

Calibration & Use of a Capintec CAPTUS 3000 Portable Thyroid Uptake System for Iodine-125 Bioassay Measurements

Todd W. Baker, MSPH, CHP





The Effects of Environmental Chemicals on Neuroendocrine Function: Mode and Mechanism for Adverse Outcomes

Presenters: R.L. Cooper, T. E. Stoker, J.M. Goldman and S.C. Laws

Contributing Organizations: NHEERL, NCCT, NCSU



B · O · S · C HUMAN HEALTH PROGRAM REVIEW

RESEARCH TRIANGLE PARK, NORTH CAROLINA JANUARY 13-15 2009

RESEARCH & DEVELOPMENT

LTG 1 Poster 05

Science Questions

Can a strategy be developed that will provide the means to determine the neuronal/molecular targets for environmental toxicants that alter the hypothalamic/pituitary control of gonadal function?

Research Goals

The hypothalamus and pituitary play critical roles in reproduction. In the female rat (and human), the mid-cycle (or ovulatory) surge of luteinizing hormone (LH) is a functional endocrine event that stimulates the final stages of follicular and oocyte maturation prior to ovulation. We have shown that environmental toxicants from a variety of chemical classes are able to block this highly coordinated endocrine event. For example, the formamides and dithiocarbamate pesticides block the catecholaminergic control of gonadotropin-releasing hormone (GnRH), and the chlorotriazines herbicides and intermediate metabolites affect the release of GnRH by a mechanism yet to be determined. We have also demonstrated that the disruption of the LH surge is a key event in the mode of action that leads to adverse reproductive outcomes including impaired pregnancy maintenance, impaired embryo development, reduced fertility, delayed puberty, premature reproductive senescence and mammary gland tumors in the female.

Due to the complexity of the observed toxicant-induced disruption of the neuroendocrine control of LH secretion and the numerous mechanisms that can be involved, we are developing a strategy to rapidly identify those cellular and molecular targets within the brain-pituitary that are affected by exposure to a test chemical. This strategy combines a series of *in vitro* (cell culture) and *ex vivo* (tissue perfusion, brain slices) procedures to identify the numerous mechanisms associated with the disruption of pituitary-gonadal and pituitary-adrenal function. We are coordinating our test chemical selection in order to integrate our results with those of the National Computational Center for Toxicology's ToxCast Research Program based on the NovAScreen® database. This approach will provide the information needed to determine the mode of action underlying the adverse effects of the chemical and provide the basis for across species comparisons. A related effort examines the extent to which toxicant-induced changes in the pituitary-adrenal axis contribute to alterations in reproductive physiology.

Identifying the Range of Chemicals

Effects of Environmental Agents on Neuroendocrine Function: Identifying Common Modes of Action and Adverse Outcomes

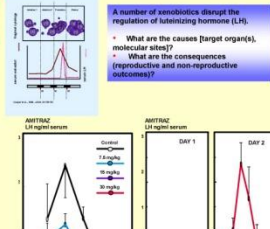
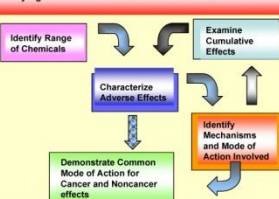


Table 1. Range of Chemicals That Modify the LH Surge

Formamides	Chlorotriazines	Organophosphates
Chloridifens	Atrazine (+)	Malathion
Acetates	Propazine (+)	Trioxolon (-)
Diphenyl Herbicides	Diuron (+)	Pyrethroids
2,4-D (diphenyl-(-)-isomers)	Dequalylazine (+)	Permethrin
Thiocarbamates	Dequalylazine (+)	Cypermethrin
Methidathion	Acetochlor (+)	Fenprothrin
Telidate	Acetochlor (+)	Fenprothrin
Dithiocarbamates	Halocacetic Acids	Formaldehyde
Tioxins (+)	Pharmaceuticals	
Metham Sodium	Sodium Valproate	
Fenitrothion	Triclosan (-)	
Sodium-Dimethylthiocarbamate	Pyrethroids	
Organophosphate	Pyrethroids	
Malathion	Permethrin	
	Cypermethrin	
	Fenprothrin	

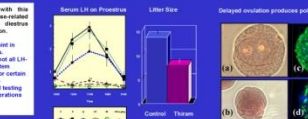
Red letters indicate positive effect on LH surge (+) effect or (-) no effect or onset of puberty.

Characterizing Adverse Outcomes

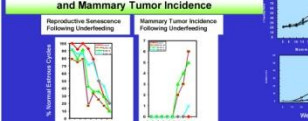
Evaluating Ovarian Cycles using Vaginal Cytology



Evaluating Oocyte Function and Fertility



Correlation Between Reproductive Senescence and Mammary Tumor Incidence



Evaluating Effects of Compounds on Pubertal Development



Impact and Outcomes

By establishing parallel dose-relationships among effects on hypothalamic catecholamines, GnRH neuronal activation and the LH surge in response to DMDC, our work has identified the toxic pathway for the adverse effect of this class of metal-chelating pesticides on female reproductive function.

Our research strategy enabled us to elucidate common (or disparate) modes of action of atrazine metabolites and other chlorotriazine herbicides, information that contributed to the assessment of the cumulative risks of this widely used class of compounds by OPP and OVR.

In vitro and *in vivo* studies defining the effects of atrazine and metabolites on aromatase and steroidogenesis have been key for interpreting recent reports in the literature suggesting that the adverse effects of this compound are mediated by alterations in aromatase activity in cell lines and wildlife species.

Furthermore, this research has now identified perturbations in the hypothalamic-pituitary-adrenal axis on the reproductive tract as new areas of concern.

Future Directions

- * Further identify the extent to which changes in the adrenal axis mediate the adverse effects of xenobiotics on reproduction.
- * Characterize the extent to which HPA may mediate selected chemical-induced developmental dysfunction (e.g., pubertal development, blood pressure changes etc.)
- * Characterize toxic effects on the relationships among the neuroendocrine factors contributing to the hypothalamic secretion of GnRH.
- * Collaborate with NCCT investigators, using their databases, to more rapidly:

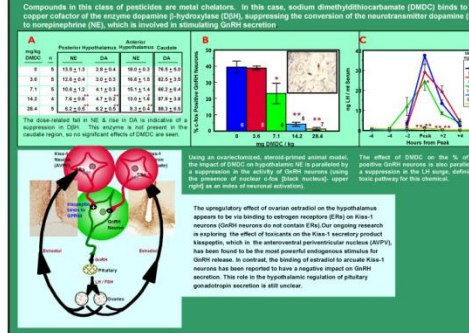
- identify the cellular mechanisms associated with altered neuroendocrine function, and
- identify compounds for further evaluation of their effects on these two important neuroendocrine systems

