



Cytek[®] Aurora User's Guide



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FCC Information

WARNING: Changes or modifications to this unit not expressly approved by the party responsible for compliance could void the user's authority to operate the equipment.

NOTICE: This equipment has been tested and found to comply with the limits for a Class A digital device, pursuant to Part 15 of the FCC Rules. These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. This equipment generates, uses, and can radiate radio frequency energy and, if not installed and used in accordance with the instruction manual, can cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause harmful interference in which case the user will be required to correct the interference at his or her own expense.

Shielded cables must be used with this unit to ensure compliance with the Class A FCC limits.

This Class A digital apparatus meets all requirements of the Canadian Interference-Causing Equipment Regulations.

Cet appareil numérique de la classe A respecte toutes les exigences du Règlement sur le matériel brouilleur du Canada.

CDRH Information

This equipment complies with CDRH Class I requirements.

Regulatory Information

For Research Use Only. Not for use in diagnostic or therapeutic procedures.

History

Revision	Date	Change
52-70001-0A	10/2017	Initial release
52-70001-0B	1/2018	Added details to safety, acquisition, and unmixing
52-70001-0C	1/2019	Added Loader, yellow-green laser, keywords, SSC off blue laser, label and lot-specific unmixing, and group-specific unstained controls
52-70001-0D	3/2019	Added ultraviolet laser
N9-20006 Rev. A	6/2019	Updated SpectroFlo QC beads lot number, corrected laser bandwidth table, changed user's guide part number
N9-20006 Rev. B	10/2019	Updated to SpectroFlo v2.2 to include reference control QC (benchmarks), clog and bubble detection

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Introduction





About this Guide

This guide provides information on the Aurora flow cytometer, daily workflow, SpectroFlo® software features, instrument maintenance, and cytometer specifications. It also includes troubleshooting tips and service information.

Safety

Safety Symbols

The Aurora is intended for research use only; not for diagnostic or therapeutic procedures. The following table lists symbols used throughout this guide.

Symbol	Meaning
	Caution: hazard or unsafe practice that could result in material damage, data loss, minor or severe injury, or death
	Risk of electric shock
	Biological risk
	Laser radiation

General Safety

- Read all safety instructions completely before using the equipment. Keep the instructions in a safe place.
- Follow all instructions when operating the instrument.
- If the equipment is used in a manner not specified by the manufacturer, the protection provided by the equipment may be impaired.
- Do not place any object on top of the instrument.

- Do not block any ventilation openings.
- Do not place the unit near any heat sources such as radiators, heat registers, stoves, or other devices (including amplifiers) that produce heat.
- Use only attachments/accessories specified by the manufacturer.
- Use only with the cart, stand, tripod, bracket, or table specified by the manufacturer or sold with the equipment. When a cart is used, use caution when moving the cart/equipment combination to avoid injury from tipping over.
- Unplug the instrument when it will not be used for long periods of time.
- Refer all servicing to qualified service personnel. Service is required when the unit has been damaged in any way, such as: if the power-supply cord or plug is damaged, if the unit is dropped, if liquid is spilled onto the unit or objects fall into the unit, if the unit is exposed to rain or moisture, or if the unit does not operate properly.
- Do not expose the instrument to temperatures outside the range of 17°C to 28°C.
- Concentrations of sodium hypochlorite (bleach) higher than 10%, as well as other cleaning agents, can damage the instrument. Use a 10% solution of household bleach to clean, where indicated. A 10% bleach solution is prepared by adding 1 part household bleach to 9 parts deionized water.
 - **NOTE:** Household bleach contains 5–7% sodium hypochlorite.
- Before turning on the cytometer, visually inspect all containers. Wear the recommended protective laboratory attire such as protective gloves, eyewear, and lab coat.
- Purge the sheath filter if air bubbles are visible in the sheath filter, or if the plenum or sheath container have run dry.
- Fill the sheath container as needed. Never use tap water as sheath solution. Never use surfactant-based sheath solutions.
- Do not run bleach or detergent through the sheath filter. It is difficult to remove cleaning solutions from the sheath filter.
- Check the cytometer periodically for fluid leaks or crimped lines. If evidence of a leak is detected, contact Cytek Technical Support immediately. Do not attempt to repair the instrument.
- When performing Daily QC, always select the current bead lot number.

Electrical Safety

- Do not place liquids on top of the instrument. Any spill into the ventilation openings could cause electrical shock or damage to the instrument.
- Do not use this equipment near water.



WARNING: To reduce the risk of fire or electric shock, do not expose this apparatus to rain or moisture.

- Use only the power supply cord specified by the manufacturer. The power cord of the unit is equipped with a 10A three-prong power plug. Do not remove the ground pin of the power plug under any circumstances. Make sure the plug is securely plugged into the power outlet to prevent fire. If the power supply cord needs to be replaced, the cross section area of the conductor should be at least 16 AWG. This is to prevent electric fire or shock.
- Use only the fuse specified by the manufacturer. The fuse is 250 VAC, 5 A, size 5 x 20 mm.

- Protect the power cord from being tread upon or pinched, particularly at the plug and the point where it emerges from the equipment. Be sure that the power outlet is located near the equipment so that it is easily accessible.

Biological Safety

- Empty the waste container when filling the sheath container or as needed to prevent leakage. Take care to avoid damaging the fluid level sensor in the waste tank.
- Biological samples are potentially dangerous and/or life threatening. Adhere to proper handling procedures for samples and reagents. Wear appropriate laboratory attire such as protective gloves, eyewear, and lab coat.
- Any instrument surface in contact with biological specimens can transmit potentially fatal disease. Use universal precautions when cleaning the instrument or replacing parts.

Laser Safety

The Aurora is a Class 1 Laser Product and complies with the US FDA Center for Devices and Radiological Health 21 CFR 1040.10 and 1040.11 except for deviations pursuant to Laser Notice No. 50, dated June 24, 2007. No laser radiation is accessible to the user during normal instrument operation.

The lasers are fully contained within the instrument and do not require any special work area safety except when service procedures are being performed. These procedures are performed only by Cytek service personnel.



CAUTION: Modification or removal of the optics covers or laser shielding could result in exposure to hazardous laser radiation. To prevent damage to skin and eyes, do not remove the optics covers or laser shielding, or attempt to service the instrument where laser warning labels are attached.

Use of controls or adjustments or performance of procedures other than those specified in this user's guide may result in hazardous radiation exposure.

To prevent exposure to laser radiation:

- Do not defeat any safety interlocks on the instrument.
- Do not use controls, make adjustments, or perform procedures other than those specified in this user's guide.
- Do not attempt to perform service procedures on the lasers.

Technical Support

For instrument support within the US, call 1-877-92-CYTEK. Outside the US, call 1-510-657-0102.

Visit our website, www.cytelbio.com, for up-to-date contact information.

When contacting Cytek, have the following information available:

- Serial number
- Any error messages
- Details or screen shots of recent system performance
- SpectroFlo software version and system firmware version, located in the Help module

Overview

Aurora System

The Aurora system consists of the Aurora flow cytometer and a computer workstation running SpectroFlo software for acquisition and analysis. SpectroFlo® QC beads are also included. The system is intended to be used for analyzing cells in the fields of immunology, biochemistry, biology, oncology, and hematology research.

The cytometer is an air-cooled, compact benchtop instrument. It is equipped with up to five lasers, 64 detection channels for fluorescence, and three channels for scatter (blue laser FSC, blue laser SSC, and violet laser SSC). Sheath and waste fluids are contained in either 4-L tanks, included with the system, or 20-L cubitainers. Software indicators notify you when the sheath is getting low or the waste is getting full. The fluidics system includes a plenum for storing sheath, allowing you to fill the sheath tank during operation.

An Automatic Micro-Sampling System (AMS) is an optional feature available to automate sample delivery. The system, referred to throughout this guide as the Loader, holds one 96-well standard-height plate.

The workstation is a dedicated USB-compatible PC with monitor, keyboard, and mouse. It runs Microsoft® Windows® 10 Pro with a 64-bit operating system, which is required for SpectroFlo software.



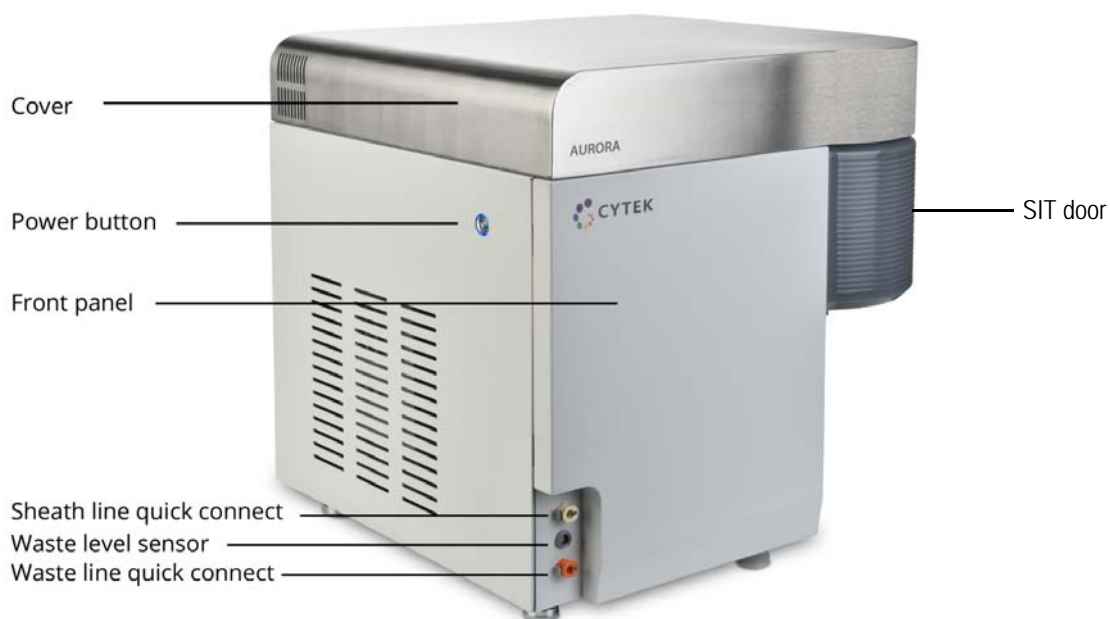
Cytometer Overview

Solid-state lasers transmit light through a flow cell where particles in suspension are focused, single file for interrogation by the laser. Proprietary, high-sensitivity, semiconductor detector APD arrays are equipped with up to 16 channels per laser to capture the emission spectra of dyes that emit in the 365 to 829-nm wavelength range. The resulting fluorescence and scatter signals are then collected and converted into electronic signals. On-board electronics convert these signals into digital data that can be acquired and recorded on the workstation.

The cytometer power button is located on the left side of the upper panel of the cytometer. When the cytometer is powered on, the power button is illuminated.

The front panel opens on hinges to the left to reveal the fluidics system. The SIT door to the right of the front panel opens to reveal the sample injection tube (SIT) assembly. The top cover opens to reveal the optics plate.

Front of Cytometer



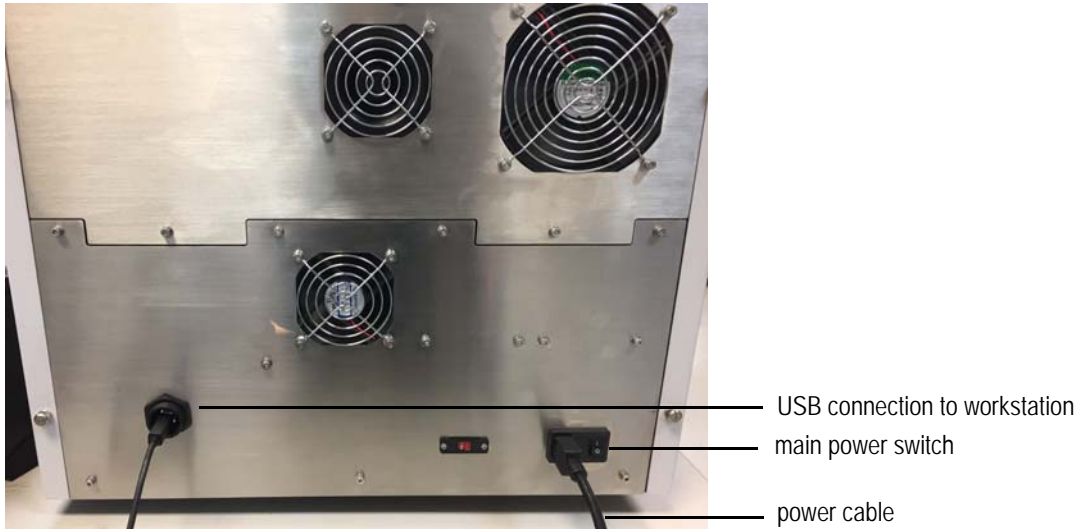
Do not place any object on top of the instrument.



Do not place liquids on top of the instrument. Fluid leaking into the cytometer could cause electrical shock or damage to the instrument.

Back of Cytometer

Allow 12.7 cm (5 in) between the back of the cytometer and the wall for proper ventilation.

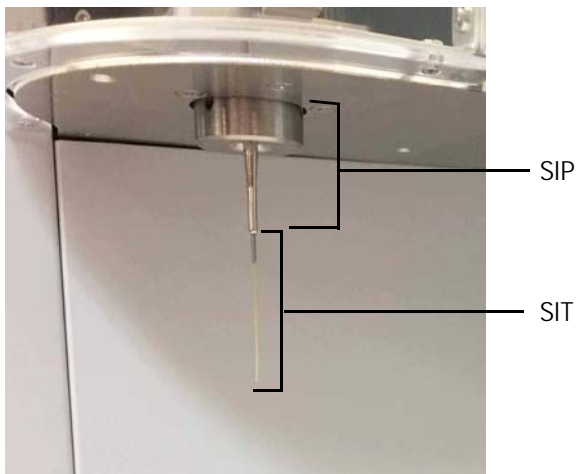


Fluidics System

Sample Injection Port and Sample Injection Tube

Sample, contained in a standard 12 x 75-mm tube, enters the cytometer through the sample injection tube (SIT) that is contained within the sample injection port (SIP). The 12 x 75-mm sample tube snaps into place under the SIP requiring no additional tube retention support. The SIT extends from the SIP during acquisition and retracts when the cytometer is not acquiring.

When using the optional Loader, a 96-well plate is used for sample delivery in place of 12 x 75-mm sample tubes. See the Loader chapter starting on [page 93](#) for more information.



Fluid Containers

The Aurora draws sheath solution directly from a 20-L sheath cubitainer or the 4-L sheath tank provided by Cytek. It expels waste into an empty 20-L cubitainer or the 4-L waste tank provided by Cytek.

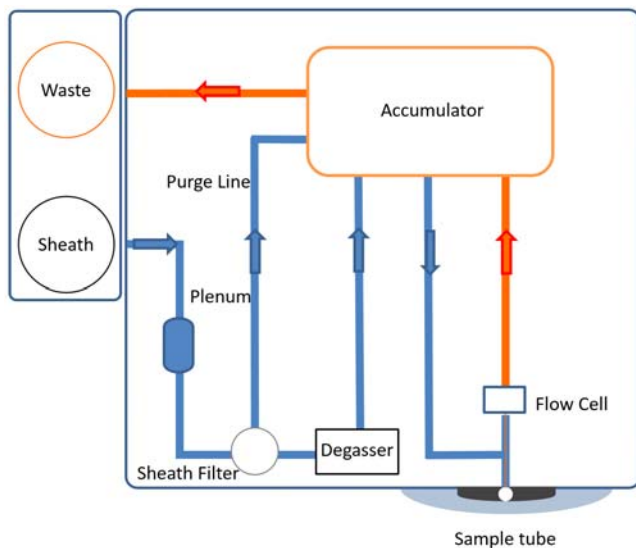
The fluidics tanks are contained in a holding reservoir located on the left side of the cytometer. The 4-L tank with the transparent fluidic line is for sheath solution. The 4-L tank with the orange fluidic line is for waste.



Fluid Flow

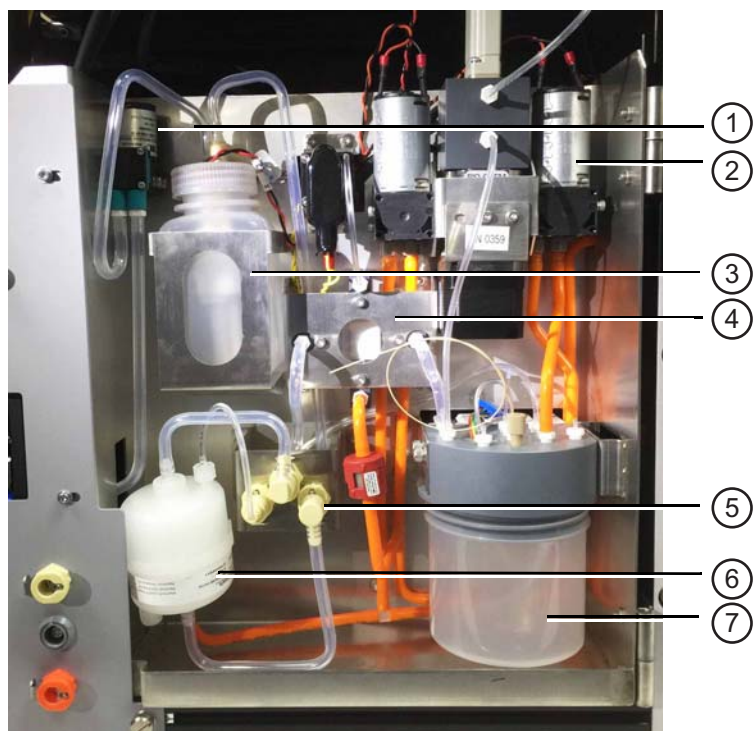
The Aurora fluidics are driven by vacuum. An accumulator vessel is the source of vacuum for the system. Sheath solution is drawn into and stored in the sheath plenum before passing through a sheath filter, where debris and contaminants are removed. Before reaching the flow cell, the sheath stream passes through a degasser, which removes air bubbles. After passing the laser interrogation point, the combination of sheath solution and sample travels to the waste container.

Sheath and waste fluid levels are monitored by sensors. The waste level sensor is located underneath the waste tank cap. The sheath level sensor is located underneath the sheath plenum cap. Both sensors are monitored by the software.



Fluidics Components

The following figure shows the fluidics components.



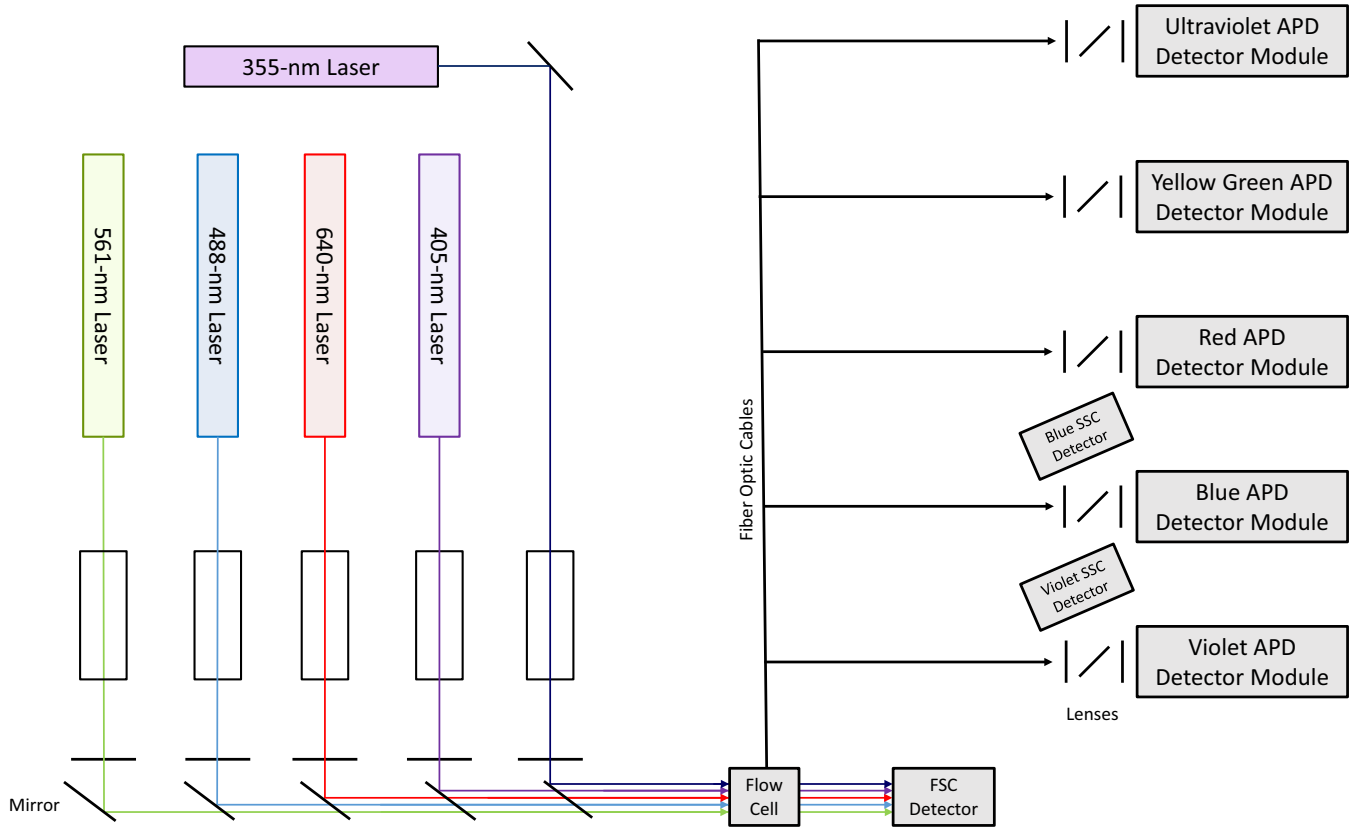
The following table describes the fluidics components.

No.	Component	Description
1	Plenum pump	Pulls sheath from the sheath tank to fill the plenum
2	Vacuum pump	Maintains the vacuum in the accumulator
3	Plenum	Storage vessel for sheath fluid before it flows to the sheath filter
4	Degasser	Removes air bubbles from the sheath fluid
5	Sheath filter quick connects (x3)	Sheath filter fluid input, fluid output, and vent line quick-connects
6	Sheath filter	Filters debris and particles from the sheath fluid
7	Accumulator	Vacuum source for the fluidics system

Optics

Unlike conventional flow cytometers that direct specific bandwidths of fluorescence light into discrete detectors or photomultiplier tubes (PMTs), the Aurora uses a solid-state, multi-channel, narrow-beam detector array for each laser. Each array can be configured with up to 16 detectors that are used to capture a part of the emission spectrum from each particle passing through the laser beam. The detector channels from up to five lasers are used to capture the entire emission spectra from each fluorescent-labeled particle. Spectral deconvolution (unmixing) algorithms calculate the contribution of the known individual fluorophore's spectra to the total collected signal.

For excitation, a proprietary flat-top laser design enables a constant power distribution across the width of the sample core stream.



Optical configurations are as follows:

Laser	Excitation	Channels for detection	Detector names
Ultraviolet	355 nm	16	UV1-UV16
Violet	405 nm	16	V1-V16
Blue	488 nm	14	B1-B14
Yellow Green	561 nm	10	YG1-YG10
Red	640 nm	8	R1-R8

■ **NOTE:** Different laser configurations are available. Yellow Green and Ultraviolet lasers are not available on the Northern Lights cytometers.

