem. Phys. 55, 572-579 (1971).
35. F. Vögtle et al., Angew. Chem. Int. Ed. 40, 2468-2471 (2001).
36. G. Rapenne, C. Dietrich-Buchecker, J.-P. Sauvage, J. Am. Chem. Soc. 118, 10932-10933 (1996)
37. K. S. Chichak et al., Science 304, 1308-1312 (2004)
38. R. A. Bilbeisi et al., Chem. Sci. 7, 2524-2531 (2016)
39. A. M. Saitta, P. D. Soper, E. Wasserman, M. L. Klein, Nature 399, 46-48 (1999).
40. Y. Liu et al., Science 351, 365-369 (2016).

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## SUPPLEMENTARY MATERIALS

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Materials and Methods
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Editor's Summary

## Three strands ironed closely together

It is not uncommon when braiding hair or bread to intertwine three different strands. At the molecular level, however, synthetic knots have thus far been restricted to architectures accessible from two-strand braids. Danon et al. used iron ion coordination to guide three organic ligand strands to form a knot geometry with eight separate crossings.

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