Other Researchers Involved

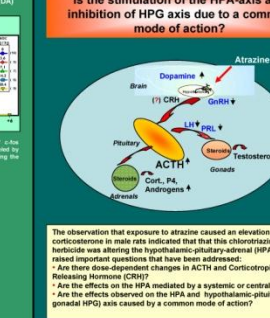
Michael Narotsky, John Laskey, David Dix, Keith Hawk, Angela Bucklewell, Michelle Holckins, Ashley Munn

Contributors: NHEERL: Goldman; Frailes; Narotsky, Laskey, NCSU: Cole of Veterinary Medicine: Zornis, Tiroff; NCCT: Dix, Hawk

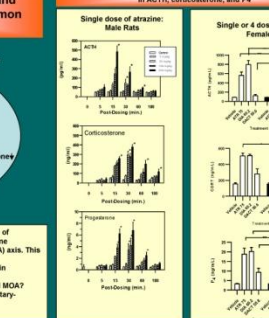
Defining Toxicity Pathways



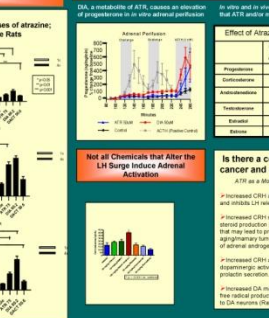
Is the stimulation of the HPA-axis and inhibition of HPG axis due to a common mode of action?



Additional Evidence that Atrazine Alters Steroidogenesis



Not all Chemicals that Alter the LH Surge Induce Adrenal Activation



Use of Mechanistic Data in Risk Assessment

This poster does not necessarily reflect EPA policy. Mention of trade names or

DIRECT AFFECTS OF ATRAZINE AND ITS METABOLITE DEISOPROPYL-ATRAZINE (DIA) ON PITUITARY AND ADRENAL HORMONE RELEASE USING *IN VITRO* PERFUSION

Hotchkiss MG, Cooper RL, and Laws SC

U.S. Environmental Protection Agency, Office of Research and Development, National Health & Environmental Effects Laboratory
Endocrinology Branch, Reproductive Toxicology Division, Research Triangle Park, NC

Abstract

Atrazine, a broad spectrum herbicide, is one of the most widely used herbicides in the United States, with current total annual use of approximately 76 million pounds of active ingredient. Previous work in our laboratory has shown that atrazine (ATR) and its metabolite deisopropyl-atrazine (DIA) are able to induce a dose-dependent increase in plasma ACTH, serum corticosterone (CORT) and progesterone (P4), at 15 and 30 min after a single oral administration *in vivo*. The mechanism for this increase in hypothalamic-pituitary-adrenal axis (HPA-axis) activation is currently unknown. To test whether ATR and DIA have a direct agonistic effect on the pituitary or adrenal glands, we conducted an *in vitro* perfusion, exposing pituitary and adrenal tissue to 50µM concentrations of either ATR or DIA during two twenty minute challenges over a three hour period. A positive control challenge was also used for both pituitary (10nM CRF) and adrenal (1nM ACTH) tissues, as well as a final challenge to test for viability of the tissue at the completion of the perfusion. Results indicate that ATR and DIA have no direct agonistic effect on pituitary or adrenal release of ACTH or CORT respectively; however treatment of adrenal tissue with 50µM DIA caused an approximate 100% increase over basal P4 release over the three hour perfusion, representing an increase from 3 pg/mg/min to 7 pg/mg/min at the 230min timepoint. While these results do not elucidate the mechanism by which ATR and DIA activate the HPA-axis after a single administration *in vivo*, increased release of P4 from the adrenal observed after *in vitro* treatment with DIA demonstrates a direct agonistic effect of the chemical on adrenal tissue, and one potential mechanism by which ATR and DIA are able to alter steroidogenesis and produce adverse reproductive outcomes *in vivo*.

Methods/Approach

Animals. For all experiments, male Wistar rats (80 days-old), obtained from Charles Rivers Laboratories, Raleigh, NC, and were housed 1 per cage under controlled temperature (20-24°C), humidity (40-50%), and light conditions (12h light/12h dark, lights on 0600) with Purina Laboratory chow (5051) and water available *ad libitum*.

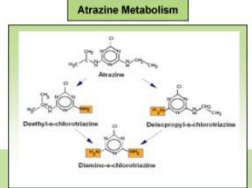
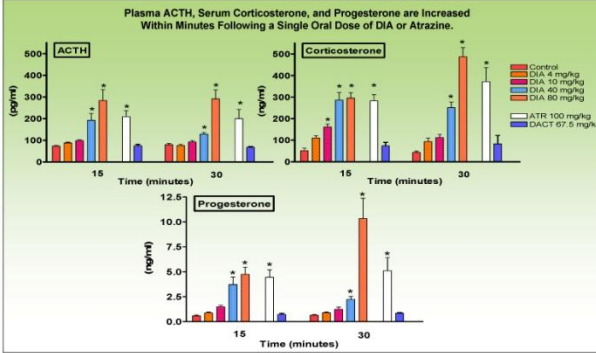
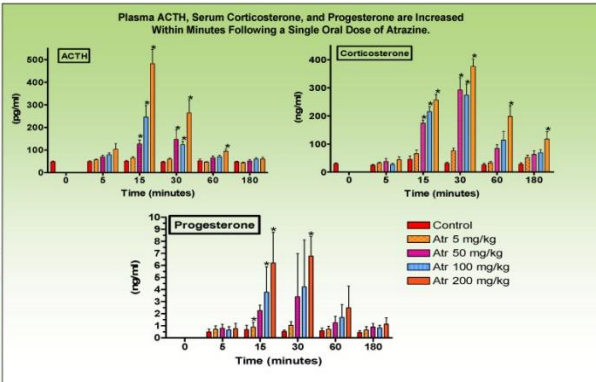
Dosing solutions and procedures. For all experiments except the pituitary perfusion, rats were pre-dosed with vehicle (methyl cellulose) for 7 days to acclimate animals to handling and dosing. All dosing was conducted from 0700-0800 when both ACTH and corticosterone were at basal concentrations. Atrazine, DIA, or DACT (97% purity, a gift from Syngenta Crop Protection, Greensboro, NC) were prepared as a suspension in 1.2% methyl cellulose.

Hormone Experimental design. Groups (n=10) of animals were dosed with a single dose of 0 (vehicle), 5, 50, 100, or 200 mg/kg bw atrazine, 67.5mg/kg bw DACT, or 4, 15, 40, or 80mg/kg bw DIA by oral gavage in a volume of 5.0 mL dosing solution/kg bw, and killed by decapitation 5, 15, 30, 60 or 180 minutes later. Plasma and serum were collected and frozen for the ACTH, progesterone, and corticosterone assays, respectively.