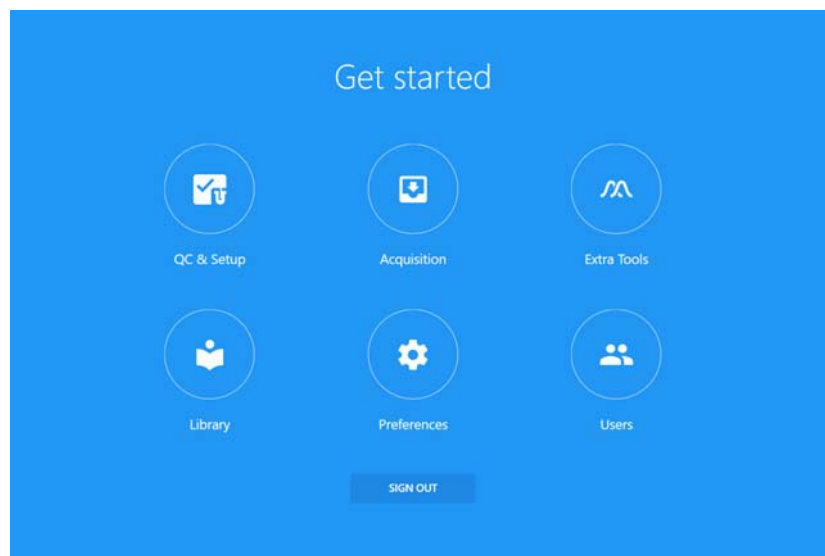
The wavelengths detected by each detector (channel) increase across the array. See the table on [page 71](#) for details.

Software Overview

SpectroFlo software allows you to acquire and analyze samples and adjust instrument settings. Once you log into the software, a Get started menu appears.

Get Started Menu

The Get started menu provides six modules that allow you to perform various functions. These same six modules are also accessible across the upper-right corner of each module screen.



The following table describes options in the Get started menu.

Module	Description
QC & Setup	Daily QC ensures that the instrument is in optimal condition for use. Run SpectroFlo QC Beads daily to assess system performance and to adjust settings to account for day-to-day variation. Levey-Jennings reports keep track of trends in system performance. Setup allows you to create reference controls. See “QC & Setup” on page 27 for information.
Acquisition	The Acquisition module allows you to create experiments to acquire and analyze data. Experiments can be created through a guided wizard or created from previously saved templates. See “Acquisition” on page 41 for information.
Extra Tools	Here, FCS files can either be unmixed or compensated using virtual filters. See “Unmixing and Compensation” on page 57 for information.
Library	The library allows you to store experiment templates, worksheet templates, user settings, fluorescent tags, SpectroFlo QC bead information, label information, keywords, and Loader settings. See “Library” on page 75 for information.
Preferences	Software preferences can be changed to customize the software. Default plot sizes, fonts, gate colors, print layouts, statistics box table option, and more can all be changed in the Preferences. See “Preferences” on page 81 for information.
Users	The Users module contains user management options and administrative controls. See “Users” on page 90 for information.

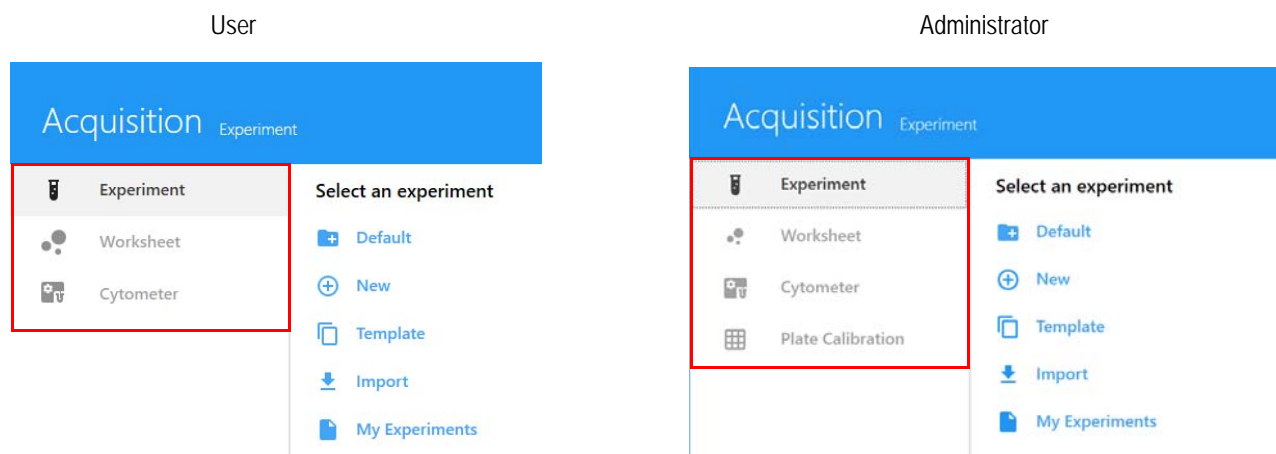
About Experiments

The Acquisition module provides the tools necessary to acquire data, such as the acquisition controls used to start, stop, and record data, and the instrument controls used to set the threshold and adjust the detector gains. See “[Experiment Display](#)” on page 42 for more information on these controls. Experiments contain the fluorescent tags and labels used in the experiment, the stopping criteria, and the groups of tubes/wells run, which can include the reference control group. You can create groups for your samples, if you wish, to conveniently organize samples by type or staining panels, for example.

Opening an Experiment

When you click Acquisition from the Get started menu, the Acquisition Experiment menu (below) is displayed, allowing you to open a default or template experiment, create a new experiment, or import an experiment. A wizard walks you through the steps to create a new experiment.

■ **NOTE:** By default the Acquisition menu in the left pane is collapsed, showing only the icons for Experiment, Worksheet, Cytometer, and Plate Calibration. To expand the menu to show the labels, click the arrows (>>) at the bottom of the pane.



Experiments can be created using several different methods. The following table describes the options in the Acquisition Experiment menu:

Method	Description
Default	Opens a new experiment with one group containing one tube and a set of labels and fluorescent tags in a default experiment worksheet template. The default experiment is user configurable. It is the quickest way to begin sample acquisition.
New	Opens the New Experiment Wizard to guide you through creating an experiment.
Template	Allows you to select from a list of saved experiment templates (see page 19).
Import	Imports an experiment ZIP file that was exported.
My Experiments	Allows you to select from a list of saved experiments. NOTE: Original experiments can be duplicated without data, which is equivalent to opening an experiment template. Right-click an experiment in My Experiments and select Duplicate (with or without data).

Completed experiments can be accessed through the My Experiments option in the Acquisition Experiment menu. Use the column headers to sort the list of experiments. For every tube recorded, two FCS files are saved, one raw and one unmixed. Use My Experiments to open experiments you already ran, as you may want to review the data or acquire more samples. You can also export experiments from My Experiments (below). A ZIP file is exported, containing all the raw data files, if applicable (and unmixed files for unmixed experiments), as well as the worksheet templates and experiment template.

Experiment	Created By	Date Created	Date Modified
Experiment_005	Admin	August 08, 2018 - 17:00 PM	August 08, 2018 - 17:00 PM
Experiment_004	Admin	August 08, 2018 - 14:47 PM	August 08, 2018 - 14:47 PM
Experiment_003	Admin	August 08, 2018 - 13:31 PM	August 08, 2018 - 13:31 PM

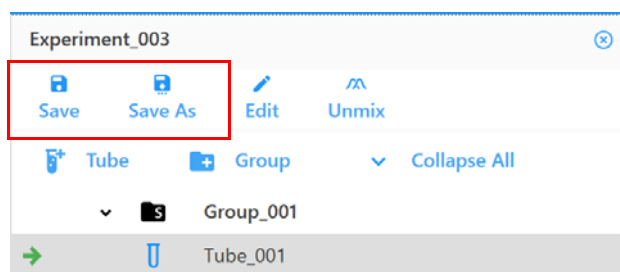
FCS Files

FCS files generated from an experiment are stored in the Export folder by default, or the folder you set as the default. See [“Storage Preferences” on page 88](#) for information. Experiments can contain the following types of FCS data files for each tube run:

- raw data files only (for samples that were acquired in an experiment)
- raw data files + unmixed data files (for samples that were acquired and unmixed live during acquisition)
- unmixed data files only (for samples that were unmixed post acquisition)

Experiment Templates

Use the Save As option above the experiment’s tube/group (hierarchy) list to save the current experiment as a template, which can then be used for running similar experiments. Experiment templates include fluorescent tags used in the experiment, reference controls, groups/tubes, labels, worksheets, and stopping criteria. Templates are saved in the library. To open and use a template, select Template from the Acquisition Experiment menu. See [“Experiment Templates” on page 78](#) for more information on experiment templates.



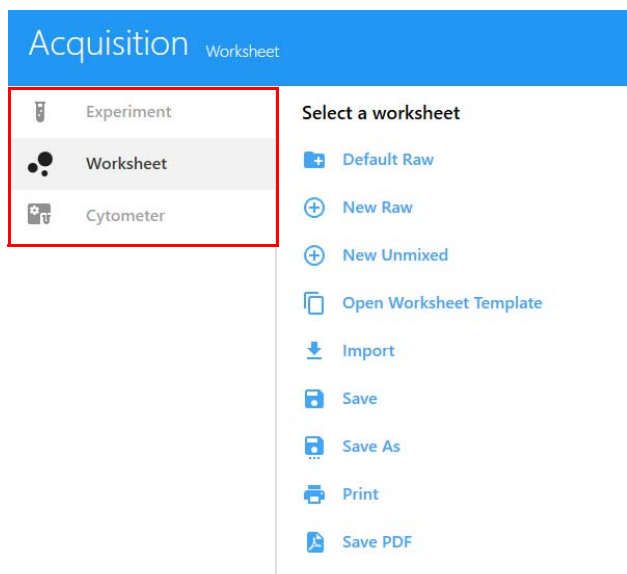
About Worksheets

Worksheets are used to visualize the data in the experiment. Each experiment requires at least one worksheet. A worksheet allows you to view the data in plots during acquisition, as well as perform analysis functions. Worksheets contain the tools necessary to create plots, gates, annotations, statistics, and the population hierarchy. Worksheets are saved with the experiment and can be saved separately and reused across experiments.

Two types of worksheets are available—worksheets for raw data and worksheets for unmixed data. You must select the appropriate worksheet to view the corresponding type of data. When viewing raw data, the parameters on the plots in a raw worksheet reflect the channel names, for example, B1-A, R1-A, V1-A. When viewing unmixed data, the parameters on the plots in an unmixed worksheet reflect the fluorescent tags, for example, PerCP-A.

Opening a Worksheet

To select a worksheet, click Worksheet in the Acquisition menu. The Select a worksheet menu appears (below). You can open a new raw or new unmixed worksheet. These worksheets open with a single FSC vs SSC plot. Use the worksheet tool bar to add plots and other elements. All worksheets are saved as template files (WXML) and can be opened using the Open Worksheet Template option. You can also import worksheets that were exported, as well as save, print, and save a worksheet PDF.

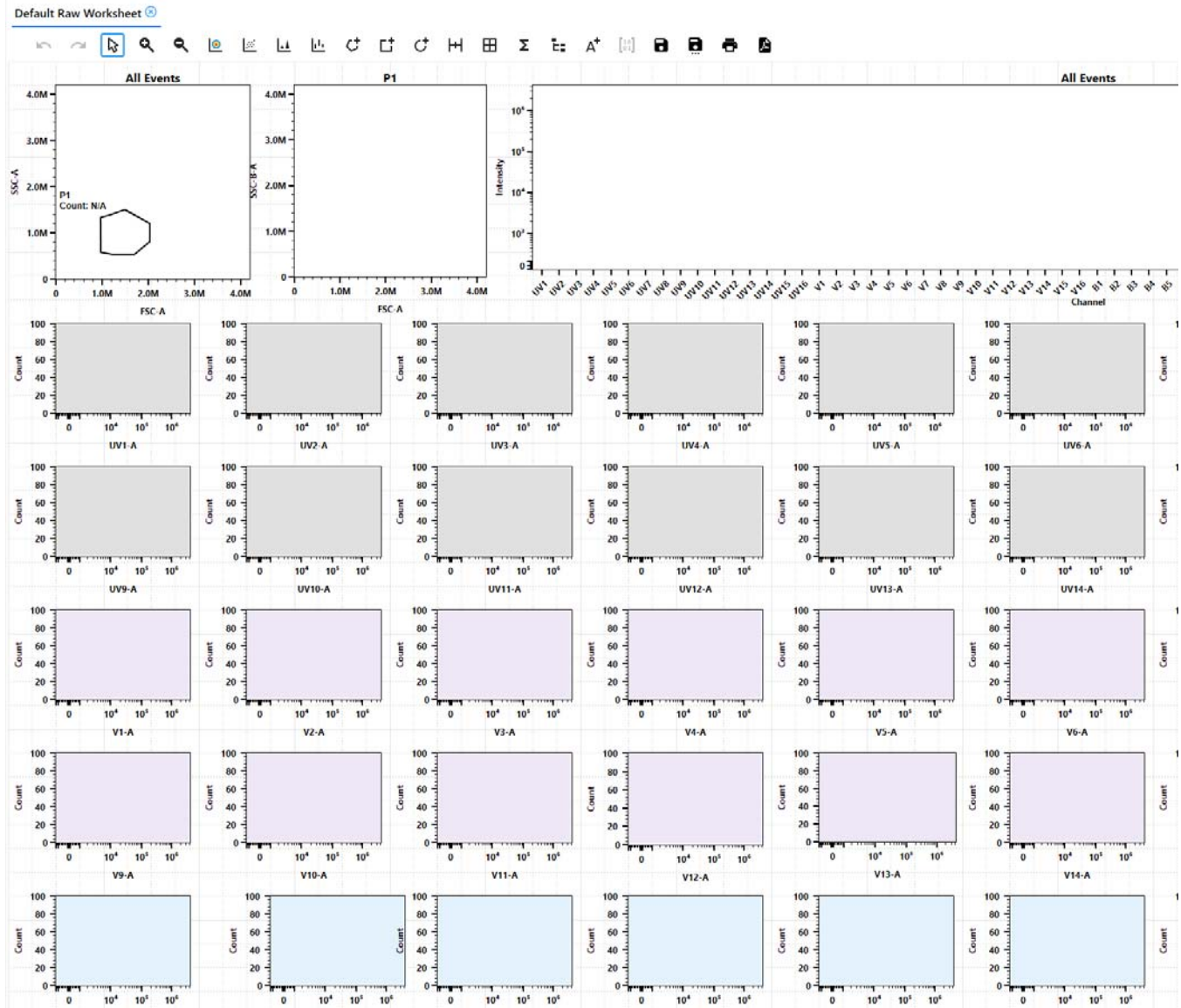


The following table describes the options in the Acquisition Worksheet menu:

Method	Description
Default Raw	Opens a default raw worksheet that can be used for experiments where reference controls will be acquired. Do not overwrite this worksheet. Always use Save As to save this worksheet with a new name.
New Raw	Opens a new raw worksheet.
New Unmixed	Opens a new unmixed worksheet.
Open Worksheet Template	Allows you to select from a list of saved worksheet templates. A default raw and default unmixed worksheet are provided.

Method	Description
Import	Imports a worksheet template that was exported.
Save, Save As, Print, Save PDF	Saves the worksheet, saves the worksheet with a new name, prints the worksheet, saves a PDF of the worksheet.

You can have multiple worksheets open at a time. The currently displayed worksheet appears with a blue line under the worksheet name. Because you can select different worksheets for different groups or tubes in an experiment, each tube will have a worksheet associated with it.



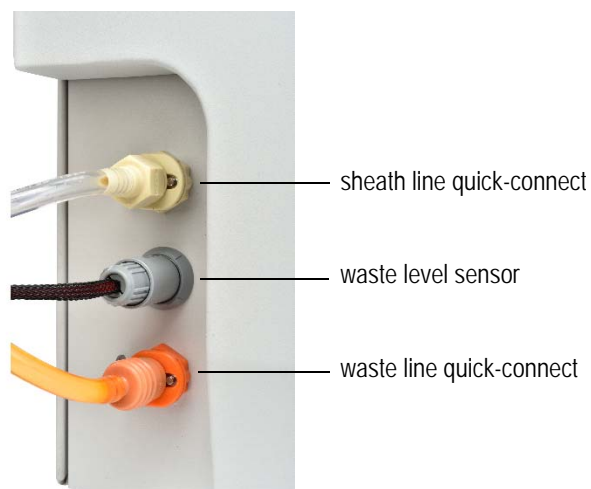
Each user can define a default raw worksheet and a default unmixed worksheet. Open the default worksheet and set it up for your experiment, then select Save As to save this worksheet with a new name. The worksheet will be available to select when you create an experiment. You can use this worksheet, open a template worksheet, or create a new worksheet.

All worksheets are saved in the library. See [“Worksheet Templates” on page 77](#) for more information on worksheet templates.

Startup & Shutdown

Filling the Sheath and Emptying the Waste

The color-coded sheath and waste quick-connects and the waste level sensor connector are located at the lower-left corner of the front panel.



■ **NOTE:** When using the Loader, place fluid tanks on the floor beneath the instrument, otherwise fluid can backflush into the wash well. We recommend using the 20-L cubitainers instead of the 4-L tanks for systems with a Loader. If your Loader has been upgraded to version 1.5, this is no longer necessary, as a check valve in the wash well fluidics line now prevents backflush from flooding the wash well.

Filling the Sheath

Fill the sheath container with manufacturer-provided sheath solution, MilliQ™ water, phosphate-buffered saline (PBS), or deionized (DI) water.

Sheath can be drawn from either the supplied 4-L sheath tank or directly from a 20-L cubitainer. Sheath solution can be added to the sheath container while the instrument is running.




Before turning on the cytometer, visually inspect all containers for leaks or cracks. Wear the recommended protective laboratory attire such as protective gloves, eyewear, and lab coat.



Fill the sheath container as needed. Use only the appropriate sheath solution. Never use tap water or surfactant-based sheath solution.

Filling Sheath into a Cytek 4-L Sheath Tank or a 20-L Cubitainer

- 1 Remove the sheath fluidics line cap from the cubitainer or sheath tank lid from the sheath tank.
- 2 Add the appropriate sheath solution.
- 3 Replace the fluidics line cap or sheath tank lid. Do not over-tighten.
- 4 If the cytometer is powered on and the software is connected, verify that the software sheath indicator is green.

 **Sheath**  **Waste**  **Connected**

Emptying the Waste

Waste can be expelled into either the supplied 4-L waste tank or directly into an empty 20-L cubitainer.



Empty the waste container when filling the sheath container or as needed to prevent leakage. The software indicator for waste will be yellow or red when the container needs to be emptied. Take care to avoid damaging the fluid level sensor in the waste tank.



Biological samples are potentially dangerous and/or life threatening. Adhere to proper handling procedures for samples and reagents. Wear appropriate laboratory attire such as protective gloves, eyewear, and lab coat during this procedure.



Always treat the contents of the waste container with household bleach (10% of the total volume). Contents of the waste container may contain biohazardous material.

Removing Waste from a Cytek 4-L Waste Tank or a 20-L Cubitainer

- 1 Disconnect the waste line orange quick-connect from the cubitainer cap or 4-L waste tank. Disconnect the waste level sensor.
The waste level sensor connector for the cubitainer is on the cubitainer cap. The waste level sensor connector for the 4-L tank is on the front of the cytometer.
- 2 Remove the waste cap from the cubitainer or the lid from the 4-L waste tank, taking care not to damage the liquid level sensor.
- 3 Dispose of the waste per local regulations.
- 4 Add 2 L of undiluted bleach to the waste cubitainer, or 400 mL of bleach to the waste tank.
- 5 Replace the waste cap/lid to the container. Hand-tighten the cap/lid until it is fully closed.
- 6 Reattach the waste line and level sensor line to the cap/lid and front of the cytometer.
- 7 If the cytometer is powered on and the software is connected, verify that the software waste indicator is green.

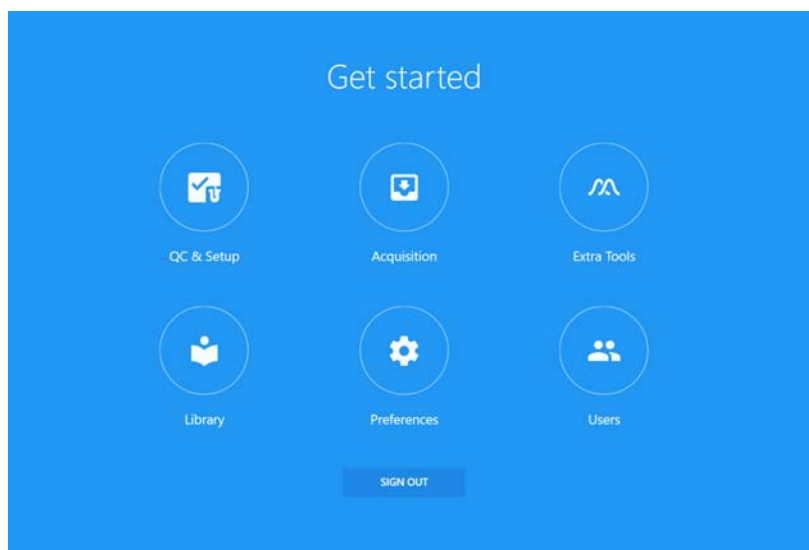
Starting Up the System

- 1 Turn on the workstation, then turn on the cytometer.
 - **NOTE:** Ensure that a tube containing 1 mL of deionized (DI) water is loaded on the SIP before launching SpectroFlo software. The tube is required for the SIT depth calibration and to flush liquid through the flow cell to remove bubbles that may have formed.
- 2 Launch SpectroFlo software and log in by entering your user name and password and clicking SIGN IN.

You can start typing your user name to display a list of names beginning with the letter(s) you type.

The cytometer initialization procedure begins. Sheath fluid is flushed through the fluidics lines to prevent any saline buildup, and the system calibrates the SIT depth.

- 3 Select QC & Setup from the Get started menu.



- 4 Check the status indicators in the lower-right corner of the screen.
 - Ensure the indicator for Connected is a green checkmark. It may take a few minutes for the indicators to update.



If the indicator shows the instrument is not connected, check to ensure that the USB connection between the cytometer and workstation is plugged into the appropriate ports. See [“Back of Cytometer” on page 13](#).

- Ensure the status indicators for sheath and waste are green before proceeding.

Fluid Indicator	Meaning	What to do
Yellow sheath	Sheath tank is low. Acquisition will continue for 5 minutes before the sheath is empty. You can refill the sheath during this time.	Fill the sheath tank (see “Filling the Sheath” on page 23).
Red sheath	Sheath tank is empty.	
Yellow waste	Waste tank is nearing capacity.	Empty the waste tank (see “Emptying the Waste” on page 24).
Red waste	Waste tank is full.	

- 5 Wait 30 minutes before running Daily QC. See [“Performing Daily QC”](#) on page 27.

Shutting Down the System

Run the shutdown procedure at the end of each day that you use the instrument. The shutdown procedure flushes the flow cell and sample lines with 10% bleach solution, DI water, 25–50% Contrad 70, and DI water. The procedure takes approximately 5 minutes.

■ **IMPORTANT:** If you are operating in plate mode, you must switch to tube mode before performing Fluidics Shutdown. In tube mode, the SIT will remain extended from the SIP at the end of shutdown to ensure the SIT does not dry and form clogs.

To shut down the system:

- 1 Follow the Fluidics Shutdown procedure. See [“Fluidics Shutdown”](#) on page 108.
- 2 When the shutdown procedure is complete, exit SpectroFlo software by clicking the X in the upper-right corner of the application window.
- 3 Turn off the cytometer and workstation.

QC & Setup

Daily QC

Run Daily QC using SpectroFlo QC beads prior to acquiring samples to ensure that the cytometer is performing optimally. Daily QC assesses the instrument's optical alignment and the system performance drift by measuring rCVs and gains needed to place the beads at the target locations for each detector. During QC, laser delays and area scaling factors are optimized and gain settings are adjusted to account for day-to-day instrument variability. Upon completion of Daily QC, a QC report is generated. QC reports can be reviewed under the Reports tab.

Performance can be tracked and charted over time in the Levey-Jennings tab. The software can be configured to display a warning if the QC result on the QC report exceeds user-defined criteria. See ["Alarm Ranges" on page 39](#).

Performing Daily QC

If you have a Loader and plan to perform Daily QC in a microtiter plate, you will need to change to plate mode. See ["Using the Loader" on page 94](#) for information.

- 1 Allow 30 minutes to pass after turning on the system to ensure the optics compartment is warmed up.
- 2 Prepare SpectroFlo QC beads (1 drop of beads in 0.3 mL DI water or PBS) in a tube or plate well.

■ **NOTE:** For Aurora systems that include a UV laser, use SpectroFlo QC beads with a 2000 series lot number.

■ **NOTE:** Always prepare beads in the same solution used for the sheath solution on the instrument. The bead diluent and instrument sheath solution must match. If you use DI water to prepare the beads, the beads will begin to degrade within a few hours. Do not reuse the beads prepared in DI water.

The SpectroFlo QC beads are 3- μ m hard-dyed, polystyrene beads that have a single fluorescence intensity. They can be excited by each laser and emit fluorescence in all detector channels.

3 Select QC & Setup from the Get started menu.



4 Select the current bead lot from the Bead Lot menu.

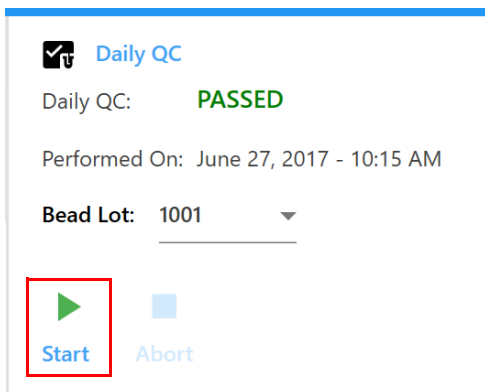
Each time you open a new lot number of SpectroFlo QC beads you must import the bead lot ID into the library so it is accessible when you run QC. Bead lot files can be downloaded from the Resources section at <http://www.cytexbio.com>.




Different bead lots have different fluorescence intensities. Always select the correct bead lot when performing Daily QC.

5 Start acquisition:

- Tube: Load a 12 x 75-mm tube of the beads onto the SIP. Select Start to begin acquisition.



- **Loader:** Ensure the correct plate type is selected (96 U-, V-, or Flat-Bottom). Click in the plate to select the well where the QC beads are located. Well A1 is selected by default, but you can choose any well. Select Eject, load the plate onto the plate stage, then click Load followed by Start to begin acquisition. Or, select Start to automatically load the plate and begin acquisition.


 **Daily QC**

Daily QC: **PASSED**

Performed on August 21, 2018 - 11:32 AM

Bead Lot: 1002 96 U-Bottom

Select QC well:

	1	2	3	4	5	6	7	8	9	10	11	12
A		<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
B	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
C	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
D	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
E	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
F	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
G	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>
H	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>






As the instrument begins acquiring the QC beads, they appear in the scatter plot. The laser delays are initially set to 0, then optimized thereafter. The performance measurements are established and compared to the pass/fail criteria (see ["Pass/Fail Criteria" on page 30](#)).

The procedure takes approximately 3 to 5 minutes to complete. Once acquisition is complete, two SIT Flushes are automatically performed to clear the beads from the sample line.

The following message is displayed when Daily QC passes. To view the QC report, click View Report (see ["QC Report" on page 30](#)).

 **Setup Completed**

Daily QC has completed and PASSED!

[Close](#) [View Report](#)

If QC fails, remove the tube or plate and follow the guidelines that appear. The recommended solution will vary depending on the reason for the failed test.

Daily QC Failed

Daily QC failed to place the QC beads on scale.

Please try the following:

1. Make sure the concentration of your beads sample is appropriate (between 100 - 1000 events per second).
2. Go to Acquisition to run your beads sample in an experiment and make sure your sample is appropriate.
3. Remix your beads sample run QC again.

If the problem persists, please contact Cytek Service.

OK

You are now ready to run reference controls, if applicable, or acquire samples.

QC Report

At the completion of Daily QC, a QC report is generated. The report includes the following sections:

- The header section contains the Pass/Fail status of the run, name of the instrument, instrument configuration, date the Daily QC was run, user who ran the Daily QC, instrument serial number, and SpectroFlo QC bead lot and expiration date.
- The results section contains the gain, gain change, median fluorescent intensity of the QC bead, %rCV, and a pass/fail indicator for each detector channel. The center wavelength of the detector is shown in parentheses next to the detector name.
- The Laser Settings section contains the laser delays for all non-primary lasers, and area scaling factors for all lasers and the FSC detector.

Pass/Fail Criteria

The pass/fail criteria are the following:

- %rCV must not exceed 6% for the FSC channel
- %rCV must not exceed 8% for the SSC and SSC-B channels
- %rCV must not exceed 6% for the third channel of each laser (V3, B3, R3, YG3, and UV3)
- % delta gain change for all channels must not exceed 100% from the last Daily QC run performed by Cytek Service personnel.

QC reports are automatically exported as CSV files to the Setup folder (C:\CytekbioExport\Setup). The number of reports listed in the Reports screen can be set in the Preferences. See [“QC Setup Preferences” on page 89](#) for more information.

An example QC Report from a five-laser system is shown.

Reference Controls

Reference controls, obtained by running single-stained and unstained samples, provide the individual fluorescence spectra necessary to unmix the data. Either beads or cells can be stained for use as reference controls. These controls can be acquired in the reference group of the experiment during acquisition, or they can be acquired as reference controls in the QC & Setup module. If reference controls are acquired in the QC & Setup module, they are stored and can be used as reference controls for unmixing in subsequent experiments.

A wizard guides you through recording reference controls.

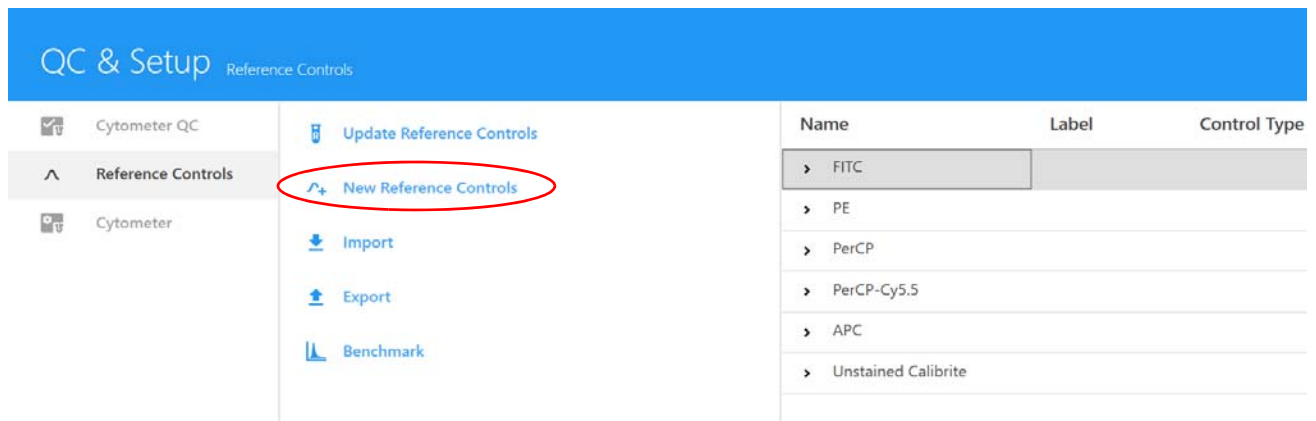
Running Reference Controls

To create reference controls you will need to select the fluorescent tags, choose the control type (beads or cells), then label the fluorescent tags (for example, using CD nomenclature). You can also append existing FCS files when running reference controls.

Run Daily QC to ensure the instrument is performing optimally before running reference controls.

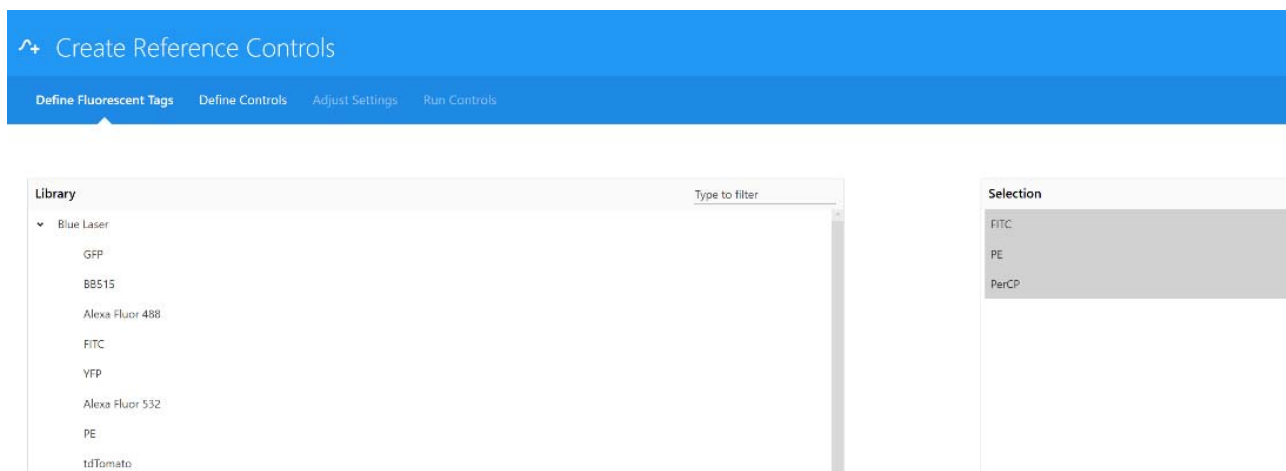
- 1 Select Reference Controls from the QC & Setup module.
- 2 Select New Reference Controls from the Reference Controls tab.

A wizard opens allowing you to create new reference controls. You can also append existing FCS files.



- 3 Select the fluorescent tags. The left pane displays the fluorescent tag groups found in the library.
 - Click the arrow to the left of the fluorescent tag group name to view the fluorescent tags in the group. (The default fluorescent tag groups are Blue Laser, Red Laser, Violet Laser, Yellow Green Laser, Ultraviolet Laser, Fluorescent Proteins, and Viability Dyes and contain a list of commonly used fluorescent tags excited by their respective laser).
 - From the expanded list of fluorescent tags, select the fluorescent tags used in the experiment. Once selected, the fluorescent tags appear in the selection pane on the right. You can select fluorescent tags by dragging and dropping, double-clicking, or using the Add button. Multiple tags can be chosen at one time. Confirm the tags selected, then click Next.

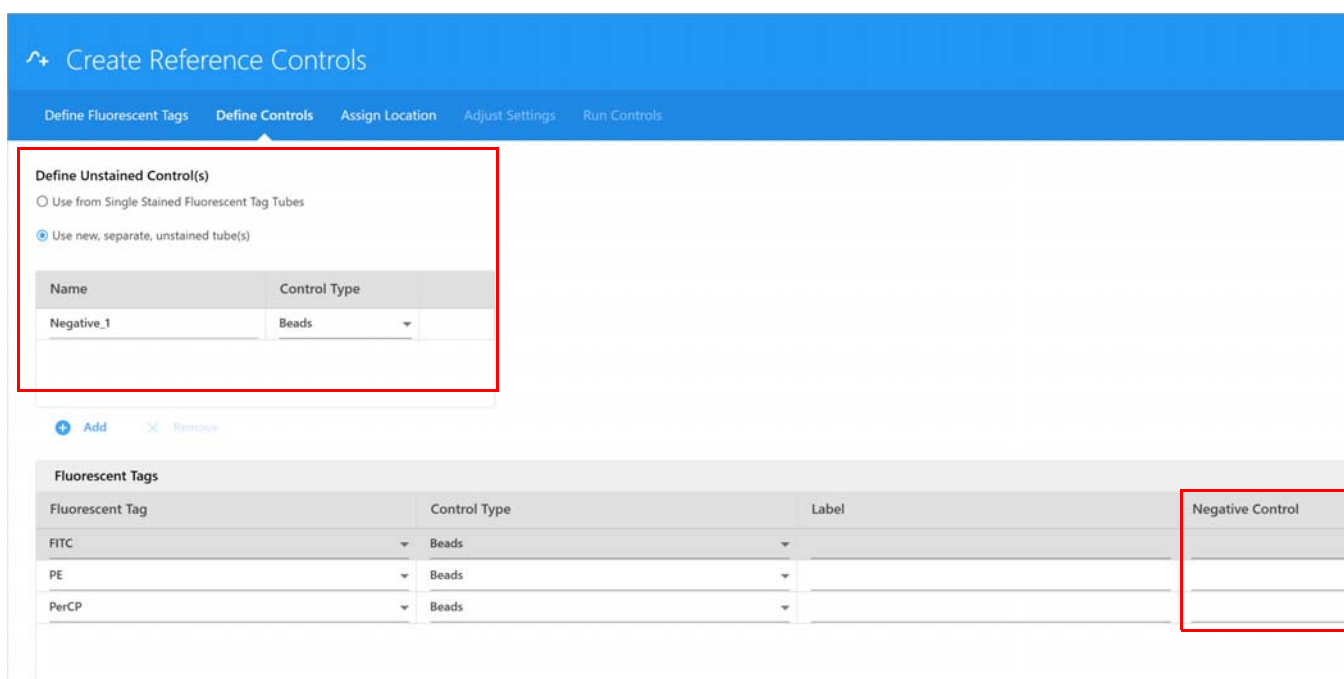
■ NOTE: The list of fluorescent tags can be edited in the library. You can use the library to add fluorescent tags that are not present in the default list. See “Fluorescent Tags” on page 75 for more information.



4 Define the unstained control(s).

Choose either single-stained fluorescent tags, or new, separate, unstained for the control tubes. If using a control that has a positive population only, you will need an unstained control of the same type as the stained.

Either beads or cells can be stained and defined as control types. If you select Use new, separate, unstained tube(s), you can add more unstained tubes, as necessary. Then select the specific unstained tube in the Negative Control column of the Fluorescent Tags table below.



- 5 Define the control type (beads or cells) for each fluorescent tag in the Fluorescent Tags table.
- 6 (Optional) Enter labels associated with the fluorescent tag for identification and tracking.
- 7 If applicable, enter the lot number(s) of the reference controls.

■ **NOTE:** If you selected Label/Lot Specific Unmixing in the Acquisition Preferences (see [page 82](#)), During unmixing, the software will search the library and experiment reference groups for reference controls that have the same fluorescent tag, label, and lot information to use the corresponding control for unmixing.

Define Unstained Control(s)

- Use from Single Stained Fluorescent Tag Tubes
- Use new, separate, unstained tube(s)

Name	Control Type
Unstained_1	Beads
Unstained_2	Beads

+ Add X Remove

Fluorescent Tags			
Fluorescent Tag	Control Type	Label	Lot
FITC	Beads		
PE	Beads		
PerCP	Beads		

- 8 Once the controls have been defined, click Next.
- 9 Load the control sample(s).
 - Tube mode: Load the appropriate control sample onto the SIP.
 - Loader mode: Assign the location of each control by dragging the control (unstained and fluorescent tags) to the appropriate well on the plate. You can use the auto-fill feature to fill the remaining wells. Click Next. Load the plate by clicking Eject, then loading the plate on the plate stage.

The screenshot shows the 'Create Reference Controls' workflow. The 'Assign Control Location' step is active, displaying a list of controls on the left and a 96 U-Bottom plate grid on the right. The controls are: Unstained_1 (Beads) assigned to A1, FITC (Beads) to A2, PE (Beads) to A3, and PerCP (Beads) to A4. The plate grid shows wells A1, A2, and A3 in the first row highlighted in blue, indicating they are selected for control assignment.

- 10 Click Start to preview the sample data. This step allows you to ensure that all populations for all control samples are on scale. If necessary, adjust gain settings.

Gain settings for all channels can be selected from the User Settings drop-down menu, or they can be individually adjusted for each channel using the detector gain spinboxes (V1–V16, B1–B14, YG1–VG10, R1–R8, and UV1–UV16).

■ **NOTE:** CytekAssaySetting was generated by measuring the optimal resolution of human lymphocytes stained with CD8 and CD4 labeled with various fluorochromes. These settings provide a place to start for most immunophenotyping applications.

The Adjust Settings screen allows you to view the data to ensure that the positively stained fluorescent particles are on scale. Adjust the threshold and FSC and SSC gains. FSC gain can be adjusted from 1-1,000. SSC and detector channel gains can be adjusted from 10-10,000. If the positive population is off-scale for any detector channels, lower the gain setting for that channel. If the positive population is not sufficiently separated from the negative population within a specific channel, verify that CytekAssaySetting is selected, as these settings ensure adequate resolution. If the positive population is still not clearly separated from the negative population, review the sample preparation and staining procedure.

Once gain settings have been confirmed, unstained and reference controls are ready for acquisition.

■ **NOTE:** Dim markers may not separate from the negative population regardless of how much the gain is increased.



11 Preview the data for the remaining controls by clicking Stop, then:

- Tube mode: Loading the next control onto the SIP and clicking Start.
- Plate mode: Selecting the next control well and clicking Start.

12 Select Next when you are satisfied with the gain settings. Proceed to running controls.

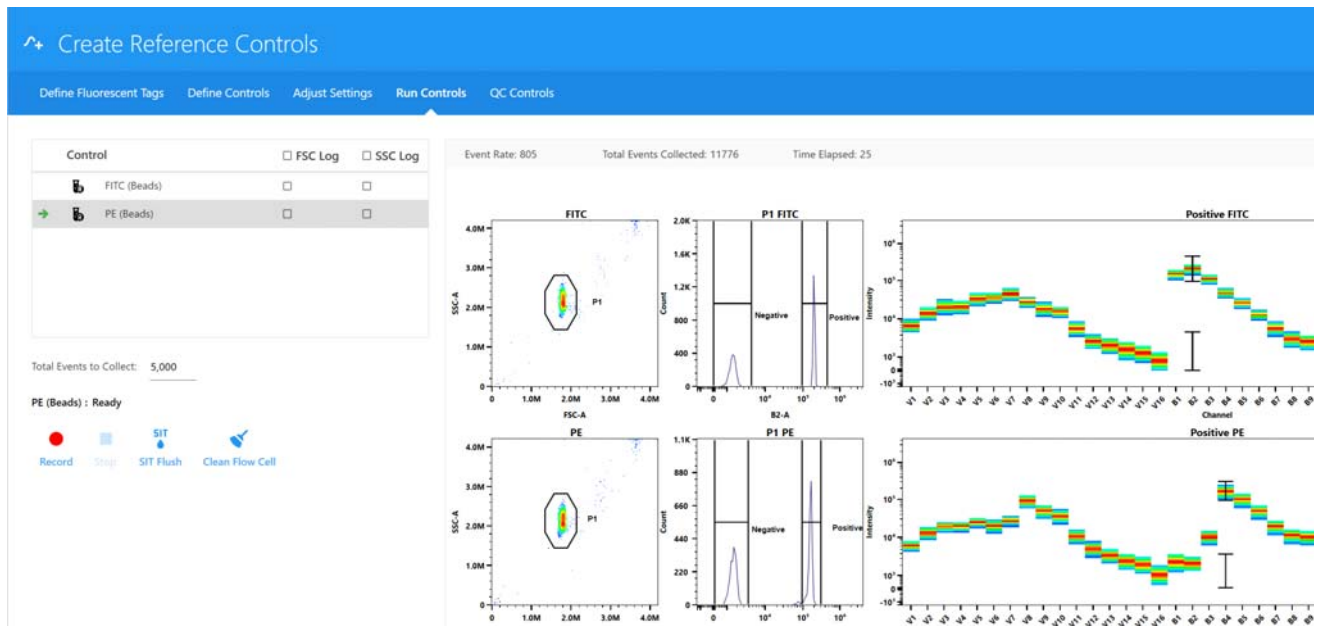
13 If you are running in Tube mode, place a tube of the appropriate single-stained particles on the SIP.

14 Click Record to begin acquiring.

Make sure to follow the order listed in the left-hand panel. When running in plate mode, you can select to record a single well/control or the entire plate.

During acquisition the spectra plot for each fluorescent control is displayed. The plots show all the channels across all lasers in the x-axis vs mean fluorescence intensity (MFI) of the fluorescent tag.

If the acquisition criteria is not met within 15 minutes, the run will stop.



15 During acquisition, obtain spectral information by moving the polygon gate on the FSC-A vs SSC-A plot to include the population of interest.

Hold down the Ctrl key while adjusting the gate to move the polygon gates for all the scatter plots at once. The gated population appears in the histogram, which is set approximately to the peak emission channel of the fluorescent tag to be acquired. The emission spectrum of the population is displayed in the spectrum plot.

Adjust the positive gate on the histogram. The software automatically displays the emission spectrum of the positive particle in the spectrum plot. SpectroFlo software sets the default gate near or on the peak emission channel. The gate can be selected manually. It is best to set the gate on the brightest emission, as this can make distinguishing the positive and negative populations easier and provides better visualization of the spectrum.

■ **NOTE:** Unmixing results are unaffected by the position of the interval gate in the spectrum plot.

Readjust the positive and/or negative gate on the histogram, if necessary.

16 Continue recording each control.

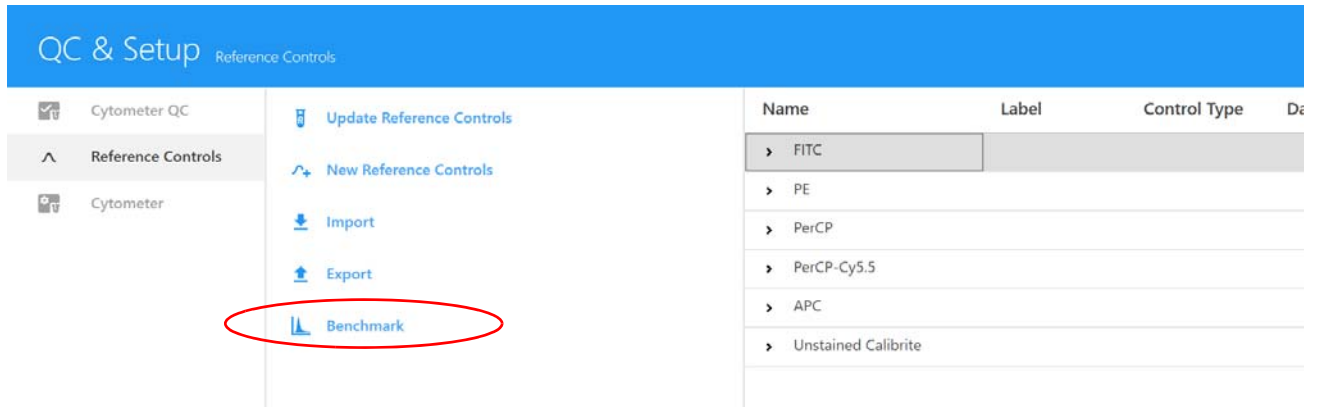
17 Select Save to save the reference controls.

FCS files for saved reference controls are stored in the export Setup folder by default, or the folder you set as the default. See [“Storage Preferences” on page 88](#) for information.

Setting Reference Controls as Benchmarks for Reference Control QC

You can select reference controls to be used as benchmarks for QC. The reference controls you select as benchmarks allow you to check the quality of your reference controls prior to unmixing. If a reference control is suboptimal, the benchmark will indicate whether the reference control spectra appears as expected. If you choose not to designate reference controls as benchmarks, the reference controls run during QC & Setup or during the experiment will be used as-is without a quality control check.

- 1 Select Reference Controls from the QC & Setup module.
- 2 Select Benchmark from the Reference Controls tab.