Pituitary and Adrenal Perfusion Experimental Design. The perfusion apparatus was set up as described in Goldman and Cooper (1993). Pituitary glands were removed and hemisected, while adrenal glands were quartered before placement in the perfusion apparatus. Treatment of the glands included two subsequent challenges with the test chemical, followed by one 60min KCl(pituitary) or 10µM ACTH(adrenal) challenge at the conclusion of the experiment to vary tissue viability. Perfusates were collected at 10min intervals and frozen for RIA analysis.

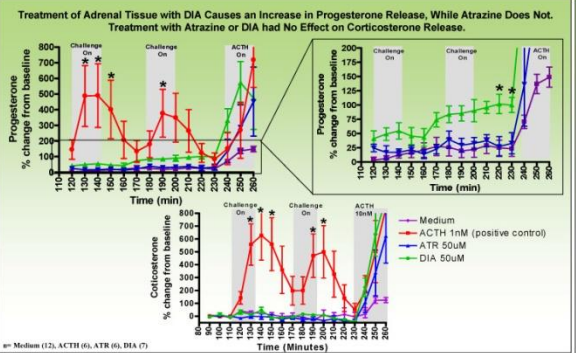
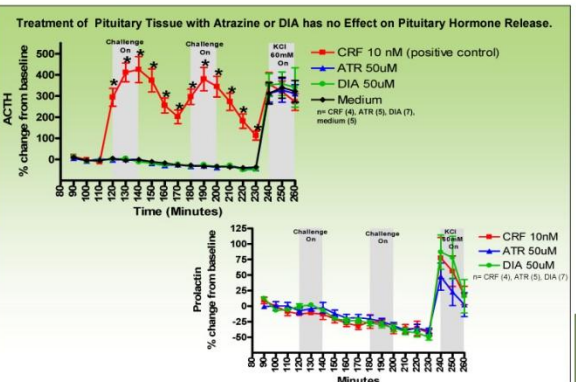
Radioimmunoassay. Serum and perfusion medium concentrations of corticosterone and progesterone were measured using coat-count radioimmunoassay kits obtained from Siemens (Los Angeles, CA). Plasma and perfusion medium concentrations of ACTH were measured using NACTH IntraChem Double Antibody Kit, MP Biomedicals, LLC.

Statistical analysis. All hormone data were analyzed using ANOVA (General Linear Models (GLM), Statistical Analysis System (SAS), SAS Institute, Inc.). When significant (p<0.05) differences were indicated by ANOVA, the Dunnett Multiple Comparison Test was used to compare each treatment with the control within a given time point. Repeated measure ANOVA was used for analyzing perfusion data. When significant (p<0.05) differences were indicated by repeated measures ANOVA, Tukey Post Hoc Tests were used to compare test doses at each time point. All perfusion data was calculated as percent change from average baseline value measured in RIA concentration/ml of sample/mg of tissue/ml.



References

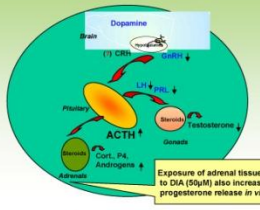
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Stevens et al., *J. Toxicol. Environ. Health*, 43:139-153(1994).
Eldridge et al., *J. Toxicol. Environ. Health*, 43:155-167 (1994); *Steroids* 64:672-678 (1999).
Cooper et al., *Reprod. Toxicol.*, 10:267-284 (1996); *Toxicol. Sci.* 53:297-307 (2000).



Laws et al., *Toxicol. Sci.*, 58(2):336-376 (2000); *Toxicol. Sci.* 76(1):61-68 (2003).
Stoker et al., *Toxicol. Sci.* 58(1):50-59(2000); *Toxicol. Sci.* 67(2):198-206 (2002).
Modic, 2004. Thesis, NC State University, <http://www.lib.ncsu.edu/theses/available/01-08052004-132003>

Goldman, J. M., and Cooper, R. L. (1993b). In *Methods in Toxicology, Vol III, Part B. Female Reproductive Toxicology*, pp. 16-33. Academic Press, Orlando, FL.

Effects of *In Vivo* Chlorotriazine exposure on the HPA and HPG axes



Conclusions

***In Vivo* Studies:**

- Dose-dependent increases in plasma ACTH, progesterone, and corticosterone were observed after a single oral dose of atrazine or DIA, with maximal increases in concentration observed at 15 or 30 minutes.
- HPA-axis activation was not observed after a single oral dose of DACT.

It is unlikely that this HPA-axis activation is due to an acute stress reaction to the dosing procedure, given that animals were pre-dosed with vehicle for one week prior to administration of the test chemicals, and animals dosed with vehicle only did not display elevations in any hormone measured.

***In Vitro* Studies:**

- Perfusion experiments were conducted to examine potential direct effects of atrazine and DIA on pituitary and adrenal tissues.

Pituitary Perfusion:

Neither atrazine (50µM) or its metabolite DIA (50µM) caused an increase in pituitary release of ACTH or prolactin at any time point.

Adrenal Perfusion:

Neither atrazine (50µM) or its metabolite DIA (50µM) caused an increase in adrenal release of corticosterone at any time point.

DIA (50µM) caused an increase in adrenal progesterone release (i.e. 220 and 230 min time points), while atrazine (50µM) did not alter adrenal progesterone release at any time point.

➤ These data suggest that the changes in ACTH and adrenal steroids observed following a single oral dose of atrazine or DIA *in vivo* may be due to a CNS effect, or in the case of DIA, may be a combination of CNS effects and a direct effect of the chemical on adrenal tissue.

➤ Further studies are needed to determine how DIA may be able to modify steroidogenesis, potentially through effects on enzyme synthesis or function in this pathway.



Acknowledgements

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Disclaimer: *(continued)*

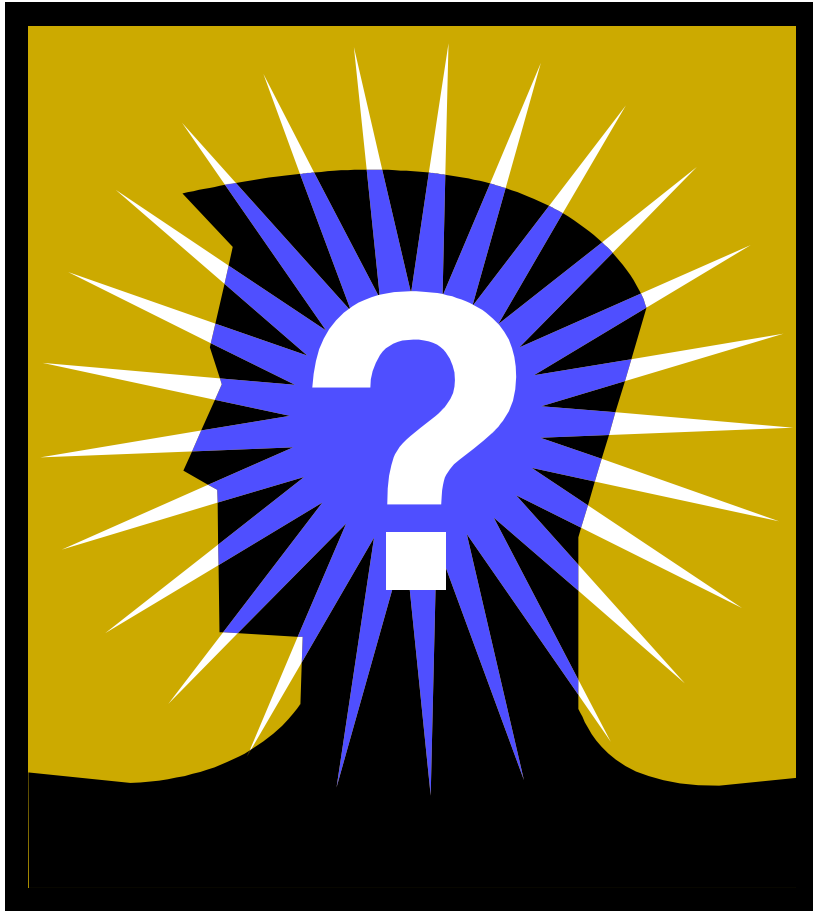
Any mention of a specific commercial product, Trade name or service is does not constitute an endorsement or recommendation for use.



Capintec CAPTUS 3000 Portable Thyroid Uptake System



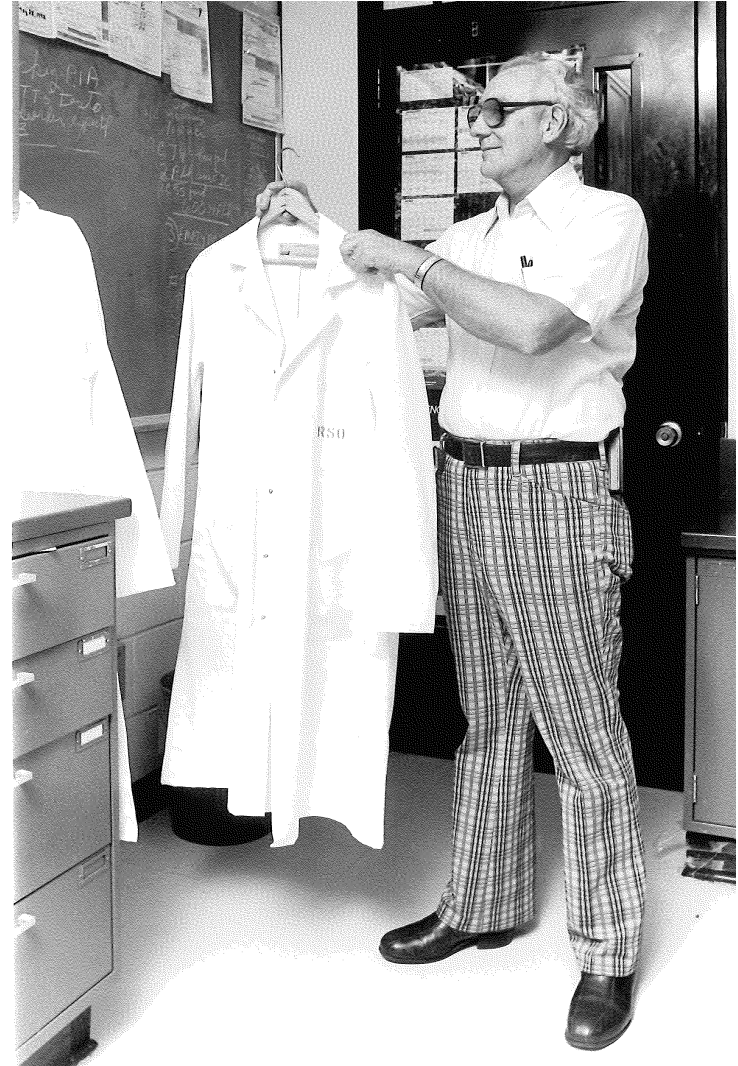
Statement of the Problem



While I still enjoy '80s music videos on VH1 Classic, using a system / method purchased in 1985 was no longer desirable.