- 3 Select the reference controls that will be the benchmarks.

Click All to select all the controls in the list.



- 4 Click Save.

Updating Reference Controls

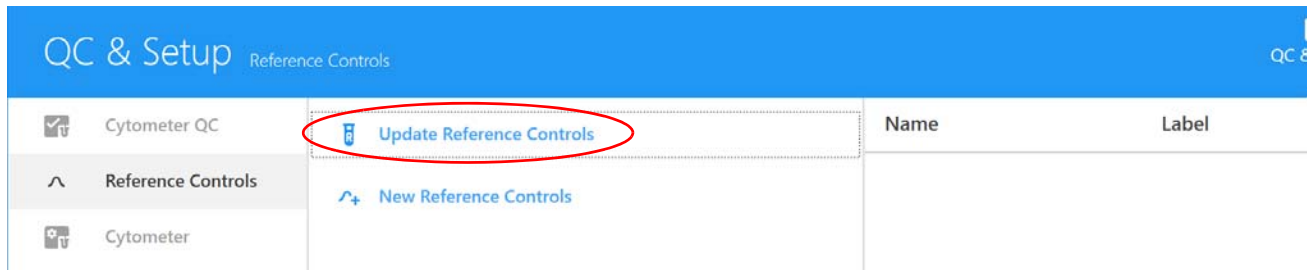
You may wish to update the reference controls if any of the following occur:

- Major service performed on the instrument
- Fluorochrome exhibiting signs of instability resulting in changes in its emission spectrum
- Staining protocols change, for example different buffers or fixatives are used

The Reference Controls tab displays the reference controls saved in the library. Click the arrow next to the control name to display the details.

To Update Reference Controls

- 1 Select Update Reference Controls from the Reference Controls tab in the QC & Setup module. A wizard opens allowing you to update reference controls.



- 2 Follow steps 3 through 12 in "Running Reference Controls" on page 32.

Levey-Jennings Tracking

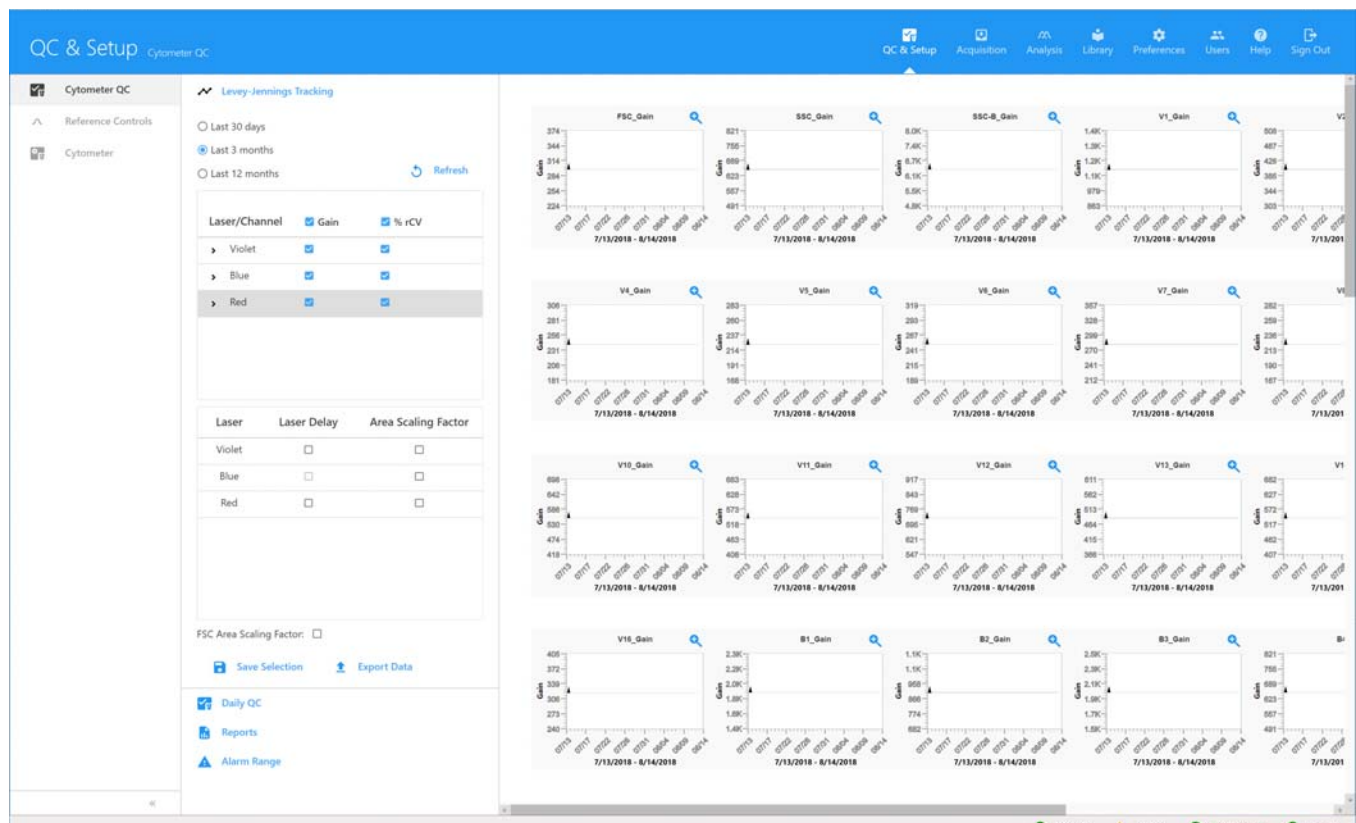
Levey-Jennings reports track the gain, %rCV, laser delay, and area scaling factor for all detector channels over time, allowing you to view the system's performance and ensure that the system is reproducing consistent results. Select the parameter(s) you wish to track.

The graphs in the report show you random errors or shifts and trends in the data for each parameter. Data from the last 30 days, 3 months, or 12 months can be included in the reports.

Select the Area Scaling Factor checkbox to plot Levey-Jennings for area scaling factor.

Use Save Selection to save the LJ tracking settings.

To export Levey-Jennings data to a .csv file, click Export Data.

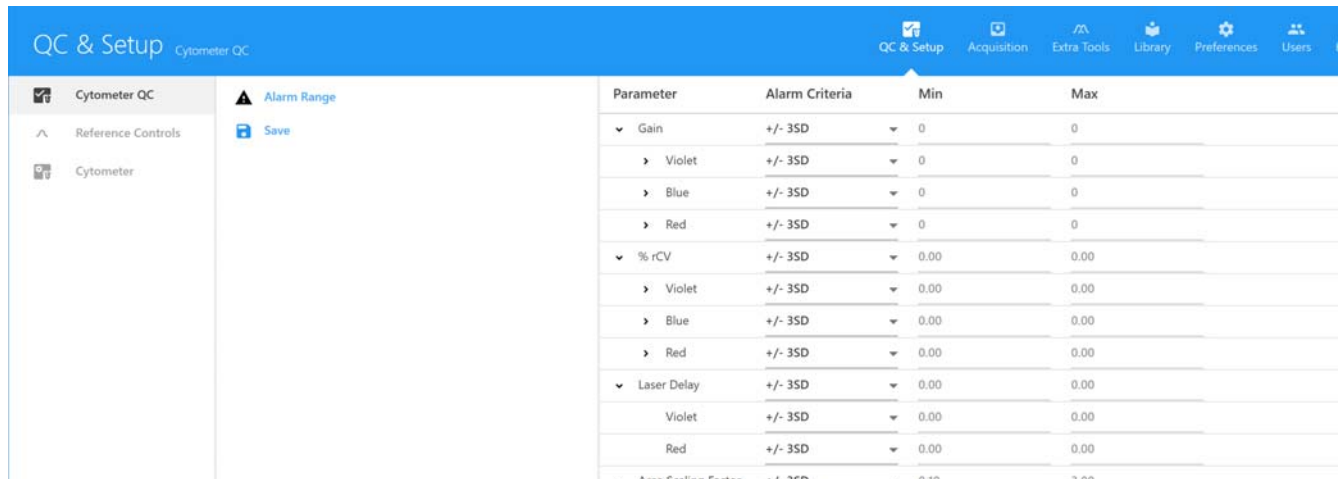


Gain Settings

The amount of signal amplification applied to each detector channel can be modulated by increasing or decreasing the amount of gain applied. The gains for every detector channel can be saved and are collectively known as the user settings. User gain settings are stored as a ratio against the Daily QC. Every time Daily QC is performed, User Settings will be adjusted accordingly.

Alarm Ranges

You can set an alarm to warn you when the gain and %rCV exceeds the passing criteria that you define. This changes the outliers (shown in red) in the LJ graphs. Select Alarm Range from the Cytometer QC tab, then adjust the SD range (plus or minus) for individual detectors for each laser.



The screenshot shows the 'QC & Setup' interface for 'Cytometer QC'. The left sidebar contains 'Cytometer QC', 'Reference Controls', and 'Cytometer'. The main area is titled 'Alarm Range' and includes a 'Save' button. A table displays the configuration for various parameters, including Gain, %rCV, Laser Delay, and Area Scaling Factor, with columns for Alarm Criteria, Min, and Max.

Parameter	Alarm Criteria	Min	Max
Gain	+/- 3SD	0	0
> Violet	+/- 3SD	0	0
> Blue	+/- 3SD	0	0
> Red	+/- 3SD	0	0
% rCV	+/- 3SD	0.00	0.00
> Violet	+/- 3SD	0.00	0.00
> Blue	+/- 3SD	0.00	0.00
> Red	+/- 3SD	0.00	0.00
Laser Delay	+/- 3SD	0.00	0.00
Violet	+/- 3SD	0.00	0.00
Red	+/- 3SD	0.00	0.00
Area Scaling Factor	+/- 3SD	0.10	3.00

Acquisition

Raw vs Unmixed Data

SpectroFlo software saves flow cytometry data in the FCS 3.1 format. Data is saved in both raw and unmixed formats. Raw data contains all the fluorescence information from each detector (ie, V1, V2, V3, etc). Each detector channel is designated by its excitation laser and position in the array. For example, B3 is the third channel of the blue laser detector array.

Unmixed data contains all the fluorescence information from each fluorescent tag in the experiment. To unmix data, single stained controls (or reference controls) for each of the fluorescent tags (as well as an unstained control) are required. During unmixing a mathematical algorithm is used for the decomposition of the fluorescent components in the sample using the reference controls. Parameters in unmixed data will display as the fluorescent tag name along with their associated labels.

The Acquisition module provides the tools that allow you to create an experiment. An experiment is a set of tubes, instrument settings, acquisition criteria (stopping rule), fluorescent tags, labels, and worksheets designed for the acquisition of samples. See [“About Experiments” on page 18](#).

New and saved experiments can be created or accessed in the Experiments tab of the Acquisition module.

Unmixing and Compensation

Raw FCS files can be spectrally unmixed in the following ways:

- Reference group from the experiment – Reference controls collected as FCS files within the experiment can be used to unmix using the Unmixing wizard in the Acquisition module.
- Reference controls run during QC & Setup – Reference controls run during QC & Setup can be used to unmix using the Unmixing wizard in the Acquisition module.
- Unmixing from the Extra Tools module – FCS files collected from different experiments can be unmixed in the Extra Tools module. FCS files can be imported and unmixed in this module.

Raw FCS files can also be compensated with the conventional method using the Virtual Filters tab in the Extra Tools module. Detector channels can be binned together to simulate the analysis of the data as if it were acquired using a filter. See [“Virtual Filters” on page 70](#) for more information.

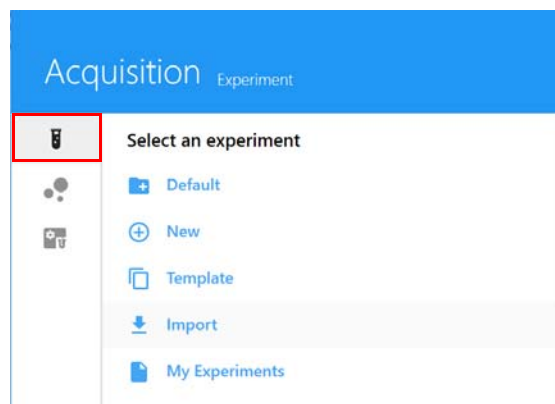
Setting Up an Experiment

An experiment can be saved as a template or created for one time use. Setting up the experiment in SpectroFlo software involves:

- 1 (Optional) Providing a name and description for the experiment. A default name is provided.
- 2 Specifying the fluorescent tags used in the experiment.
- 3 Defining the reference group with associated reference tags, labels, and lot numbers, as needed.
- 4 Adding labels and lot numbers to each of the fluorescent tags.
- 5 Adding custom keywords. Custom keywords can be defined in the Library.
- 6 Selecting an acquisition worksheet—either new or a template.
- 7 Defining acquisition criteria (stopping rule based on events, time, or volume).

Acquisition Experiment Overview

The Acquisition module provides the necessary elements for data collection within the experiment. Click the Experiment icon in the far left pane to open a template, the default, or a new experiment using the wizard.



Experiment Display

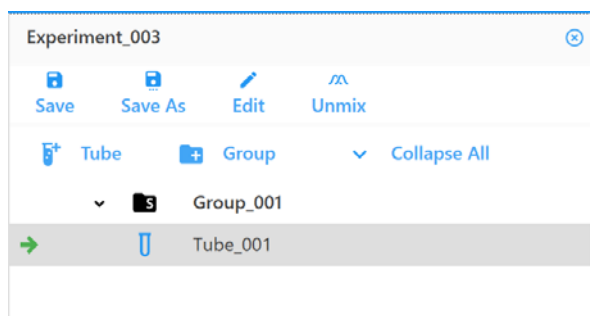
The experiment display in the Acquisition module includes the following panes. To show, hide, or undock (float) these panes from the experiment panel, click the corresponding icons in the top-right corner of the pane.

Group-Tube List and Hierarchy

The samples are listed in the upper left of the screen. Samples can be organized into groups. Use the ⊕ Tube and ⊕ Group icons to add tubes and groups. If running in plate mode, click Add Plate to add plates. Click Plate View to display a graphic image of the 96-well plate instead of the list of groups.

Click Save to save changes to the experiment, or click Save As to save an experiment template. Click Edit to edit the experiment. Worksheets can be applied to the experiment, groups, or individual tubes.

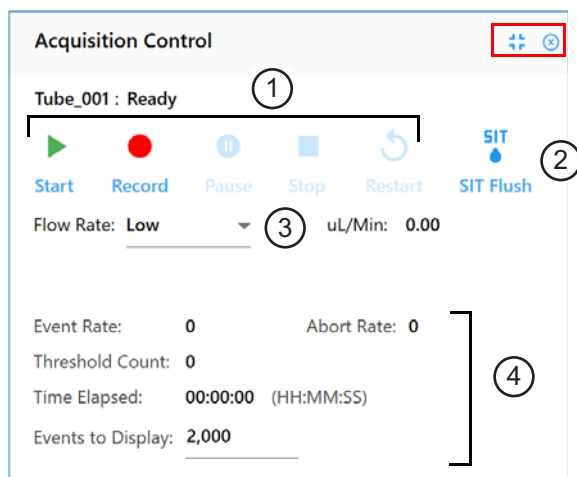
Tube mode



Acquisition Control

The Acquisition Control pane allows you to start, stop, and pause acquisition, record data, and restart acquisition counters. The acquisition controls are enabled when a tube is present on the SIP or when in plate mode. To show, hide, or undock (float) this pane from the experiment panel, use the dock/undock and hide icons in the top-right corner.

For information on the Loader Acquisition Controls when in plate mode, see “[Loader Acquisition Controls](#)” on page 96.



The following table describes the controls in the Acquisition Control pane.

No.	Control	Description
1	Start/Record/Pause/Stop/Restart	Start and Record are enabled when a tube is present on the SIP. Select Start to start acquisition Select Record to record data. Record can also start acquisition. Select Pause to pause recording. While paused you can adjust the flow rate. Select Record again to continue. Select Stop to stop acquisition. Select Restart to restart the acquisition counters. All events and results displayed are refreshed. Stop and Restart are enabled once Start is selected. Stop and Pause are enabled once Record is selected.

No.	Control	Description
2	SIT Flush	Select to perform a SIT Flush
3	Flow Rate	Select Low (15 µL/min), Medium (30 µL/min), or High (60 µL/min). The exact flow rate is displayed.
4	Event Rate, Abort Rate, Threshold Count, Time Elapsed	Displays the real-time counts during acquisition.
5	Events to Display	Enter the number of events to display during acquisition.

Instrument Control

The Instrument Control pane consists of the Gain, Threshold, Signal, and Lasers tabs for use in adjusting the instrument.

User Settings allow you to select CytekAssaySetting, Default, or any saved user settings for the experiment. We recommend using CytekAssaySetting as a starting point. This setting provides the optimal resolution for each channel, accommodates bright signals, and minimizes spread.

The screenshot shows the Instrument Control interface with the following components and callouts:

- Callout 1:** Points to the Gain tab in the top navigation bar.
- Callout 2:** Points to the FSC channel and its threshold value (100,000) in the Threshold tab.
- Callout 3:** Points to the Scatter Channels section, specifically the FSC, SSC, and SSC-B rows.
- Callout 4:** Points to the Area Scaling Factor table in the Lasers tab.

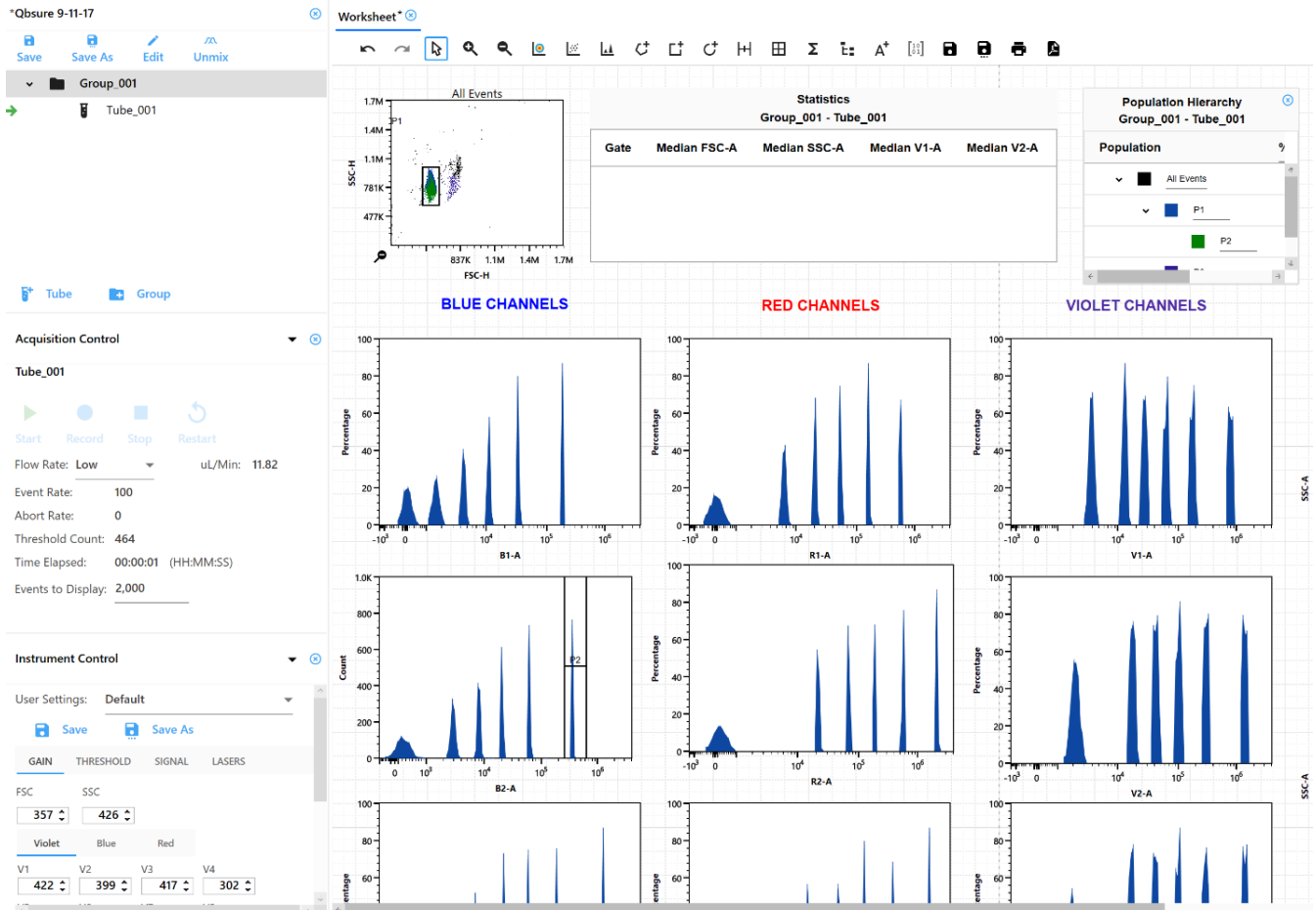
Laser	Area Scaling Factor
Violet	1.19
Blue	0.93
Red	0.99

The following table describes the tabs in the Instrument Control pane.

No.	Tab	Description
1	Gain	<p>Gains can be adjusted for all detector channels for all lasers using the gain spinboxes. FSC gain can be adjusted from 1–1,000. SSC and fluorescence detector gains can be adjusted from 10–10,000. To change the value that the gain increments, see “Acquisition Preferences” on page 81.</p> <p>Use All Channels % to increase/decrease all gains for a selected laser by the percentage you select.</p>
2	Threshold	<p>Use the Threshold tab to set the threshold parameter and minimum threshold channel value. Multiple parameters can be set as a threshold using either the AND or OR operator. Use OR when at least one parameter is available.</p>
3	Signal	<p>Use the Signal tab to select area, height, or width for each signal. Area and height can be selected for all channels. Width can be selected for only one channel per laser.</p>
4	Lasers	<p>Use the Lasers tab to set the area scaling factor and laser delay. These values are automatically set and updated in all user settings upon completion of the Daily QC.</p> <p>■ NOTE: If you run large cells and the lowest FSC gain setting is not low enough to see your cells, lower the FSC area scaling factor (for example, 0.5).</p>

Worksheet

The worksheet allows you to view the data in plots and create plots, statistics, population hierarchy, and gates. Click the worksheet icon in the far left pane to select, import, save, and print worksheets.



Worksheet Toolbar

A toolbar at the top of the worksheet area allows you to undo/redo, zoom; create plots, gates, statistics, population hierarchy, annotations; and save, print, and save a PDF of the worksheet. Hover the cursor over an icon to see a description and keyboard shortcut.



Plots

Four plot types can be created in the worksheet:

- dot plots
- pseudocolor plots (density plots)
- histogram plots
- spectral plots

To change the properties of a plot, right-click the plot and select Properties. You can select the plot type, parameters, scale, background color, and labels.

Plot Properties ⓘ

General

Plot Gate: All Events ▾

Plot Type: Pseudocolor Plot ▾

Parameters

X Axis Parameter: FSC-A ▾

X Axis Scale: Linear ▾

Y Axis Parameter: SSC-A ▾

Y Axis Scale: Linear ▾

Layout

Width 338 Height 254.6666

Density Plot Options

Density Levels: 10 ▾

Miscellaneous

Background Color:

Include Population Name

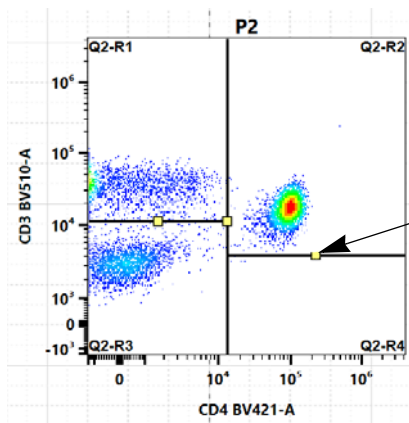
Include Tube Name

Include Custom Title

Gates

Gates types include:

- rectangle
- oval
- polygon
- interval
- quadrants and staged quadrants (select and drag an offset handle to move the quadrant segment up or down)



Click and drag handle to move quadrant segment up or down.