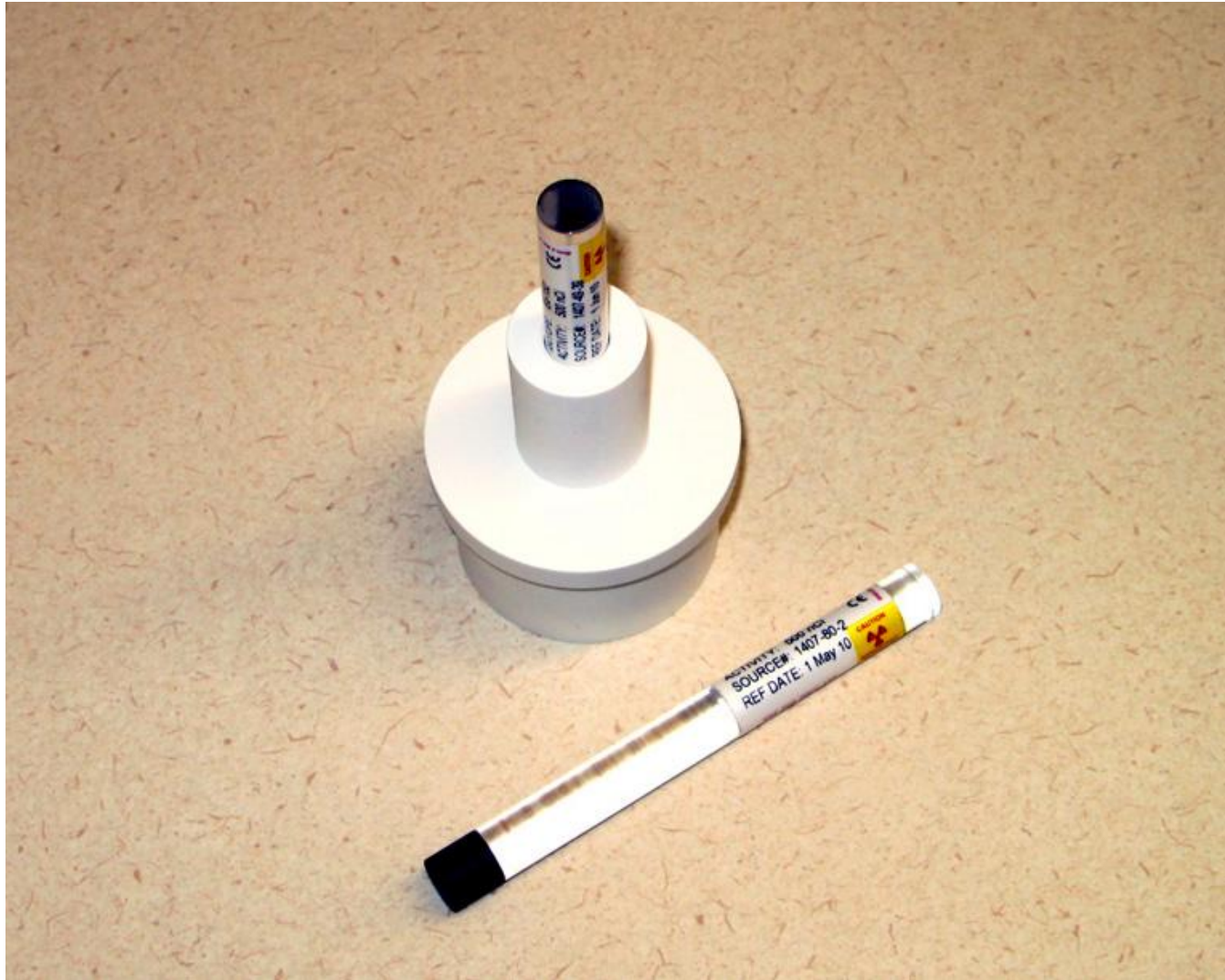
Graham Hair, EPA/RTP RSO 1973-1994

Agreements for bioassay measurements at other institutions like Duke University and NIEHS no longer feasible.



Capintec CAPTUS 3000 Portable Thyroid Uptake System











Objectives

- Review **Iodine-125** bioassay requirements
- Review **Iodine-125** detection options.
- Review techniques to determine **Iodine-125** efficiency.
- Discuss materials, methods and data observations.
- See the practical results of the calibration.

External Monitoring

- Whole Body (Badge) Monitoring

- Higher energy beta emitters; photon emitters

Examples – P-32, Cr-51, I-125, As-73

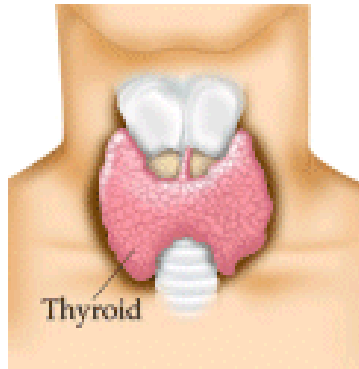
- Extremity (Ring Badge) Monitoring

- Higher energy beta emitters
- Examples – P-32, P-33

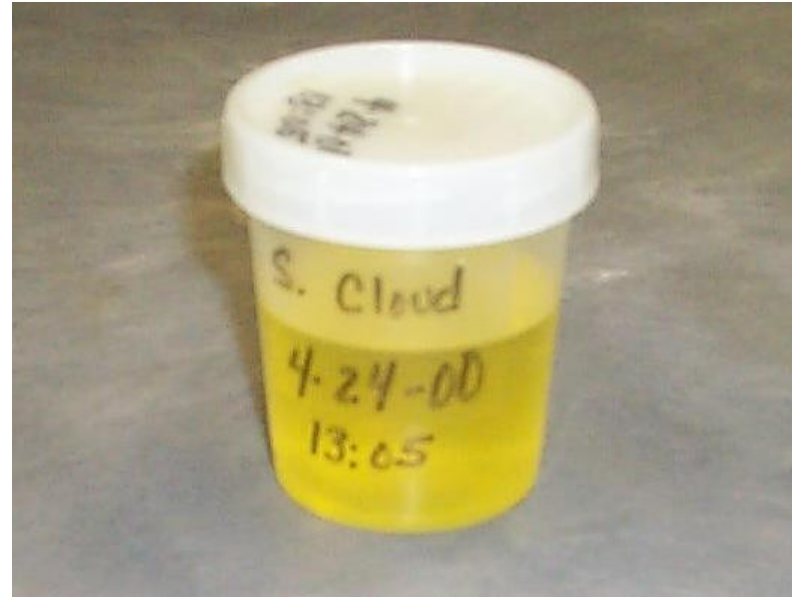


**Picture Credits: Landauer Corp.
ldrsolutions.landauerinc.com**

Internal Dose Assessment



**Thyroid Scan
for I-125 & I-131**



**Urine Analysis
for H-3, C-14, P-32, & S-35**

Picture Credit: (Thyroid gland)

http://www.uos.harvard.edu/ehs/radsafety/pur_i125.shtml

10 *CFR* 20 Limits on Intake

- Allowable Limit on Intake (ALI):
 - Inhalation: 40 μCi
 - Ingestion: 60 μCi
- Derived Air Concentration (DAC):
3 x 10⁻⁸ $\mu\text{Ci}/\text{mL}$

But what about thyroid burden?

- Follow-up required for > 0.12 μCi from Reg. Guide 8.20

Internal Exposure Monitoring

TABLE 1
Activity Levels Above Which Bioassay
for I-125 or I-131 Is Necessary

Isotope	Activity Limit** (mCi)	
	Volatile or Dispersible	Bound to Non-Volatile Agent
I-125	1.0	10