Gate Properties

Gates properties can be changed by right-clicking the gate. You can change the name of the gate, the color, and gate boundary line weight. You can also select whether to display the count and/or the % parent events within the gate, as well as the gate parameters.

Gate Properties ⊙

Gate Name: P2 _____

Gate Color: ■

Count % Parent

Gate Boundary Line Weight: **Normal** ▾

Parameters

X Axis Parameter: FSC-A _____

Y Axis Parameter: SSC-A _____

Logical Gates

Select multiple gates and right-click to open a menu to:

- Intersect the gates with the AND operator – events that are present in all of the selected gates are part of the intersected gate population.
- Join the gates with the OR operator – events that are present in at least one of the gates are part of the joined gate population.

Population Hierarchy Experiment_009-Group_001-Tube_001		
Population	% Parent ▾	Count
▼ ■ All Events	N/A	N/A
▼ ■ P1	N/A	N/A
■ P2	N/A	N/A
■ P3	N/A	N/A

Delete Gate

Intersect (And) Gates

Join (Or) Gates

Bring to Front

Gate Properties

Statistics

To create a statistics box, click the Statistics icon in the worksheet toolbar, then click in the worksheet area.



Select the population checkbox next to the populations that have stats to display. To add a statistic, select the statistic from the Statistics Variable list.

Select the parameter you would like to add for the statistics. Multiple parameters can be selected at once.

The software offers a counts/ μ L statistic that can be calculated for any gate.

To adjust the precision of the statistics, select the decimal place in the Decimal Places table. To remove a statistic, right-click the column header and select Delete.

Once the statistics are complete, you can export the stats as a CSV file to the location you choose. Statistics output can be appended or overwritten if you export the stats into the same CSV file.

Create Statistics Table

Title
New Statistics 01

Add Statistics

Population	Count	% Parent	% Grand Parent	% Total
All Events				
P1				
P2				
P3				

Statistics Variable: Median, rSD, % rCV, Mean, Max
Parameter: FSC-A, SSC-A, V1-A, V2-A, V3-A
Decimal Places: Median (0), rSD (0), % rCV (2), Mean (0), Max (0)

Buttons: Add Statistics, Cancel, Save

Creating a Default Experiment

Use the default experiment option to quickly begin an experiment. The default experiment contains a set of fluorescent tags, however, all parameters are user configurable.

- 1 Click Default in the Acquisition Experiment menu.

Acquisition Experiment

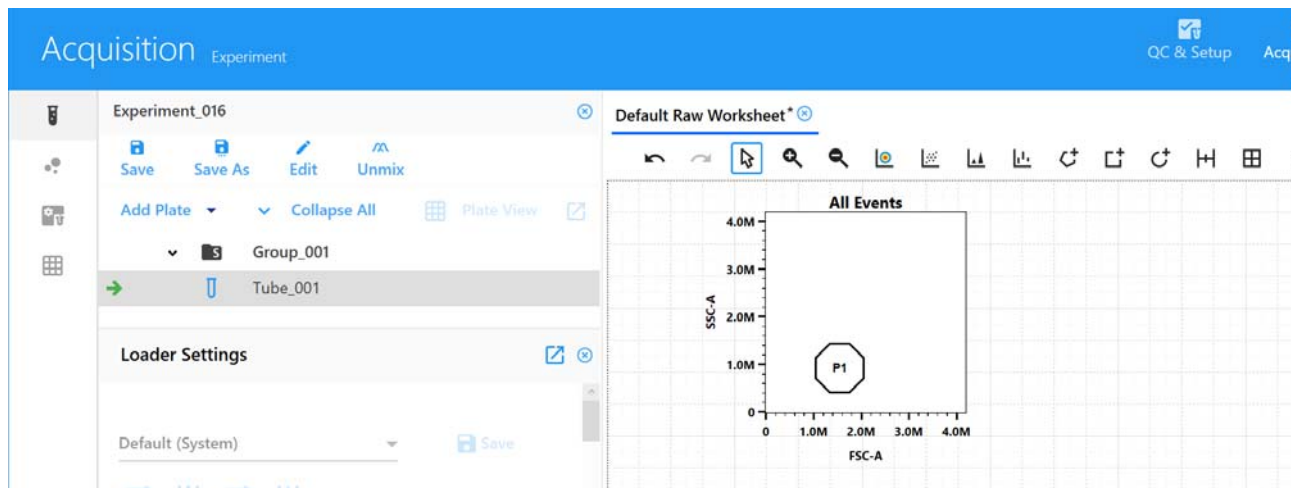
Experiment

- Worksheet
- Cytometer
- Plate Calibration

Select an experiment

- Default**
- New
- Template
- Import
- My Experiments

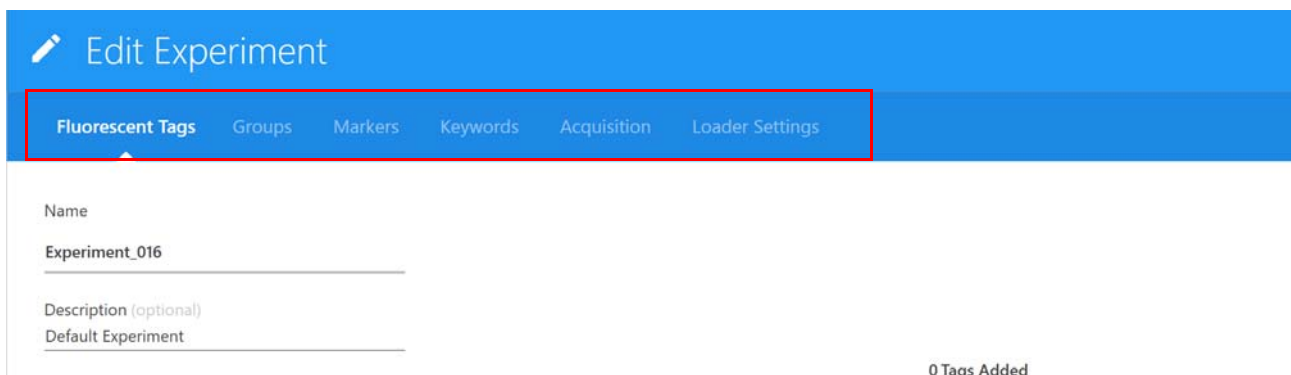
The default experiment opens with a default raw worksheet.



2 Click Edit.

The same wizard appears as when creating a new experiment. Follow the steps to select the fluorescent tags, add groups, and define makers, keywords, acquisition settings/worksheets, and Loader settings.

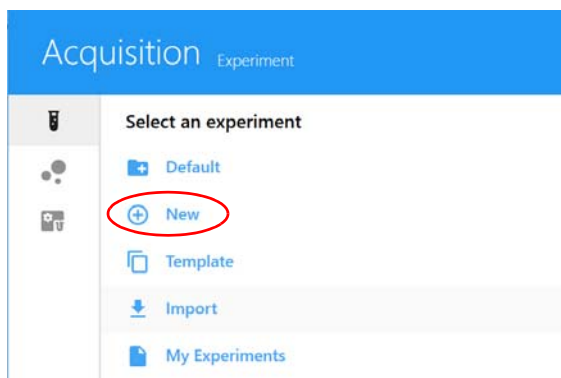
See “Creating a New Experiment” on page 50 for details on defining the information in the experiment wizard.



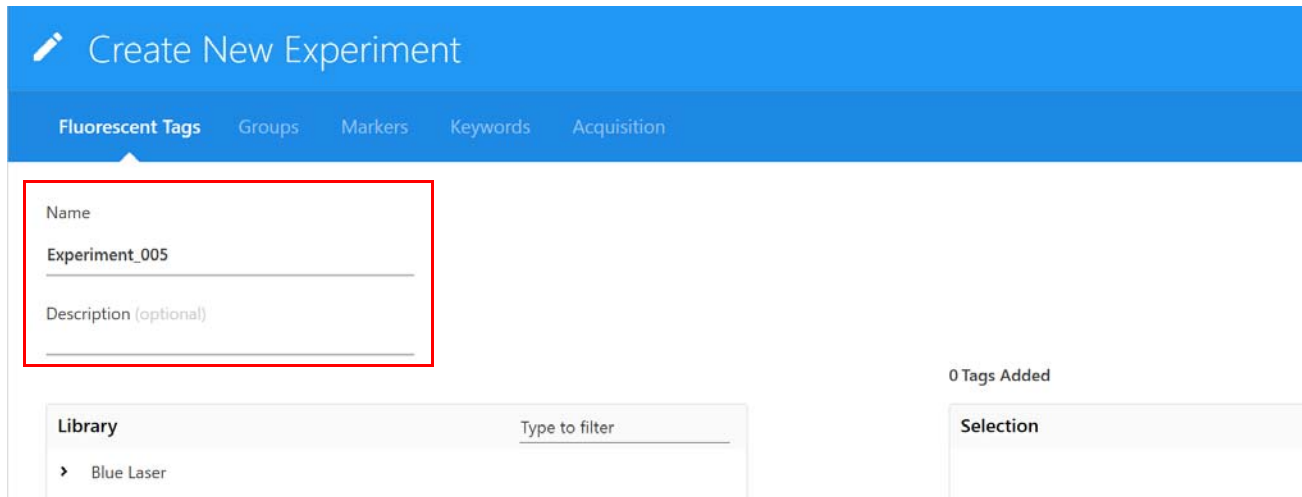
Creating a New Experiment

Selecting New in the Experiment menu opens the New Experiment wizard. The wizard walks you through the steps to create a new experiment.

1 Click New in the Acquisition Experiment menu.



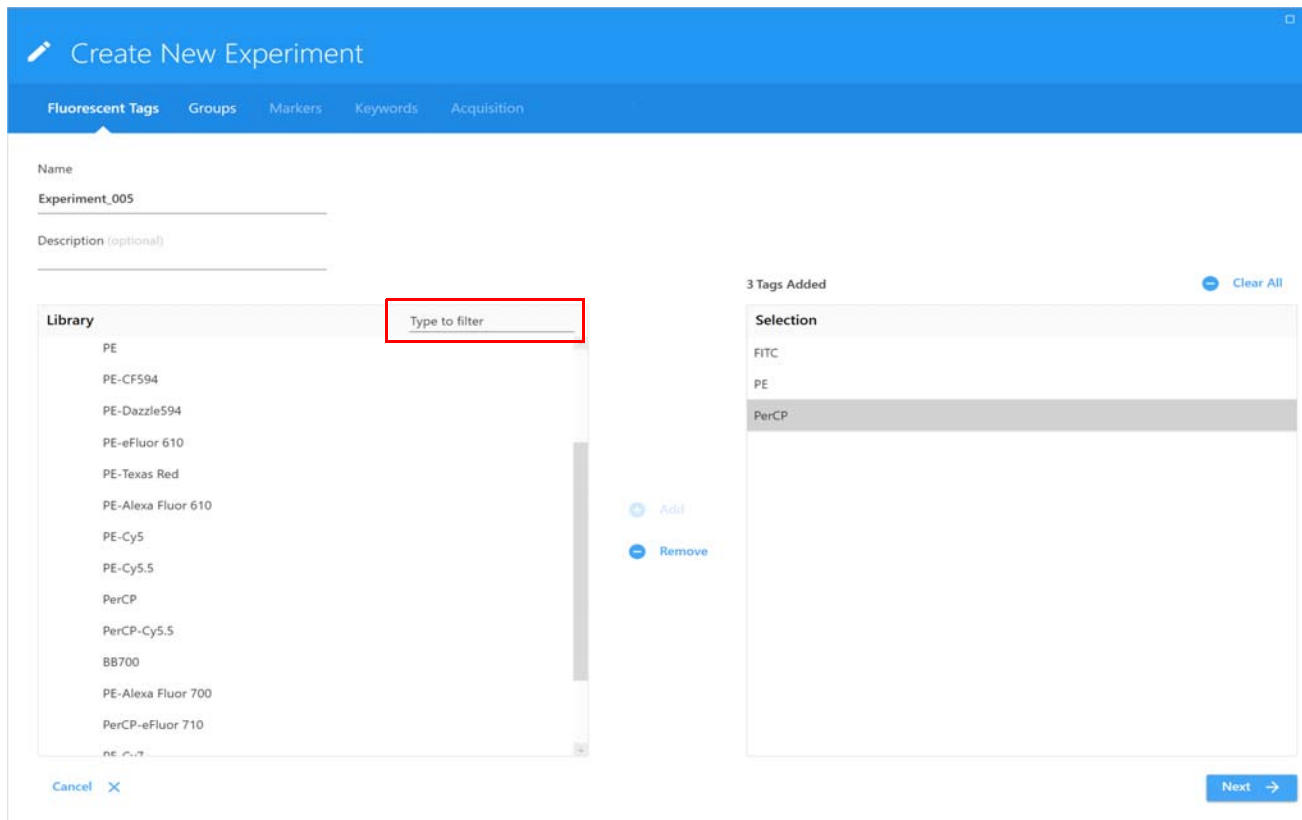
- 2 The Create New Experiment wizard opens. Specify a name for the experiment or use the default name. (Optional) Type in a description.




- 3 Click the arrow to the left of the group name (laser) in the Library pane on the left to display the list of its fluorescent tags. Select the fluorescent tags used in the experiment and click (+) Add to add them to the Selection list on the right. You can also double-click the tag to add it to the Selection list.

To quickly find a fluorescent tag, type the tag name in the Type to filter text box. A default list of fluorescent tags for each group is available in the library. See “Fluorescent Tags” on page 75.

You must select all fluorescent tags present in the experiment, as this will determine which reference controls are to be used during spectral unmixing.

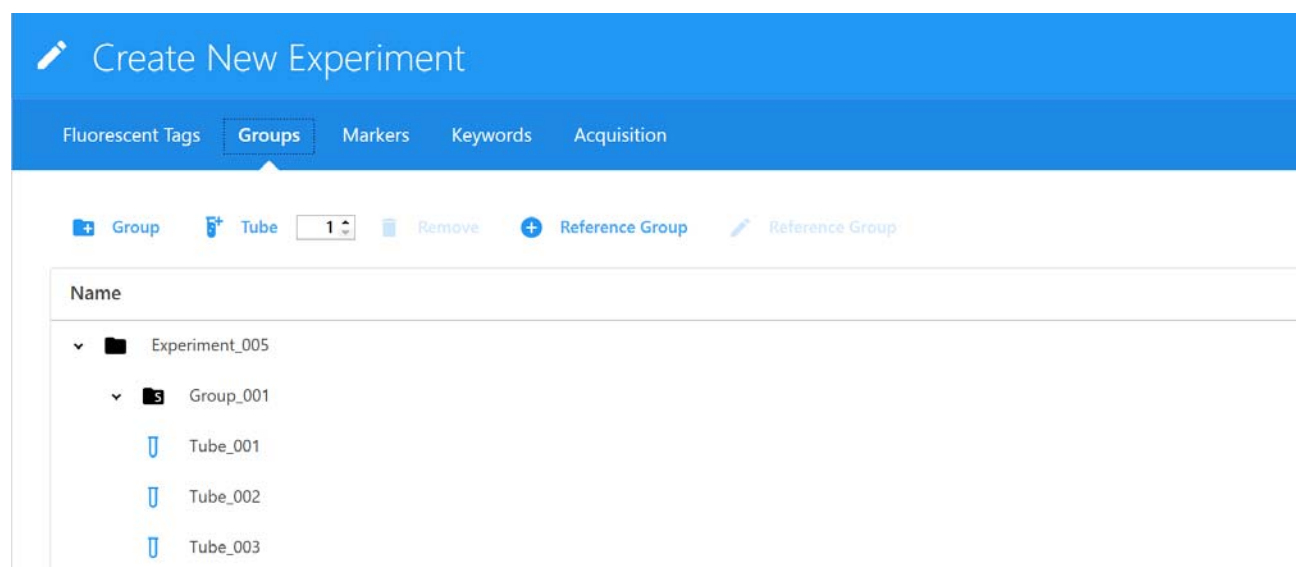



To remove individual fluorescent tags from the Selection list, click to select the tag, then click Remove. To remove all fluorescent tags, click Clear All.

- Once all fluorescent tags have been chosen from the Library list, confirm the list in the selection pane, then click Next.
- Create groups for your samples by selecting  Group. Add tubes to the groups.

For information on creating groups when in plate mode, see “Creating Groups When Using the Loader” on page 99, then proceed to [step 7](#).

■ **NOTE:** When running sticky samples, such as LNW samples, we recommend adding cleaning wells between samples to thoroughly clean the mixing probe. For example, add two wells, one with 10% bleach and the next with DI water. At the end of a plate, consider adding a group of four wells, two with 300 µL of 10% bleach and two with 300 µL of DI water. Program a long mix (15 seconds at 1500 rpm) to thoroughly clean the mixing probe.



- Select  Reference Group if you are intending to unmix with all or some controls acquired in this experiment.

This creates a list of reference control tubes for each fluorescent tag specified as part of the experiment.

■ **NOTE:** If you plan to unmix the samples using only reference controls run in QC & Setup, step 6 is not necessary.

■ **NOTE:** To mix and match reference controls acquired in the experiment with reference controls run in QC & Setup, define the controls to acquire in the reference group, acquire the controls, then after selecting Unmix, select the remaining controls from the reference controls run in QC & Setup.

- 7 **IMPORTANT:** Define an unstained control for autofluorescence by selecting its control type (beads or cells). The unstained control needs to be of the same type and prepared in the same way as the samples, as this will ensure accurate unmixing and autofluorescence quantitation.

- 8 If applicable, select Define Extra Unstained Control(s) for Spillover Calculation to use a different unstained control to calculate spillover for your reference controls. Then enter a name and control type for this extra unstained control.

For example, if test samples are cells and the reference controls are beads, all with only positive peaks, you will need to run a separate tube of negative beads for the spillover calculation. An extra unstained control is not needed if your unstained autofluorescence control (and sample) is the same type as the reference controls.

- 9 Select the control type (beads or cells) for the single-stained reference controls.
- 10 (Optional) Enter the label (for example, CD nomenclature) that is conjugated to the fluorescent tag.
- 11 If applicable, enter the lot number(s) of the reference controls.

■ **NOTE:** If you selected Label/Lot Specific Unmixing in the Acquisition Preferences (see [page 82](#)), the software will search the library and experiment reference groups for reference controls that

have the same fluorescent tag, label, and lot information in order to use the corresponding control for unmixing.

Fluorescent Tag	Control Type	Label	Lot	Unstained
FITC	Cells			
PE	Cells			
PerCP	Cells			

■ **NOTE:** Use the red trash can icon to delete an individual tube from the reference group. This may be necessary if you wish to mix and match references acquired in this experiment with reference controls run in QC & Setup. Any stored controls you plan to use should be deleted from the reference group.

12 Click Save.

Once the reference group has been created, entries for each of the references will be displayed. Each of the reference group tubes/wells will have an icon (with the letter R) associated with it under the reference group.

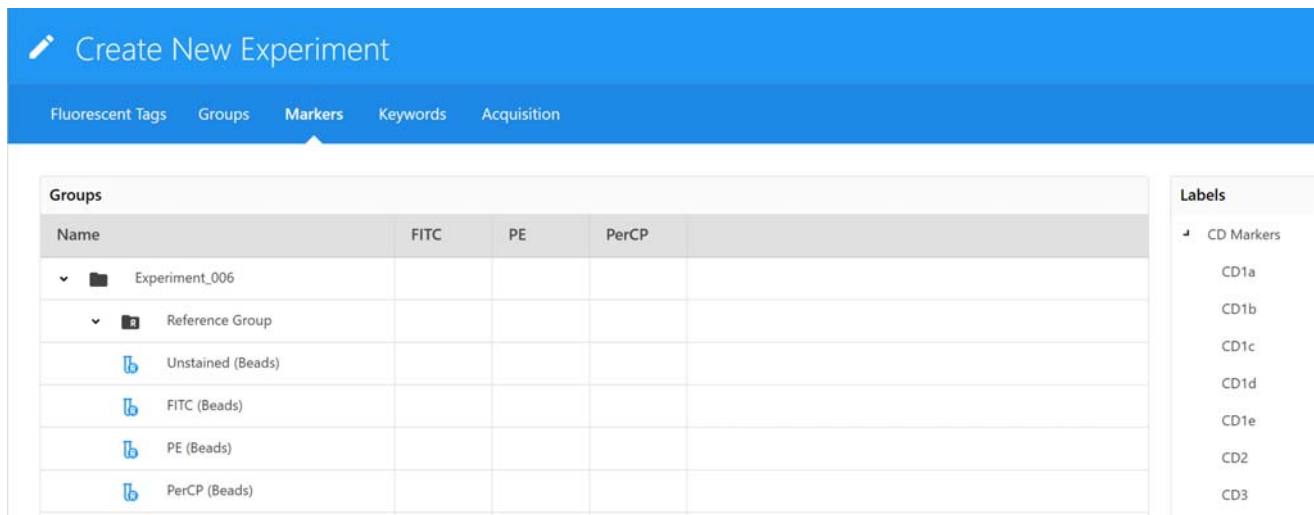
Group | Tube: 1 | Remove | Reference Group | Reference Group

- Experiment_006
 - Reference Group
 - Unstained (Beads)
 - FITC (Beads)
 - PE (Beads)
 - PerCP (Beads)
 - Group_001

13 If necessary, continue adding tubes/wells, click Next when all tubes are created.

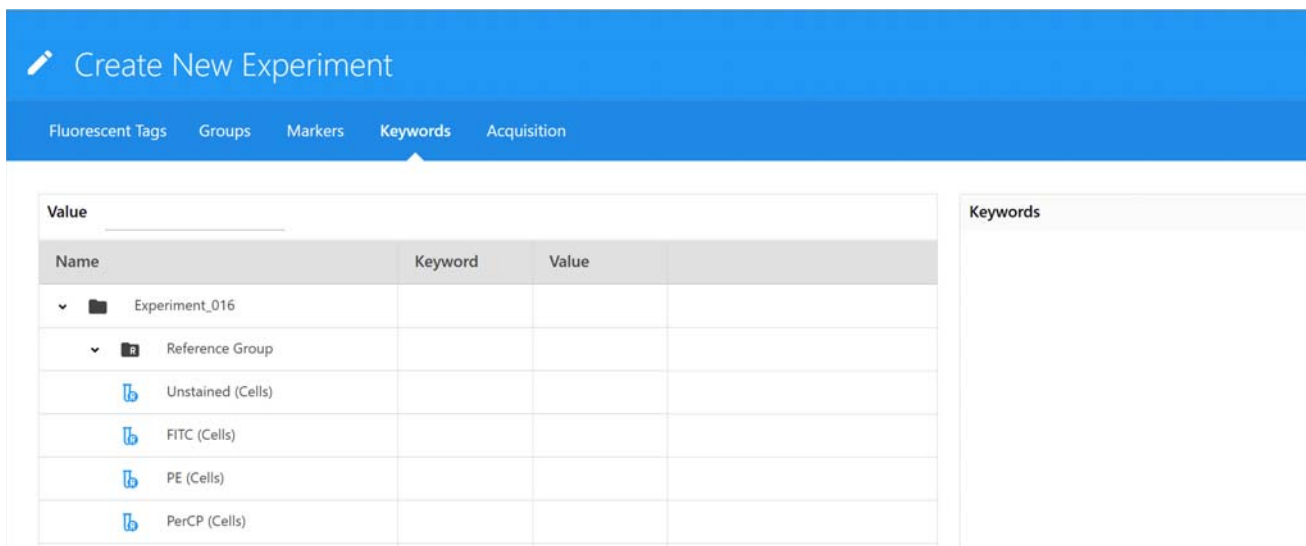
14 Add markers/labels to the remaining sample tubes before continuing. They can be chosen from the Labels list on the right, typed directly into the table, or copied and pasted. Labels can be added at the group or tube level and can be applied to multiple cells selected at once. Labels are required for reference controls if you selected Label/Lot Specific Unmixing in the Acquisition Preferences (see [page 82](#)). Click Next when all the tubes are labeled.