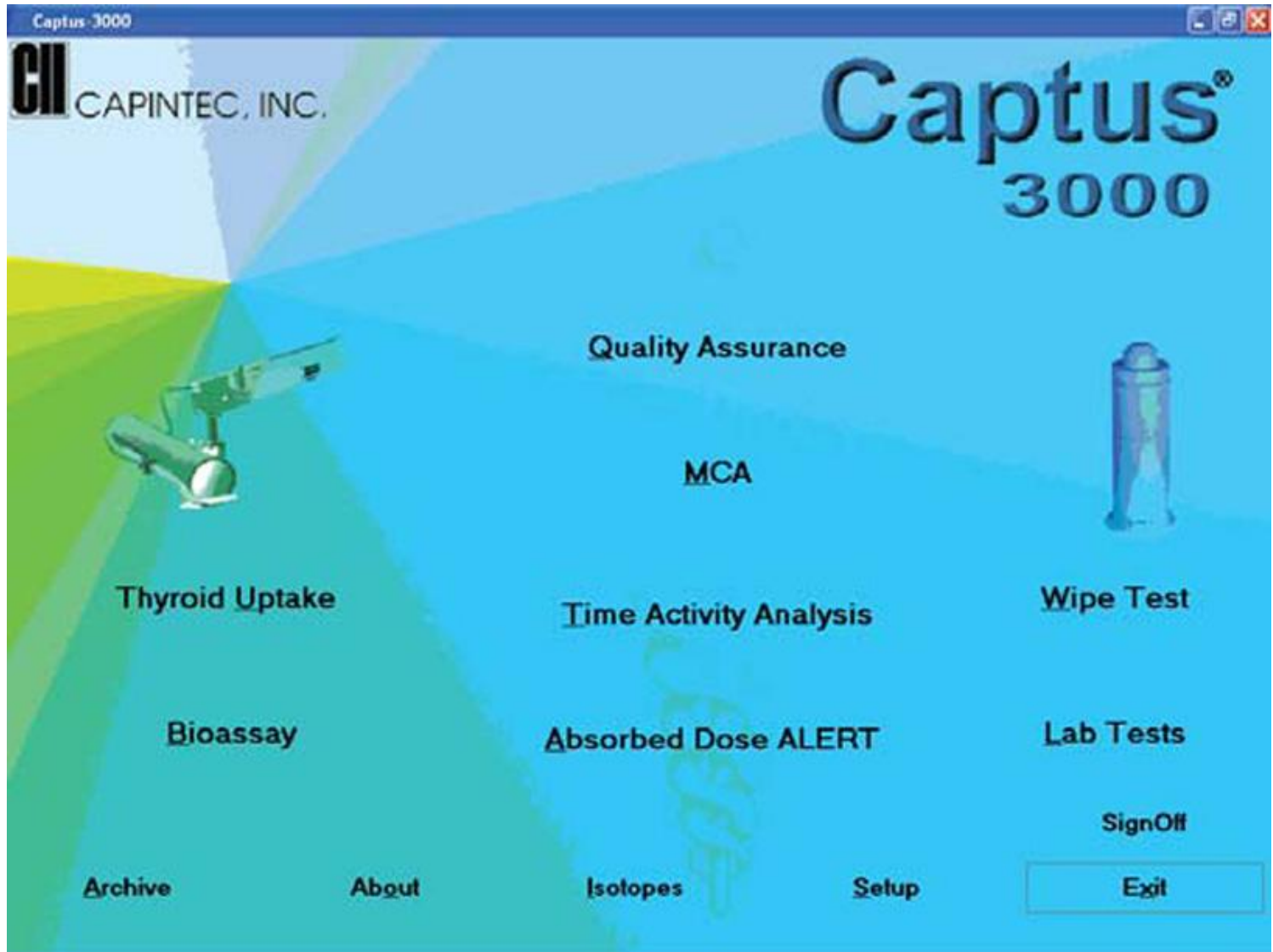
****Activity Limit applies to the larger quantity:
[1] Three Month usage/disposal or
[2] Maximum activity handled at one time.**

Reference: (Adapted)

**U.S. Nuclear Regulatory Commission Regulatory Guide 8.20,
“Applications of Bioassay for I-125 and I-131,” September, 1979.**