■ **NOTE:** If you selected Label/Lot Specific Unmixing, the software will search the library and experiment reference groups for reference controls that have the same fluorescent tag, label, and lot information in order to use the corresponding control for unmixing.



15 (Optional) Enter custom keywords and click Next.

Custom keywords can be added at the experiment, group, or tube level. You must define the custom keywords in the Library before you can add them to an experiment (see “Keywords” on page 78 for information). Drag and drop the keywords from the Keywords list on the right to the experiment, group, or tube and enter keywords values as needed. You can also copy and paste custom keywords across different tubes in this wizard.



16 Select the acquisition settings and worksheet(s). The worksheet menu lists all the worksheets for the given user.

- Select the Default Raw Worksheet (Raw) for the Reference Group and for the sample groups if you plan to perform post-acquisition unmixing.
- Select the Default Unmixed Worksheet (Unmixed) or any user-created unmixed worksheet for your sample groups if you are performing live unmixing. Worksheets can be selected at the experiment, group, or tube level.

Select the stopping gate, storage gate, number of events to record, stopping time (in seconds), or stopping volume (in μL). Acquisition stops when the first of the stopping criteria is met (time, volume, number of events). These criteria can be selected at the experiment, group, or individual level.

- If acquiring beads, we recommend collecting 5,000 singlet events.
- If acquiring cells, we recommend collecting 10,000 to 20,000 events of the desired population.

■ **NOTE:** If running in plate mode, when selecting worksheets and acquisition settings, select the top level (group or experiment) to apply the settings to all wells within the group or experiment.

■ **NOTE:** The number of events to acquire depends on the target population. For example, you may need to acquire 10,000 to 20,000 events to get 2,000 of the desired population. Approximately 1,000 to 2,000 events is needed in both the negative and positive populations of each control for accurate unmixing.

Name	Worksheet	Stopping Gate	Storage Gate	Events To Rec
Experiment_004				10,000
Reference Group	Default Raw Worksheet (Raw)	All Events	All Events	10,000
Unstained (Beads)	Default Raw Worksheet (Raw)	All Events	All Events	10,000
CD4 FITC (Beads)	Default Raw Worksheet (Raw)	All Events	All Events	10,000
CD8 PE (Beads)	Default Raw Worksheet (Raw)	All Events	All Events	10,000
CD3 PerCP (Beads)	Default Raw Worksheet (Raw)	All Events	All Events	10,000
Group_001	Default Unmixed Worksheet (Unmixed)	All Events	All Events	10,000
Tube_001	Default Unmixed Worksheet (Unmixed)	All Events	All Events	10,000

17 If running in plate mode, define the Loader settings. See [“Defining Loader Settings” on page 100](#).

18 Once the worksheet and stopping criteria have been defined, click Save and Open to open the new experiment.

To make any changes to the experiment, click Edit above the group/tube hierarchy.

19 To acquire controls and samples and perform live unmixing, see [“Live Unmixing” on page 60](#).

20 If running in Plate mode, calibrate the SIT. See [“Calibrating the SIT” on page 103](#).

Unmixing and Compensation

Spectral Unmixing

Spectral unmixing is an important concept to understand how data is generated and analyzed using the Aurora flow cytometer with SpectroFlo software. Spectral unmixing is used to identify the fluorescence signal for each fluorophore used in a given experiment.

Understanding Full Spectrum Flow Cytometry

Because fluorophores emit light over a range of wavelengths, optical filters are typically used to limit the range of frequencies measured by a given detector. However, when two or more fluorophores are used, the overlap in wavelength ranges often makes it impossible for optical filters to isolate light from a given fluorophore. As a result, light emitted from one fluorophore appears in a non-primary detector (a detector intended for another fluorophore). This is referred to as spillover. In conventional flow cytometry spillover can be corrected by using a mathematical calculation called compensation. Single-stained controls must be acquired to calculate the amount of spillover into each of the non-primary detectors.

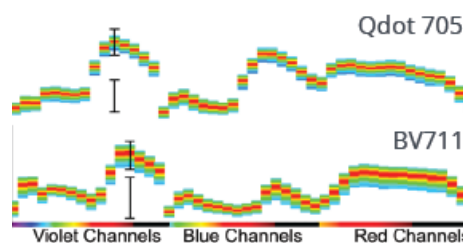
The Aurora's ability to measure a fluorophore's full emission spectra allows the system to use a different method for isolating the desired signal from the unwanted signal. The key to differentiate the various fluorophores is for those to have distinct patterns or signatures across the full spectrum. Because the system is looking at the full range of emission of a given fluorophore, and not only the peak emission, two dyes with similar emission but different spectral signatures can be distinguished from each other. The mathematical method to differentiate the signals from multiple fluorophores/dyes is called spectral unmixing and results in an unmixing matrix that is applied to the data. While not mathematically identical to conventional compensation, the overall principal is the same. Just as for compensation, single-stained controls, identified in SpectroFlo software as reference controls, are still necessary, as they provide the full fluorescence spectra information needed to perform spectral unmixing.

One advantage of unmixing is the ability to extract the autofluorescence of a sample and treat it as a separate parameter. This is especially useful when running assays with particles that have high autofluorescence and for which that high background has an impact in the resolution of the fluorescent signals. Per experiment, you can define one unstained control or multiple unstained controls (one per group), depending on whether the multicolor samples have the same or different autofluorescence signatures.

Spectrum plots from conventional spectrum viewer shows heavy overlap between Qdot 705 and BV711 peak emission spectra.



Spectrum plots from Aurora show distinct signatures for Qdot 705 and BV711.



Unmixing Workflows

Unmixing Overview

There are three unmixing workflows available in SpectroFlo software—two in the Acquisition module and one in the Extra Tools module:

- live unmixing during acquisition
- post-acquisition unmixing (in the Acquisition module)
- post-acquisition unmixing (in the Extra Tools module)

When data is acquired with live unmixing, references are acquired as raw data either in the experiment as part of the reference group or previously acquired in the QC & Setup module as reference controls. References for *all* fluorescent tags used in a given experiment must be present in the system in order for live unmixing of multicolor samples to occur. The live unmixing functionality allows you to visualize unmixed data during acquisition.

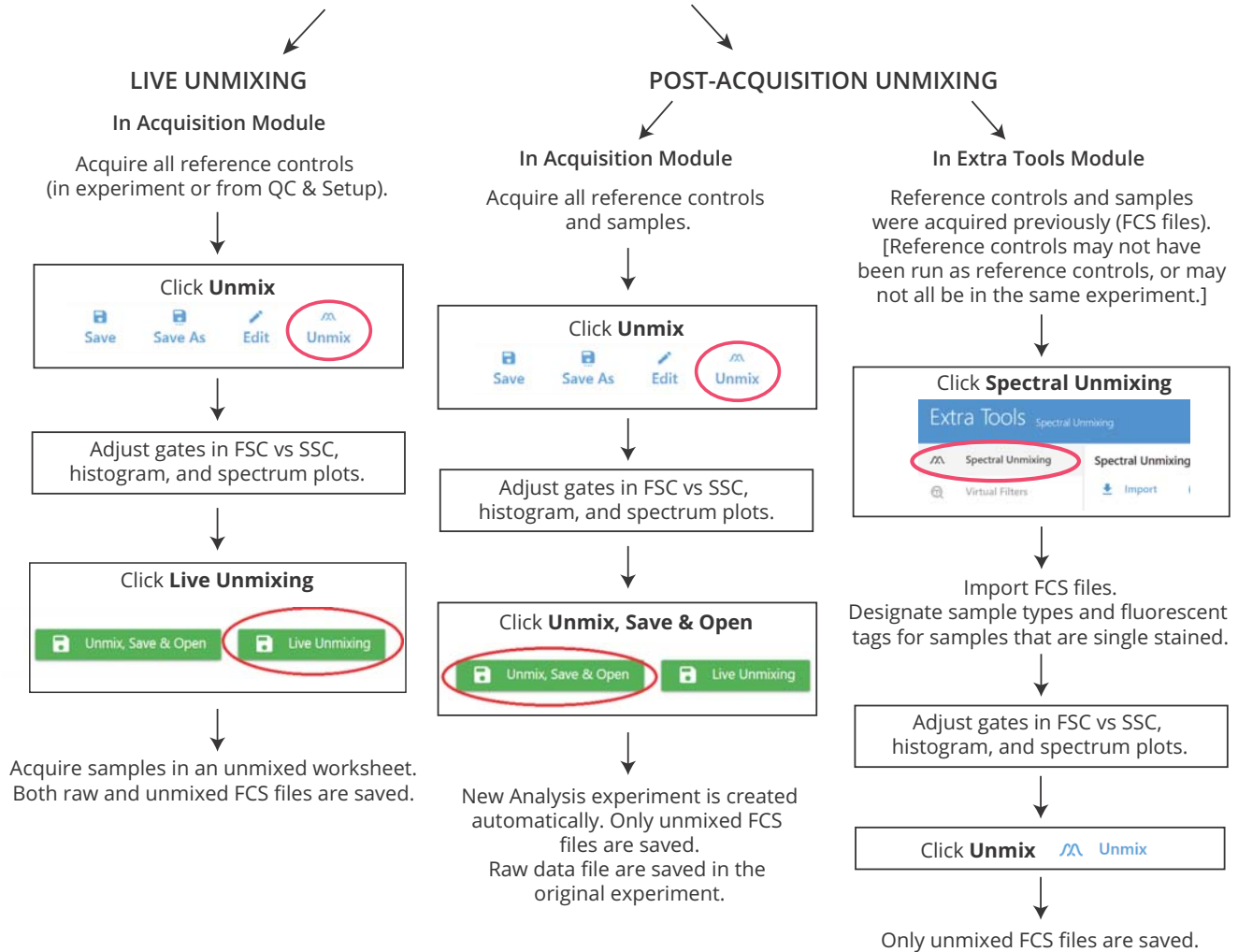
Reference Controls for Unmixing

Depending on when you unmix the data, you will use the following controls for unmixing.

Unmixing	Reference Controls
Live unmixing	Reference controls run in the experiment or reference controls run in QC & Setup
Post-acquisition unmixing in Acquisition module	Reference controls run in the experiment or reference controls run in QC & Setup
Post-acquisition unmixing in Extra Tools module	Any FCS files from samples run in any experiment

Multicolor samples can be acquired as raw data and unmixed post acquisition as well. This can be done in either the Acquisition module or the Extra Tools module.

Unmixing Workflows

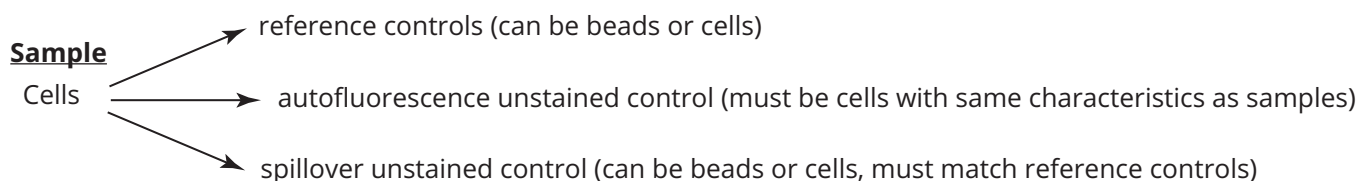


Negative/Unstained Controls

In addition to positive reference controls needed for spectral unmixing, an unstained control is also necessary to assess **autofluorescence**. The unstained control needs to be of the same type and prepared in the same way as the samples, as this will ensure accurate unmixing and autofluorescence extraction, if desired. Ideally, your reference controls, unstained control, and samples will all be the same sample type and prepared in the same way.

In addition to assessing autofluorescence, **fluorescence spillover** must also be determined. To correct for spillover, the unstained autofluorescence control can be used if it matches the sample and reference control type. However, if your reference controls do not match your sample type and do not contain a negative population in each tube (have only positive peaks), you must use a separate spillover unstained control that matches your reference control type.

Controls



Live Unmixing

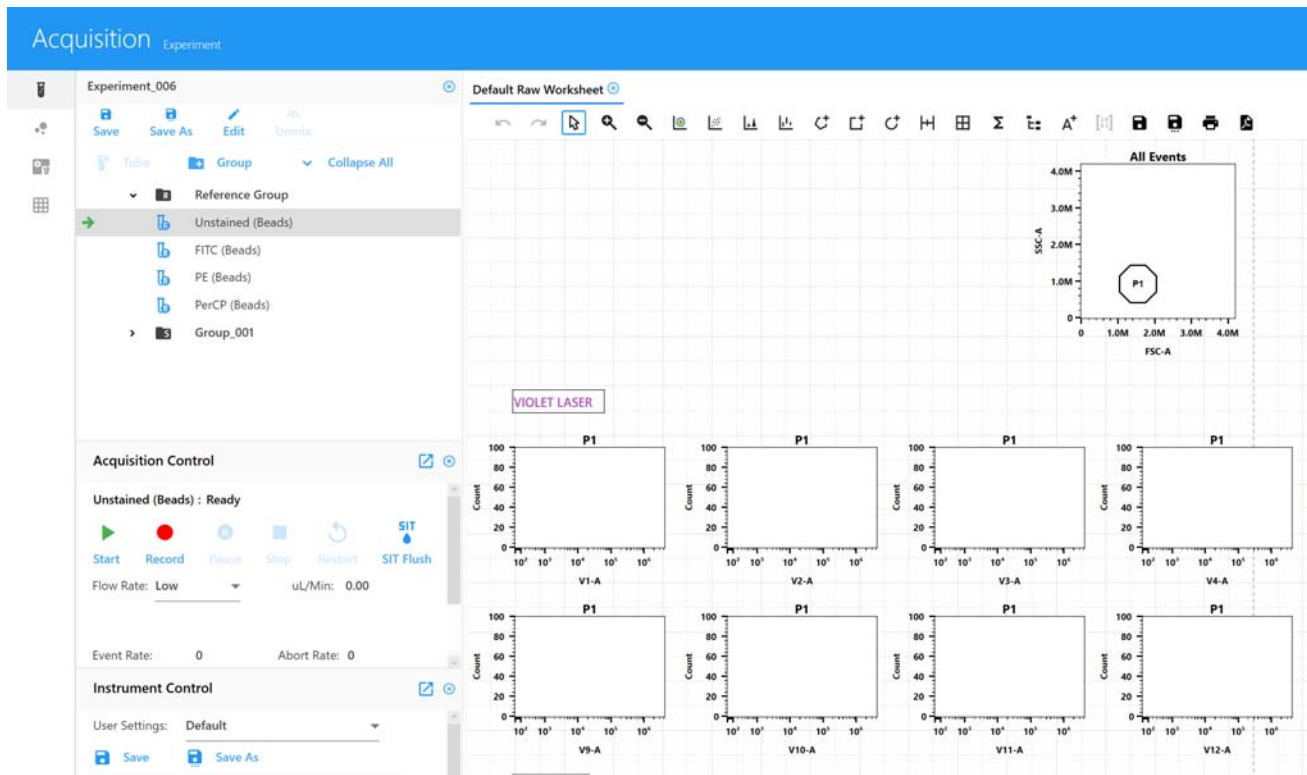
Samples can be unmixed during acquisition. Live unmixing can be performed with the reference group acquired during the experiment, the reference controls (run during QC & Setup and stored in the system), or a combination of both.

For each sample tube that is live unmixed, two FCS files are generated, one that is composed of raw data and one that is composed of unmixed data.

Live unmixed data can be analyzed in unmixed worksheets in the Acquisition module. Unmixed worksheets are different from raw worksheets, as they only display fluorescence information categorized into the defined fluorescent tags for each of the experiments.

To Perform Live Unmixing

- 1 Create a new experiment with fluorescent tags defined. Create a reference group in the experiment with the fluorescent tags, if there are any that have not already been stored as reference controls. See “Creating a New Experiment” on page 50 for details.



- 2 To view the data for the reference control tubes, make sure CytekAssaySetting is selected, then click Start. If necessary, use the Instrument Controls to adjust the settings so that all events are on scale. View all the controls, as well as the multi-color tube, and make any instrument adjustments to ensure populations are on scale before you begin recording.

To edit the acquisition criteria, click Edit at the experiment level and select the Acquisition tab. Or, to edit the properties of a single tube, right-click a tube and select Tube Properties.

■ **NOTE:** Keep in mind the more events you acquire, the longer it takes to unmix the data.

- 3 Click Record when you are ready to begin acquisition. Acquisition stops when the first stopping criterion is met.

■ **NOTE:** If necessary, you can pause to change the flow rate.

- When all reference controls are acquired, click Unmix in the upper-left toolbar.
- For the unstained controls, we recommend selecting Use Control from Experiment if unmixing with controls you acquired in the experiment.
- If necessary, for the stained controls, select Use Control from Library if unmixing with reference controls run in QC & Setup.

Checkmarks appear for those controls coming from QC & Setup. The checkbox is only active if reference controls for those fluorescent tags are already saved with the reference controls from the QC & Setup module.

- Click Next.
- Use the Identify Positive/Negative Populations tab to include the positive and negative populations for each fluorescent tag in the appropriate gate.

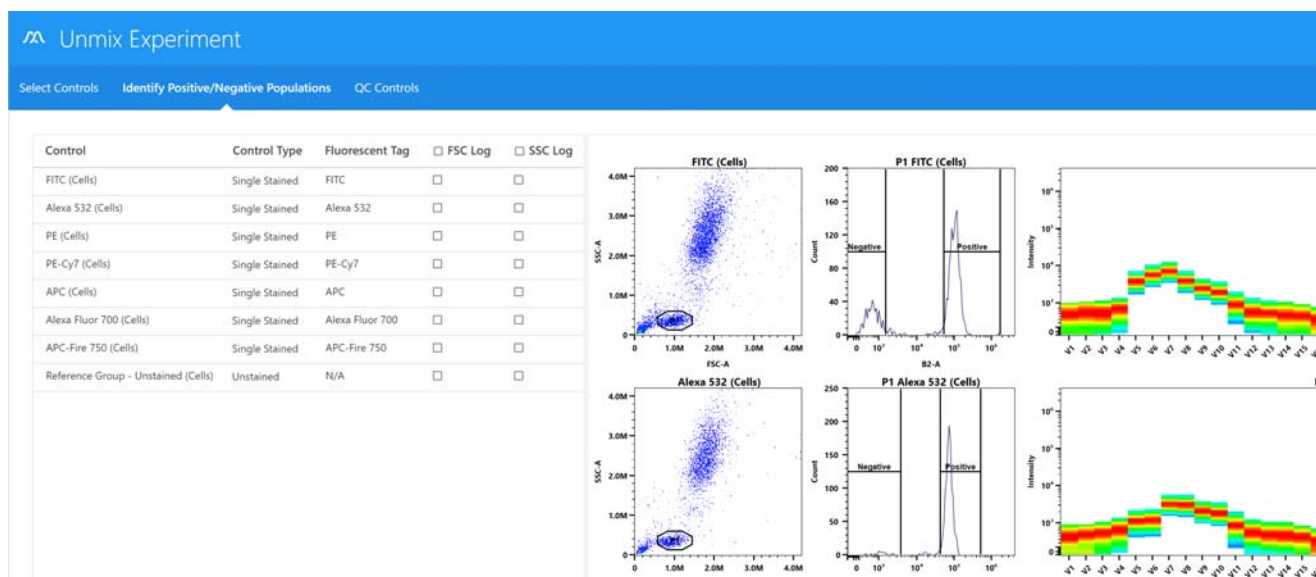
Only the data plots for the samples you acquired are displayed, not for reference controls that you chose to use from the library.

■ **NOTE:** If you need to set the FSC and/or SSC axis to a log scale, select the Log checkbox.

You may find it more efficient to view the data in columns—adjust the gates in the FSC vs SSC plots first, then adjust the histogram gates, finally adjust the gates on the spectrum plots.

- Move the polygon gate in the FSC vs SSC plot on the left to include the singlet population. Hold down Ctrl to move all the polygon gates at once.
- Move the positive interval gate in the histogram to include the positively stained population. Move the negative interval gate to include the negative population.
- Move the interval gate on the spectrum plot on the right to select the channel that exhibits the brightest fluorescence intensity. This channel is the peak emission channel for the fluorescent tag.

■ **NOTE:** If one of the controls is questionable or does not contain sufficient data, you can reacquire it or append to it, then unmix again.



- (Optional) To see how the reference controls run in the experiment compare to the benchmark reference controls, click Next.

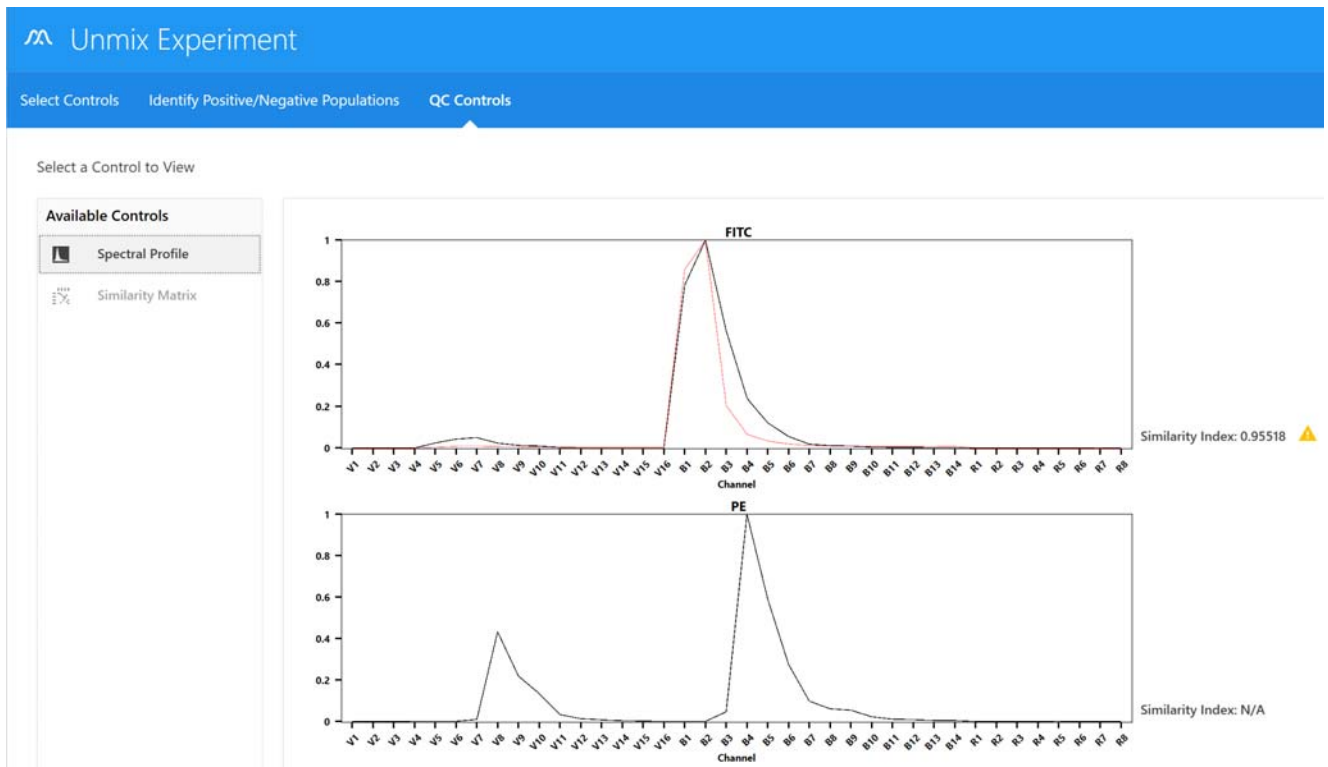
■ **NOTE:** For information on creating benchmarks, see [“Setting Reference Controls as Benchmarks for Reference Control QC”](#) on page 37.

Two options allow you to view how the two reference controls compare—spectral profile and similarity index.

- Spectral Profile displays the emission spectrum of the unmixing controls against benchmark spectra designated by the user. The benchmark reference control spectra appear in red and the reference controls appear in black.

A similarity index appears to the right of the plots. If the value is below 0.97, it will be flagged with a yellow warning symbol. This indicates a mismatch of the unmixing control spectra with the benchmark spectra. If the similarity index falls below this value, it is imperative to check the unmixing control against the reference spectra provided in the fluorochrome guideline found in the Help menu. See [“Similarity Matrix”](#) below.

If no benchmark control is established for a particular dye, that plot will only display a black line that represents the spectra of the unmixing control. The similarity index will display N/A.



- Similarity Matrix displays a similarity index matrix and a complexity index value.

The similarity index is a number between 0-1 that measures how closely the unmixing control's spectral signature matches the benchmark control's spectral signature. Click View Similarity Index above the matrix to display the indexes for each dye. The Similarity Matrix will display the similarity index for each dye against itself and all the other dyes to be unmixed in the experiment.

The complexity index is a measure of how distinguishable a collection of spectral signatures are from each other when unmixed together. It calculates this by looking at the ratio of the similarity index of the worst overlapping combination of signatures to the best overlapping

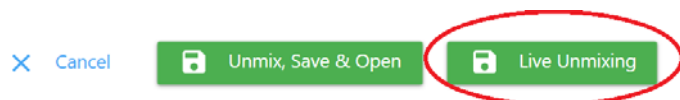
combination of signatures by looking at the ratio of the worst overlapping combination to the best overlapping combination.

The screenshot shows the 'Unmix Experiment' interface with the 'QC Controls' tab selected. On the left, 'Available Controls' includes 'Spectral Profile' and 'Similarity Matrix'. The 'Similarity Matrix' is displayed as a heatmap with the following data:

	FITC	Alexa 532	PE	PE-Cy7	APC	Alexa Fluor 700	APC-Fire 750
FITC	1	0.5	0.2	0.01	0	0	0.01
Alexa 532	0.5	1	0.67	0.02	0	0	0
PE	0.2	0.67	1	0.02	0.01	0	0.01
PE-Cy7	0.01	0.02	0.02	1	0.02	0.07	0.18
APC	0	0	0.01	0.02	1	0.45	0.16
Alexa Fluor 700	0	0	0	0.07	0.45	1	0.37
APC-Fire 750	0.01	0	0.01	0.18	0.16	0.37	1

Complexity Index: 2.77

10 Click Live Unmixing.



11 The wizard closes and the experiment reappears. The reference group now has the unmixed icon to the left of the tube(s). Select an unmixed worksheet to view the unmixed data.

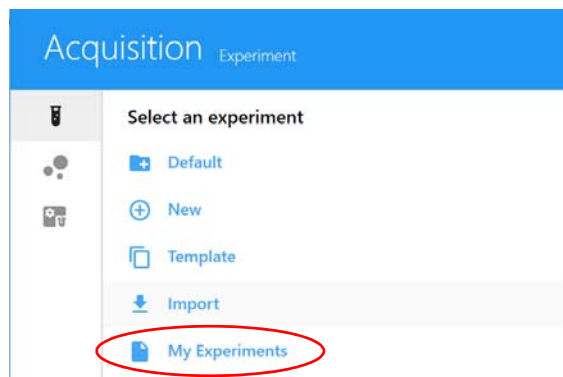
The screenshot shows the 'Acquisition' interface for experiment '*7C Demo Panel 20190826'. The 'Default Raw Worksheet' is active. On the left, a tree view shows the following structure:

- Reference Group (with an unmixed icon 'R')
- Group_001
 - Tube_001
 - Tube_002 (highlighted with a green arrow)

On the right, a plot area titled 'All Events' shows a single data point labeled 'P1' at approximately 1.0M on the y-axis (PSCH).

12 Select the sample tube you wish to acquire. The green arrow indicates the tube is selected. Click Start, then Record.

Use My Experiments to open experiments you ran if you wish to review the data or acquire more samples.



FCS files are stored in the Export folder by default, or the folder you set as the default. See [“Storage Preferences” on page 88](#) for information. FCS files for live unmixed data are saved as both raw data and unmixed data.

Alternately, you can click My Experiments, select the experiments you want to export, right-click and select Export. This will export the entire experiment as a ZIP file with all of the FCS files and worksheet templates contained inside. This experiment can be imported into other instances of SpectroFlo software, or unzipped to access the FCS files for analysis using other analysis software.

Post-Acquisition Unmixing

Samples can be acquired as raw data and then unmixed after acquisition is complete. This can be done through two methods:

- post-acquisition unmixing in the Acquisition module (see below)
- post-acquisition unmixing in the Extra Tools module (see [page 67](#))

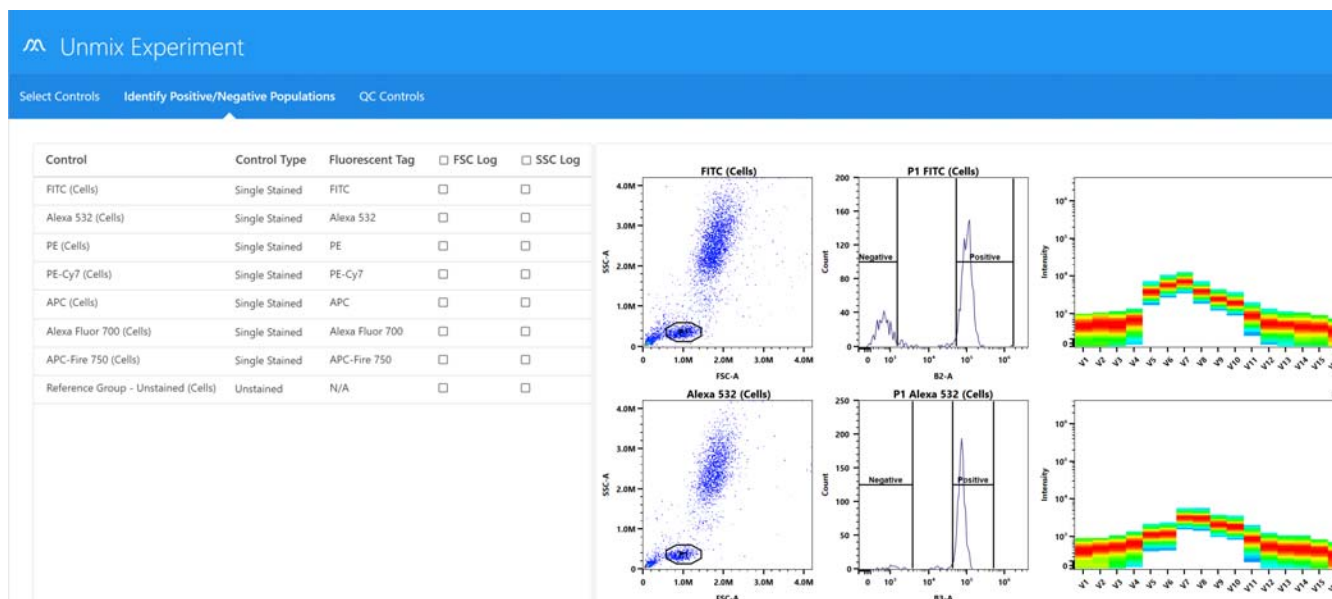
Post-Acquisition Unmixing in the Acquisition Module

The unmixing wizard in the Acquisition module limits reference controls to those coming from the reference group in the experiment or reference controls run in QC & Setup.

To perform post-acquisition unmixing in the Acquisition module, perform the same workflow as live unmixing except the following:

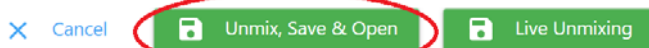
- 1 Acquire all reference control tubes and sample tubes prior to selecting the Unmix button in the upper-left pane.
- 2 You may find it more efficient to view the data in columns—adjust the gates in the FSC vs SSC plots first, then adjust the histogram gates. Finally, examine the spectra plots.
 - a. Move the polygon gate in the FSC vs SSC plot on the left to include the singlet population. Hold down Ctrl to move all the polygon gates at once.
 - b. Move the positive interval gate in the histogram to include the positively stained population. Move the negative interval gate to include the negative population.

- c. Move the interval gate on the spectrum plot on the right to select the channel that exhibits the brightest fluorescence intensity. This channel is the peak emission channel for the fluorescent tag.

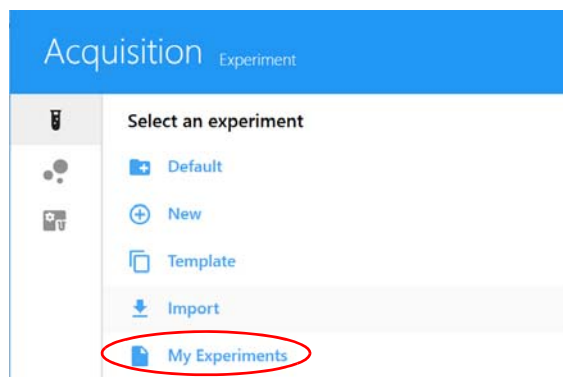


- 3 Click Unmix, Save & Open.

A new experiment opens with a new unmixed worksheet.



Use My Experiments to open experiments you ran, if you wish to review the data or acquire more samples.



FCS files are stored in the Export folder by default, or the folder you set as the default. See “Storage Preferences” on page 88 for information. FCS files for post-acquisition unmixed data are saved as unmixed data only.

Post Acquisition Unmixing in the Extra Tools Module

When performing post-acquisition unmixing in the Extra Tools module, you can pick and choose which FCS files to unmix (for example, controls coming from different experiments, reference controls run during QC & Setup, or single-stained controls that were not run as part of the reference group).

FCS files can be designated into three categories:

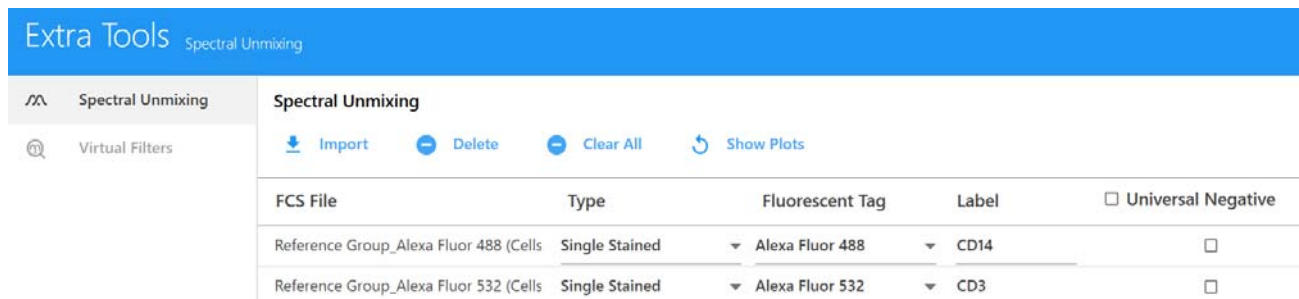
- Single Stained
- Unstained
- Sample

■ **NOTE:** There must be at least one single-stained FCS file and one unstained FCS file in the file list. Otherwise, unmixing cannot be performed.

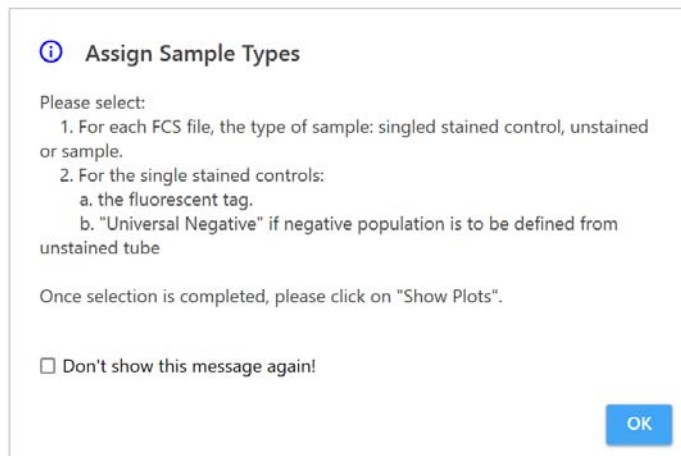
In addition, raw FCS files can also be conventionally compensated in this module through the Virtual Filters tab. This function can simulate the presence of filters and can compensate data using conventional compensation methods (see “Virtual Filters” on page 70).

To Unmix Raw Data Files:

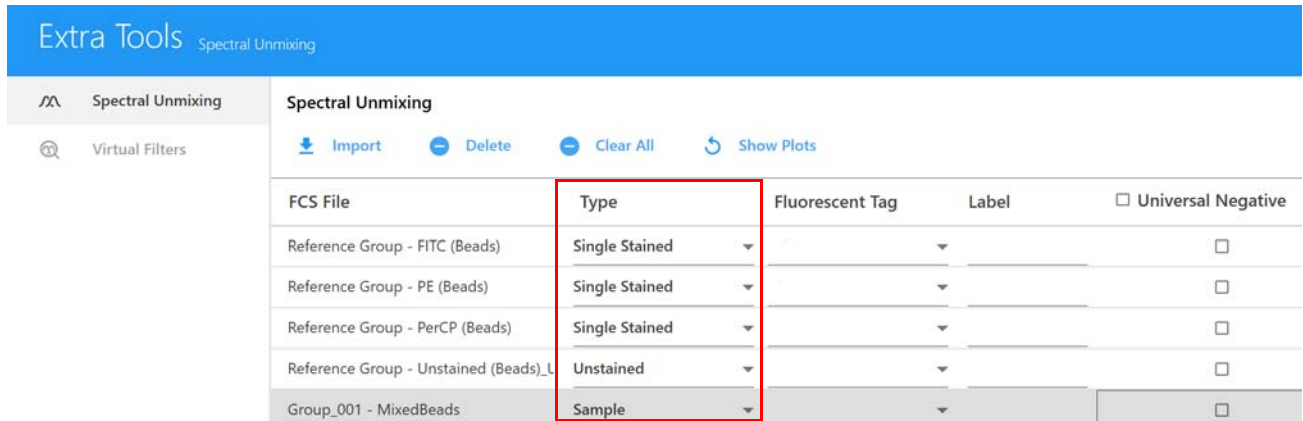
- 1 Select Spectral Unmixing from the Extra Tools module.
- 2 Click Import to import raw FCS files for unmixing.



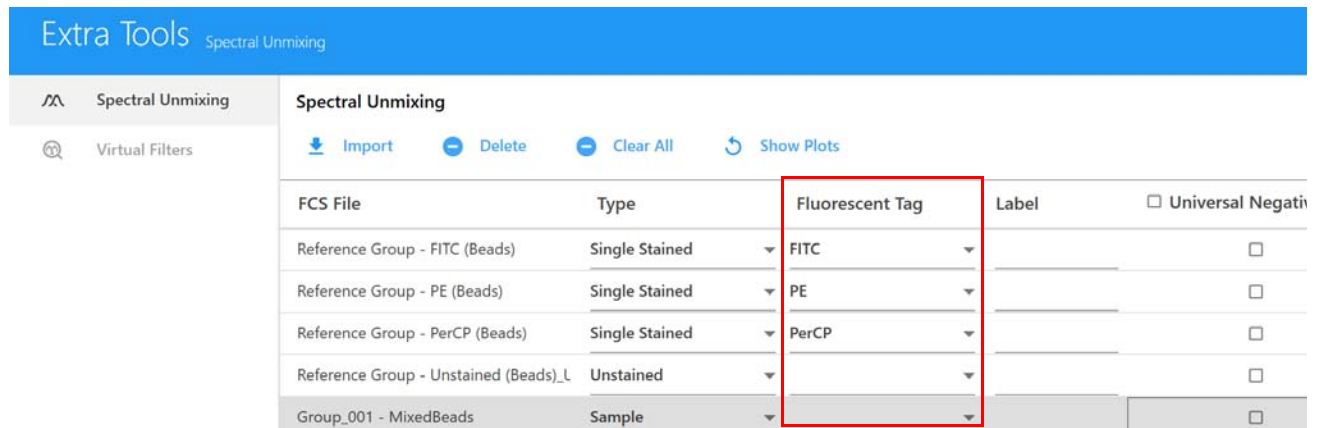
- 3 Select the files. Select multiple files using either the Shift or Ctrl key. Click **Open**.
- 4 Upon importing, a dialog box on how to assign sample types appears. Read the instructions and click OK.



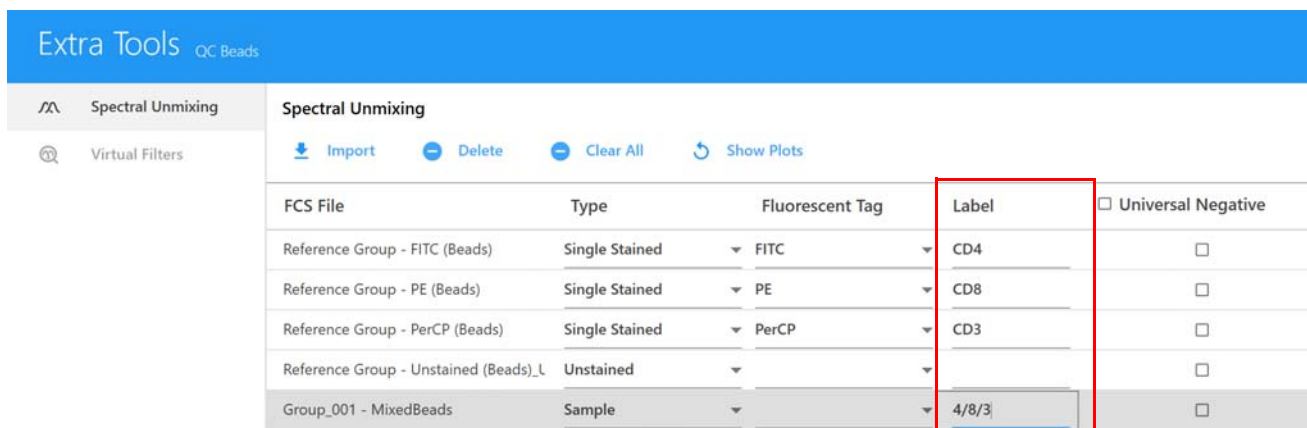
- Once FCS files have been imported, select the sample type for each FCS file as Single Stained, Unstained, or Sample. The software will automatically designate the type based upon the file name. You can manually modify these if the automatic designation is incorrect.



- FCS files designated as single-stained will require a fluorescent tag designation to specify what reference spectrum will be provided for unmixing.

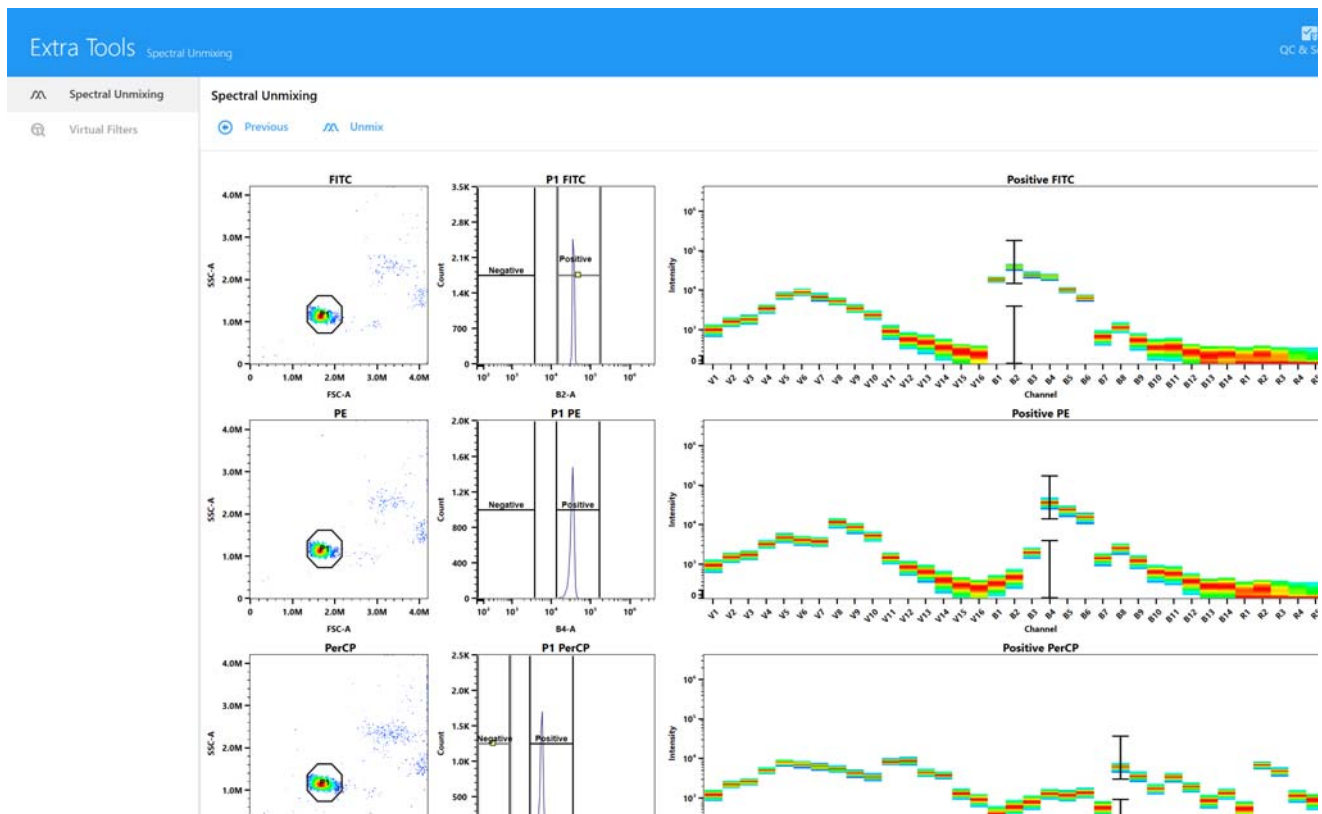


- Enter a label for each single-stained control and sample.

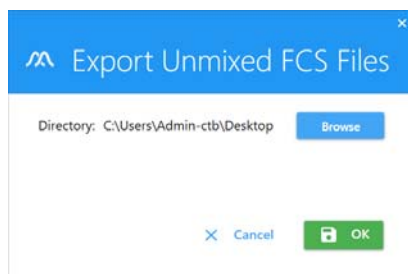


- Select Universal Negative for single-stained FCS files that do not contain a negative population, and the unstained control will be used for the negative population. In the bottom left of the screen, check whether Auto Fluorescence will be used as a fluorescent tag.

- 9 Click Show Plots to display the data in the FSC vs SSC plot, peak emission channel histogram, and spectrum plots.
- 10 The positive and negative populations need to be identified through the appropriate placement of the existing gates. Click OK to adjust the gates.
 - a. Move the polygon gate in the FSC vs SSC plot to include the singlet population.
 - b. Move the interval gate in the histogram labeled *Positive* to include the positively stained population. Move the interval gate in the histogram labeled *Negative* to include the negative population. Do not adjust the negative gate when using the Universal Negative.
 - c. Move the interval gate on the spectrum plot on the right to select the channel that exhibits the brightest fluorescence intensity. This channel is the peak emission channel for the fluorescent tag.