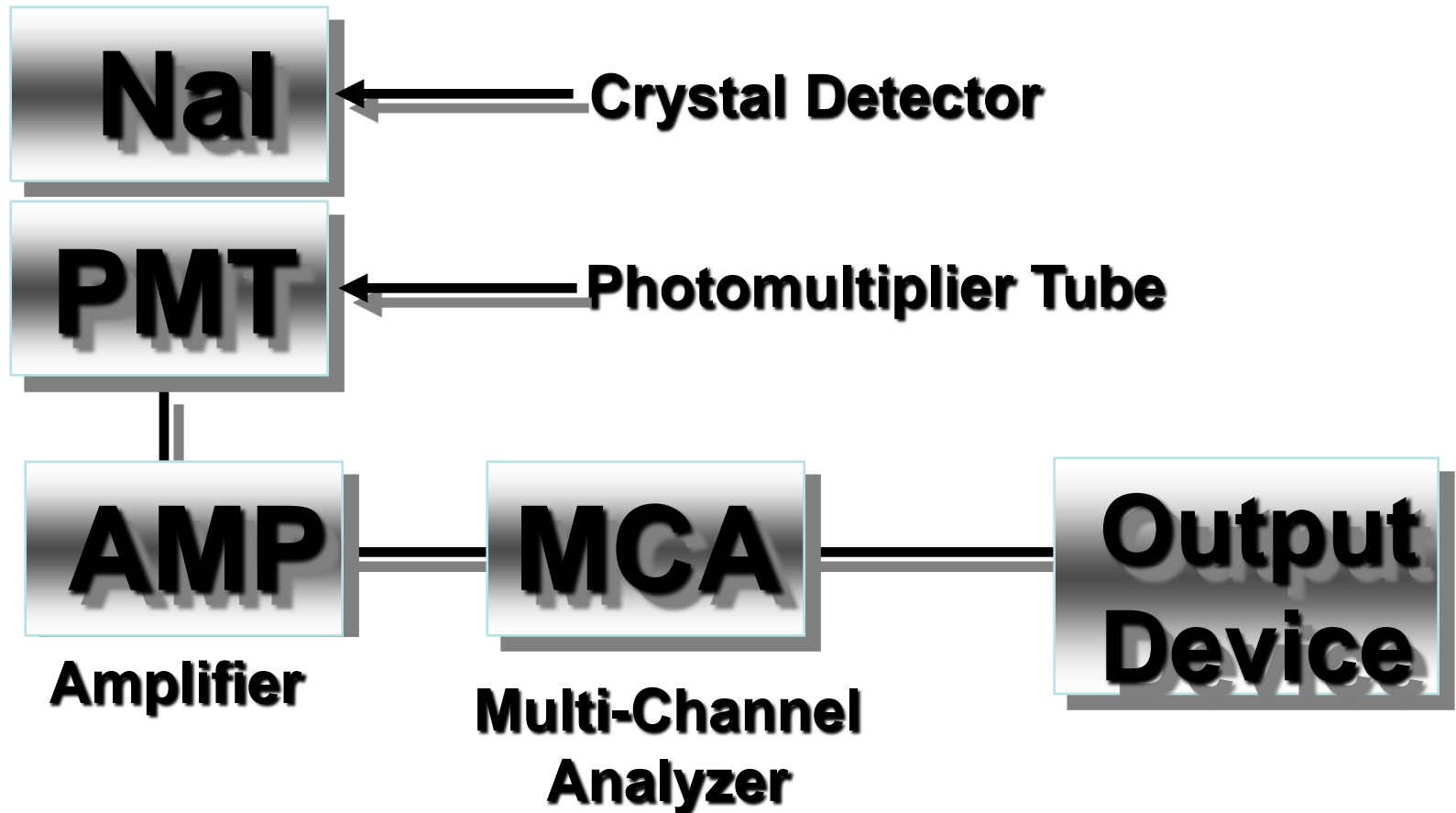


**Configured for Calibration /
Spectrum Acquisition**





NaI Scintillation Detector



Radiation Detection Scintillation Detectors

Incident Ionizing Radiation

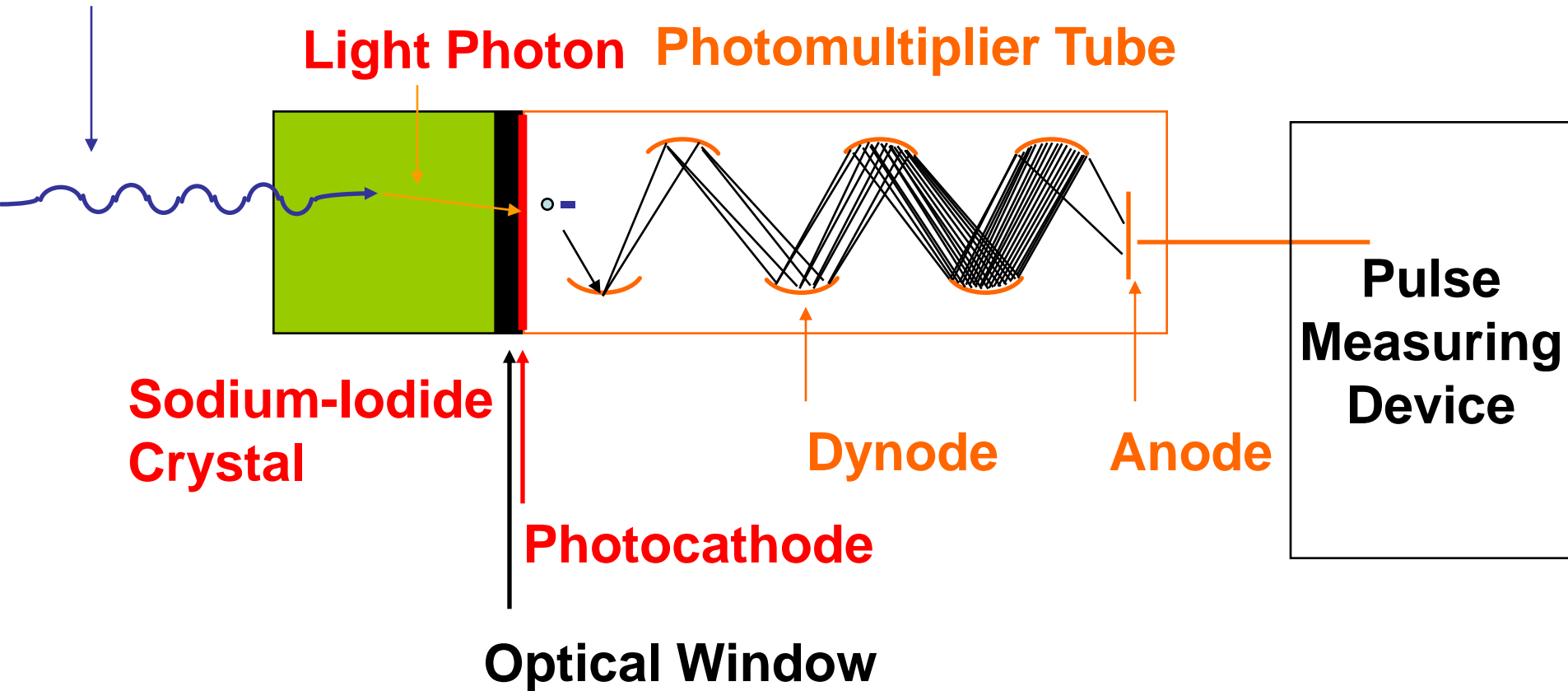


Diagram Credit:
HPS STW Technical
Presentation

Liquid Scintillation Counters (LSC)



Gamma Counters



Picture Credits:
(Various used equipment on-line sales listings.)

Modern Gamma Counters



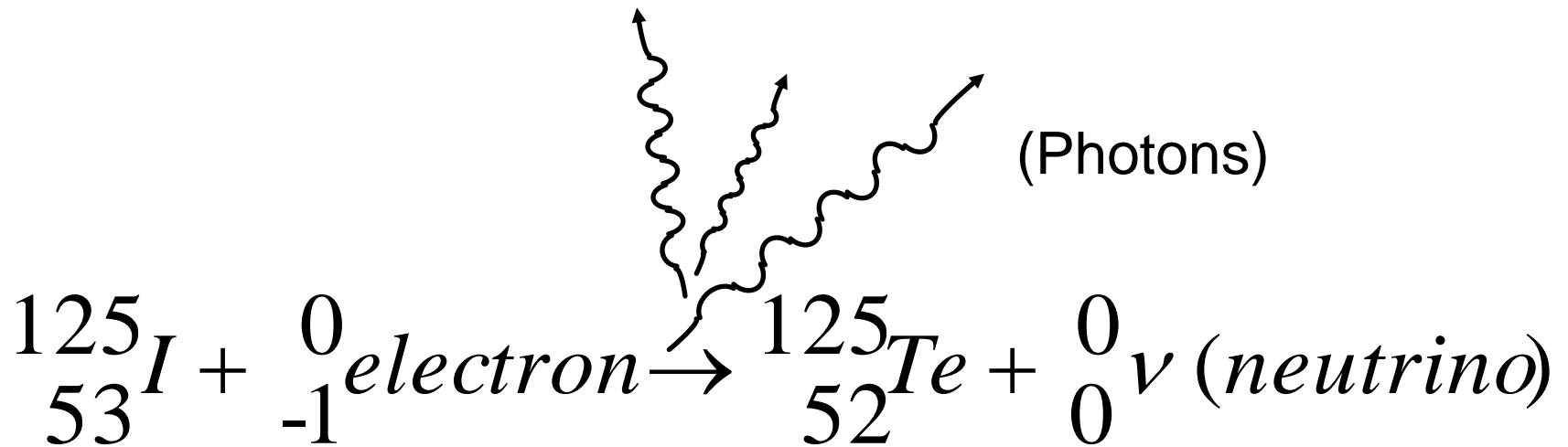
1470 WIZARD



1470 WIZARD

Picture Credit:
Product Brochure from PerkinElmer formerly Wallac

Iodine-125 Decays by Electron Capture



**Results in Multiple, Discrete
Photons Emitted**

Primary Photons for I-125

Photon Mode	KeV	% Abundance
Te K alpha 2	27.2	40.5
Te K alpha 1	27.5	75.6
Te K beta 1	31.0	20.1
Te K beta 2	31.9	4.4
γ M1+0.07%E2	35.5	6.7

Reference:

Brown, E and Firestone, R.; Table of Radioactive Isotopes, 1986.

Primary Photons for I-129

Photon Mode	KeV	% Abundance
Xe K alpha 2	29.5	20.5
Xe K alpha 1	29.8	37.8
Xe K beta 1	33.6	10.2
Xe K beta 2	34.6	2.4
γ M1+ 0.08%E2	39.6	7.5

Reference:

Brown, E and Firestone, R.; Table of Radioactive Isotopes, 1986.

Ratio of Photons

Isotope	Photon Abundance (Per decay)
Iodine-129	0.785
Iodine-125	1.473

Equivalency or Conversion Factor:

- Going from I-129 activity to I-125 activity divide by 0.533.

Reference:

Brown, E and Firestone, R.; Table of Radioactive Isotopes, 1986.

Modern Gamma Counters



1470 WIZARD



1470 WIZARD

Picture Credit:
Product Brochure from PerkinElmer formerly Wallac

Absolute Efficiency?

- “Absolute detector efficiency—for I-125, it is possible to determine the absolute efficiency of the detector by using the Horrocks method. The method does not require calibrated sources (having a known DPM value). For other isotopes, detector efficiency is obtained by dividing corrected CPM in the counting window by the isotope DPM value. This method does require the use of calibrated sources.”

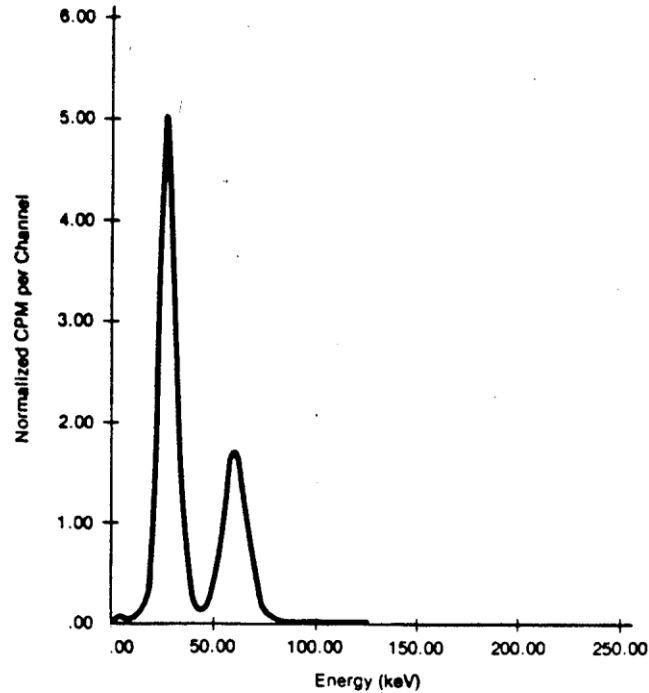
From Perkin-Elmer Website

las.perkinelmer.com/Catalog/ProductInfoPage.htm?ProductID=2470-0010

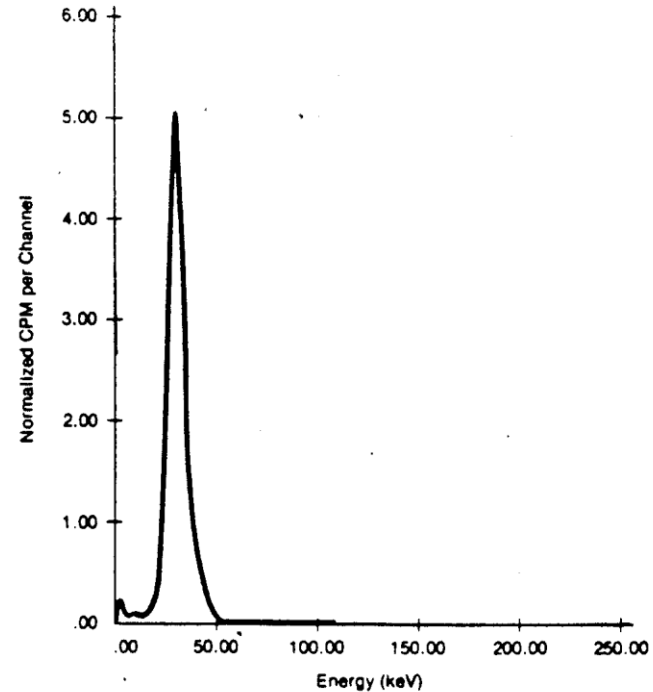
References:

1. Eldrige, *Nucleonics*, 22 (6), 56 (1964)
2. Harper et. al, *J. Nucl. Med.* 4, 277 (1963)
3. Horrocks, *Clinical Chemistry*, Vol. 21, No. 3, (1975)

I-125 and I-129 Gamma Spectra



IODINE-125
(HALF-LIFE = 1,440 HOURS)



IODINE-129

Reference: (Scanned page)

Packard AutoGamma 5000 Series Manual; Appd. B-13; 1989.

Data/Observations

Iodine-129

- 0.23% Iodine-129 Efficiency

Iodine-125

- 0.43% Iodine-125 Efficiency
 - Minimum Detectable Activity of 0.06 μCi
- Fewer counts in the sum (coincident) peak than with bench top gamma counting instruments.

Future Work

- Acquire Iodine-125 spectrum under conditions of significant dead time.
- Validate with NIST traceable standard of Iodine-125
- Cross calibrate NIST Traceable Iodine-125 with NIST Iodine-129 traceable standard.
- Check on the derivation of the Reg. Guide Organ Burden.



Reference Materials

1. Cember, Herman: *An Introduction to Health Physics*; 2nd edition, 1987.
2. U.S. Nuclear Regulatory Commission Regulatory Guide 8.20, “Applications of Bioassay for I-125 and I-131,” September, 1979.
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4. Packard Model Autogamma 5000 Operating Procedures Manual, [Operating Procedures and Appendixes.

Reference Materials

5. PerkinElmer Model 1470 Wizard Gamma Counter Instrument Manual, [Operating Procedures and Appendixes.]
6. Brown, E. and Firestone, R.; **Table of Radioactive Isotopes**, 1986.
7. Capintec CAPTUS 3000 Portable THYROID UPTAKE SYSTEM, [Owner's Manual]; Rev. D – February 2010.

Physics Before Chemistry

“Physics is the knife that cuts through the grain of nature.”

Unattributed-Unknown



Contact Information:

Todd Baker

baker.todd@epa.gov

Voice: 919-541-4307