- 11 Click Unmix.
- 12 Select the directory to which the unmixed FCS files are exported or leave the default. Click OK.

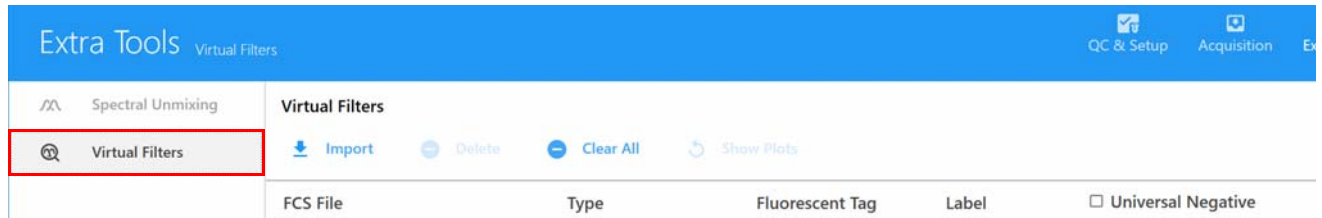


These FCS files can then be imported to an experiment for analysis or analyzed using third-party software.

Virtual Filters

The Virtual Filters option in the Extra Tools module allows you to compensate raw FCS data using conventional compensation methods.

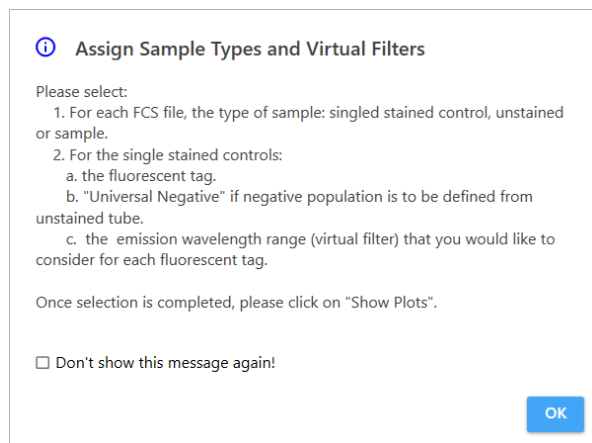
- 1 Click the Virtual Filters tab in the Extra Tools module.



- 2 Click Import to import raw FCS files for virtual filter analysis.

These FCS files can be single-stained reference controls, unstained controls, and/or sample files. However, you must include an unstained control FCS file.

- 3 Upon importing, a dialog box on how to assign sample types appears. Read the instructions and click OK.



- 4 Once FCS files have been imported, the sample type for each FCS file needs to be designated as Single Stained, Unstained, or Sample. The software will automatically designate the type based upon the file name. You can manually modify these if the automatic designation is incorrect.
- 5 FCS files designated as single stained require a fluorescent tag designation. Select the fluorescent tag for each single-stained sample.

If there is no negative population in the single-stained FCS file(s), select Universal Negative, and the unstained control will be used for the negative population.

The virtual filter is automatically assigned by the software based upon the fluorescent tag designation. (Optional) To increase the bandwidth of the virtual filter, use the channel pull-down menus to select the desired range. See the following table for wavelength ranges.



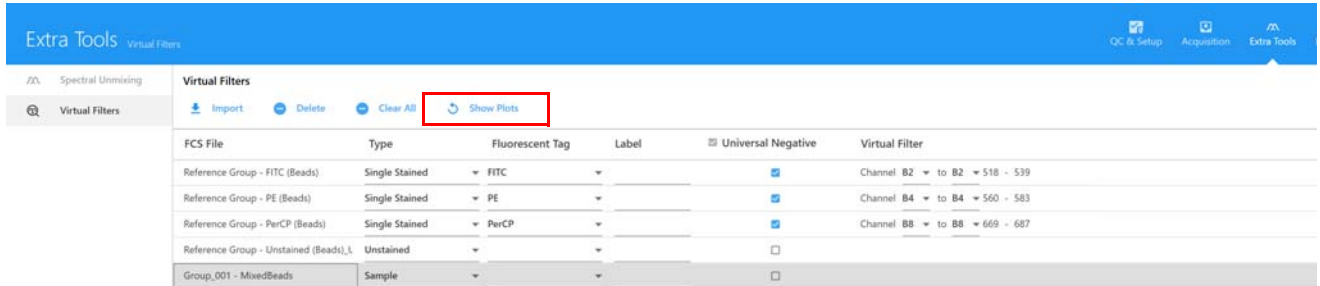
The following table shows the system's filter bandwidths.

Laser	Channel	Center Wavelength (nm)	Bandwidth (nm)	Wavelength Start (nm)	Wavelength End (nm)
Ultraviolet	UV1	373	15	365	380
	UV2	388	15	380	395
	UV3	428	15	420	435
	UV4	443	15	436	451
	UV5	458	15	451	466
	UV6	473	15	466	481
	UV7	514	28	500	528
	UV8	542	28	528	556
	UV9	582	31	566	597
	UV10	613	31	597	628
	UV11	664	27	651	678
	UV12	692	28	678	706
	UV13	720	29	706	735
	UV14	750	30	735	765
	UV15	780	30	765	795
	UV16	812	34	795	829
Violet	V1	428	15	420	435
	V2	443	15	436	451
	V3	458	15	451	466
	V4	473	15	466	481
	V5	508	20	498	518
	V6	525	17	516	533
	V7	542	17	533	550
	V8	581	19	571	590
	V9	598	20	588	608
	V10	615	20	605	625
	V11	664	27	651	678
	V12	692	28	678	706
	V13	720	29	706	735
	V14	750	30	735	765
	V15	780	30	765	795
	V16	812	34	795	829

Laser	Channel	Center Wavelength (nm)	Bandwidth (nm)	Wavelength Start (nm)	Wavelength End (nm)
Blue	B1	508	20	498	518
	B2	525	17	516	533
	B3	542	17	533	550
	B4	581	19	571	590
	B5	598	20	588	608
	B6	615	20	605	625
	B7	661	17	653	670
	B8	679	18	670	688
	B9	697	19	688	707
	B10	717	20	707	727
	B11	738	21	728	749
	B12	760	23	749	772
	B13	783	23	772	795
	B14	812	34	795	829
Yellow Green	YG1	577	20	567	587
	YG2	598	20	588	608
	YG3	615	20	605	625
	YG4	661	17	653	670
	YG5	679	18	670	688
	YG6	697	19	688	707
	YG7	720	29	706	735
	YG8	750	30	735	765
	YG9	780	30	765	795
	YG10	812	34	795	829
Red	R1	661	17	653	670
	R2	679	18	670	688
	R3	697	19	688	707
	R4	717	20	707	727
	R5	738	21	728	749
	R6	760	23	749	772
	R7	783	23	772	795
	R8	812	34	795	829

6 (Optional) Select a label for the single-stained controls and samples.

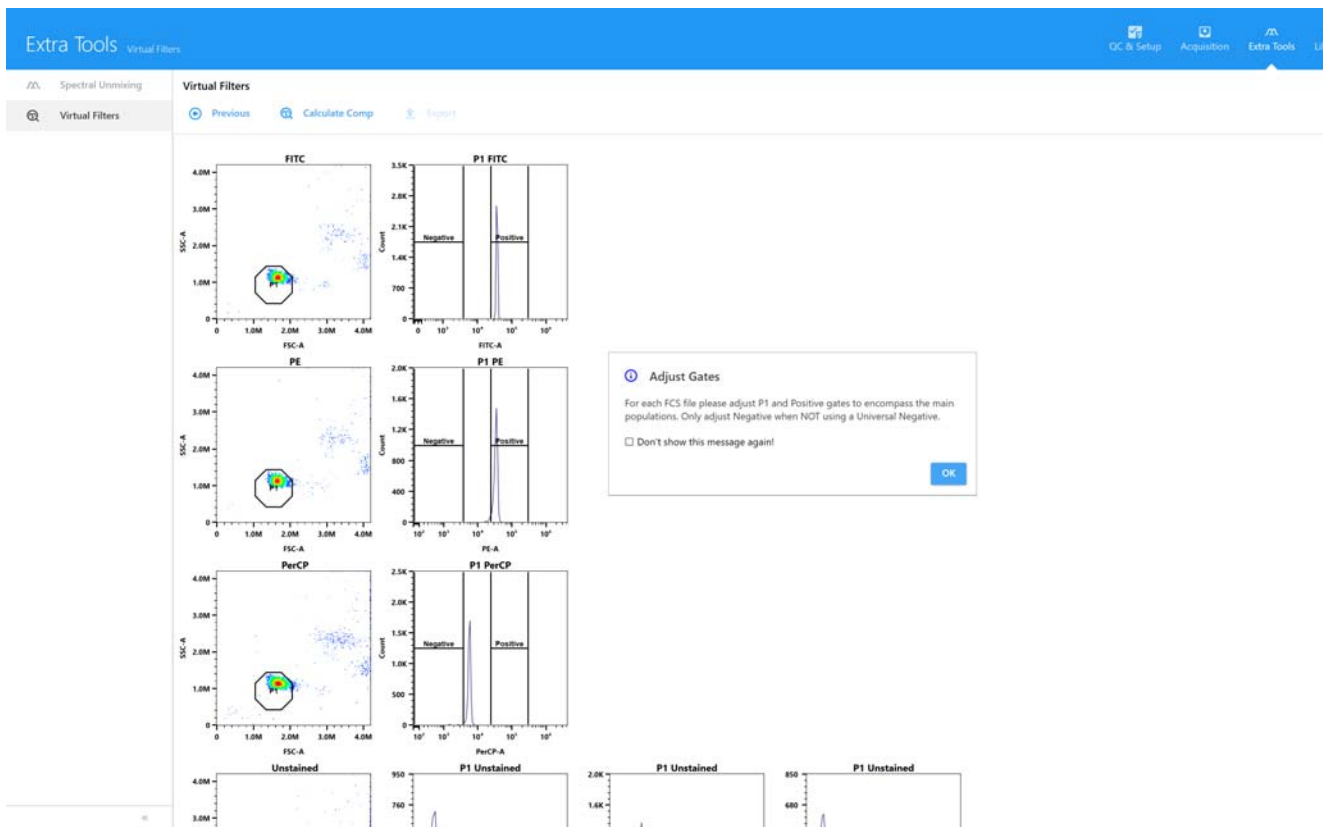
7 Click Show Plots to display the plots.



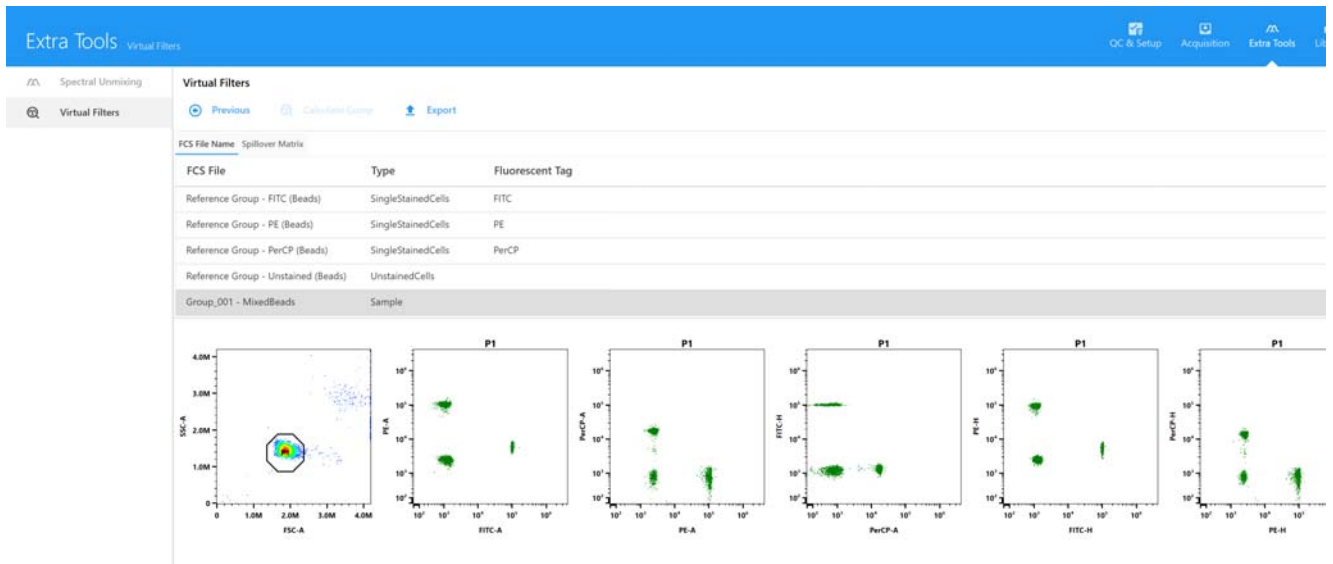
The data is displayed in the FSC vs SSC plot and fluorescent tag histogram plot.

- 8 The positive and negative populations need to be identified through the appropriate placement of the gates. Click OK to adjust the gates.
- Move the polygon gate in the FSC vs SSC plot to include the singlet population. Hold down Ctrl to move all the polygon gates at once.
 - Move the interval gate in the histogram labeled *Positive* to include the positively stained population. Move the interval gate in the histogram labeled *Negative* to include the negative population. Do not adjust the negative gate when using the Universal Negative.

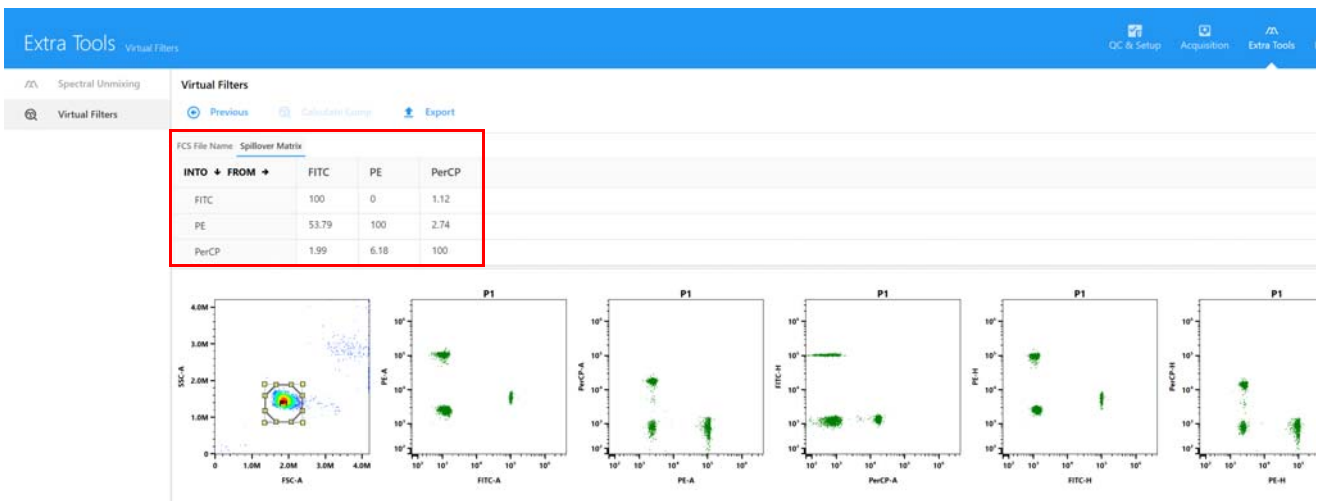
The histogram x-axes are labeled with the fluorescent tag instead of the channel/detector.



- Click Calculate Comp once gates have been set correctly. The conventionally compensated data is displayed. To view the data for a specific FCS files, select the file.



The spillover matrix is also calculated. Click Spillover Matrix to view the spillover values.



- Click Export and select the location where you wish to export the conventionally compensated data. The files are exported to a folder named Compensated followed by the current date and time. Files can then be imported back into an experiment to analyze in SpectroFlo software.

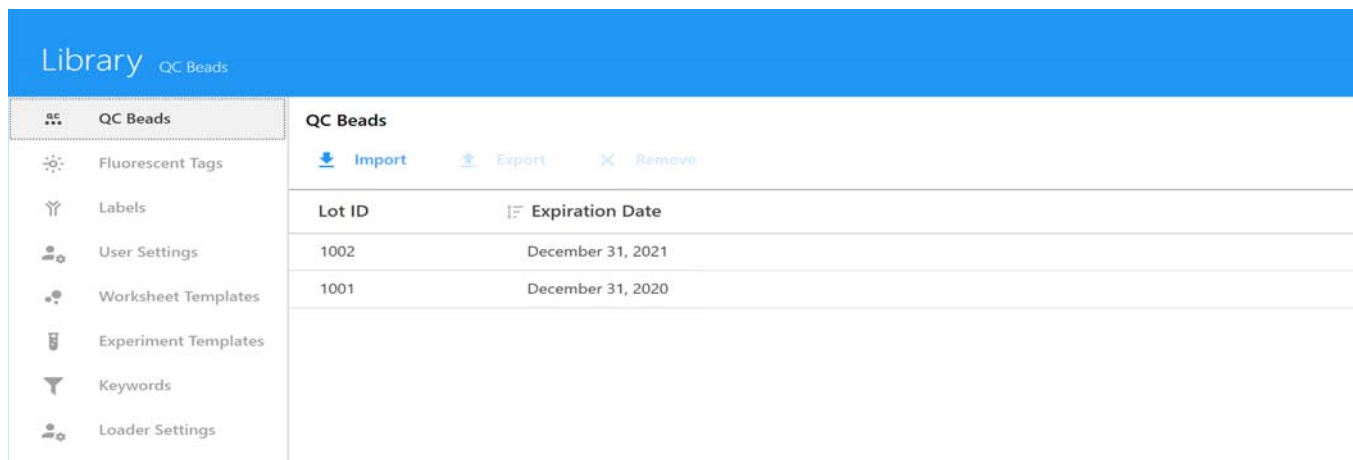
Library, Preferences, and Users

Library

The library contains information for various elements used for the experiments. Information saved in the library includes SpectroFlo QC bead lots, fluorescent tags, labels, user settings, worksheet templates, experiment templates, keywords, and Loader settings. Information stored in the library can be saved, exported, and imported for reuse.

QC Beads

SpectroFlo QC bead lot IDs and expiration dates can be imported, exported, or removed from the library. The QC bead lot for the beads used for Daily QC must be saved in the library.



The screenshot shows the 'Library' interface with a sub-section for 'QC Beads'. On the left is a navigation menu with options: QC Beads (selected), Fluorescent Tags, Labels, User Settings, Worksheet Templates, Experiment Templates, Keywords, and Loader Settings. The main area displays a table titled 'QC Beads' with 'Import', 'Export', and 'Remove' buttons. The table has two columns: 'Lot ID' and 'Expiration Date'.

Lot ID	Expiration Date
1002	December 31, 2021
1001	December 31, 2020

Fluorescent Tags

Fluorescent tags are the designation given to each distinct fluorescent molecule that can be detected by the system. This includes for example, fluorophores, fluorescent proteins, and fluorescent viability dyes. Each unique fluorophore run on the instrument must be given a fluorescent tag name.

By default, several groups of fluorescent tags are pre-installed with the software—Blue Laser, Red Laser, Violet Laser, Yellow Green Laser, Ultraviolet Laser, Fluorescent Proteins, and Viability Dyes.

These groups contain the most commonly used fluorophores excited by the system's on-board lasers. Additional tags can be added to these groups.

Name	Created By	Date Created	Date Modified	Name	Emission Wavelength
Blue Laser	System	August 20, 2019 - 10:14 AM	August 20, 2019	BB515	515
Red Laser	System	August 20, 2019 - 10:14 AM	August 20, 2019	Alexa Fluor 488	520
Violet Laser	System	August 20, 2019 - 10:14 AM	August 20, 2019	FITC	520
Fluorescent Proteins	System	August 20, 2019 - 10:14 AM	August 20, 2019	Alexa 532	554
Viability	System	August 20, 2019 - 10:14 AM	August 20, 2019	PE	576
				PE-Dazzle594	607

You can create groups or individual fluorescent tags by selecting New. Both groups and individual fluorescent tags can be imported or exported.

To edit the properties of a fluorescent tag that you added, select the fluorescent tag of interest and select Edit. Properties that can be edited include fluorescent tag name, laser excitation wavelength, emission wavelength, and display name. The default tags that are included with the software cannot be edited or deleted. However, they can be exported and imported across different systems running SpectroFlo software, v2.2.

If the fluorophore is known by another name or identified by a different spelling, those additional names or spellings can be added as synonyms.

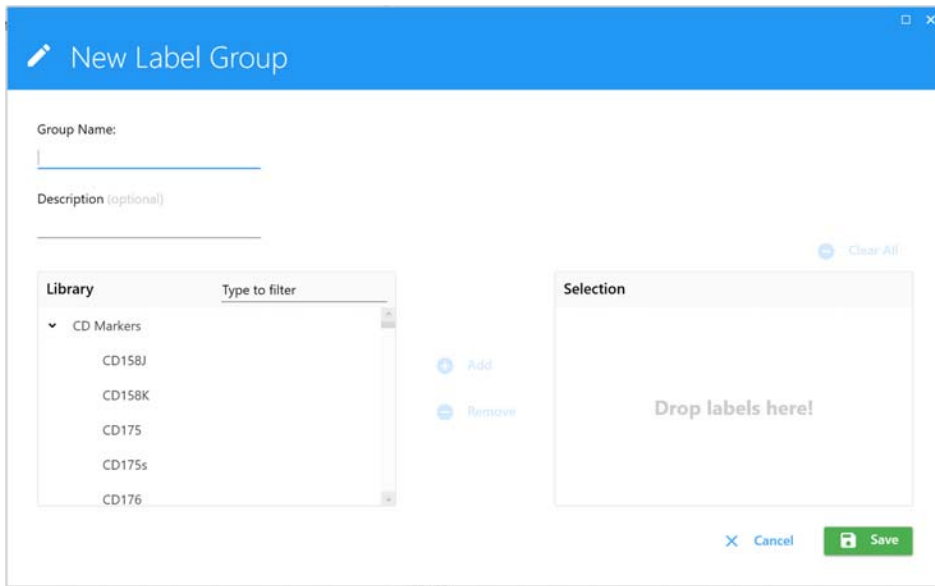
Any groups that you created can also be edited in this window. The default groups cannot be edited or deleted.

Labels

Fluorescent tags can be conjugated or attached to proteins that can specifically bind to other proteins on the cell surface or within the cytoplasm. They can also be inherently fluorescent, such as fluorescent proteins that can be fused to a variety of cellular proteins using molecular cloning techniques. The proteins that are either bound or attached to fluorescent tags can be designated as labels. The software comes with an initial set of pre-installed labels that are categorized as CD Markers, Chemokines, Chemokine Receptors, and Cytokines. Additional labels can be added by using **+** Add in the right pane.

Name	Created By	Date Created	Date Modified	Label
CD Markers	System	July 13, 2018 - 10:43 AM	July 13, 2018 - 10:43 AM	CD1a
Chemokine	System	July 13, 2018 - 10:43 AM	July 13, 2018 - 10:43 AM	CD1b
Chemokine Receptors	System	July 13, 2018 - 10:43 AM	July 13, 2018 - 10:43 AM	CD1c
Cytokines	System	July 13, 2018 - 10:43 AM	July 13, 2018 - 10:43 AM	CD1d
				CD1e

New label groups and individual labels can be created by clicking New. Label groups can also be imported and exported for use on other systems. The default labels cannot be edited or deleted.



User Settings

User Settings are the set of gain settings, threshold, and signal type for all detector channels. The date when it was created, as well as the name of user who created it are also saved. The settings are saved from the Instrument Control Pane in the Acquisition module. The name and description can be modified in this tab. Administrators can check the Shared checkbox to allow all user to see and use the settings and worksheets created by other users. These settings can be saved from the Acquisition screen.

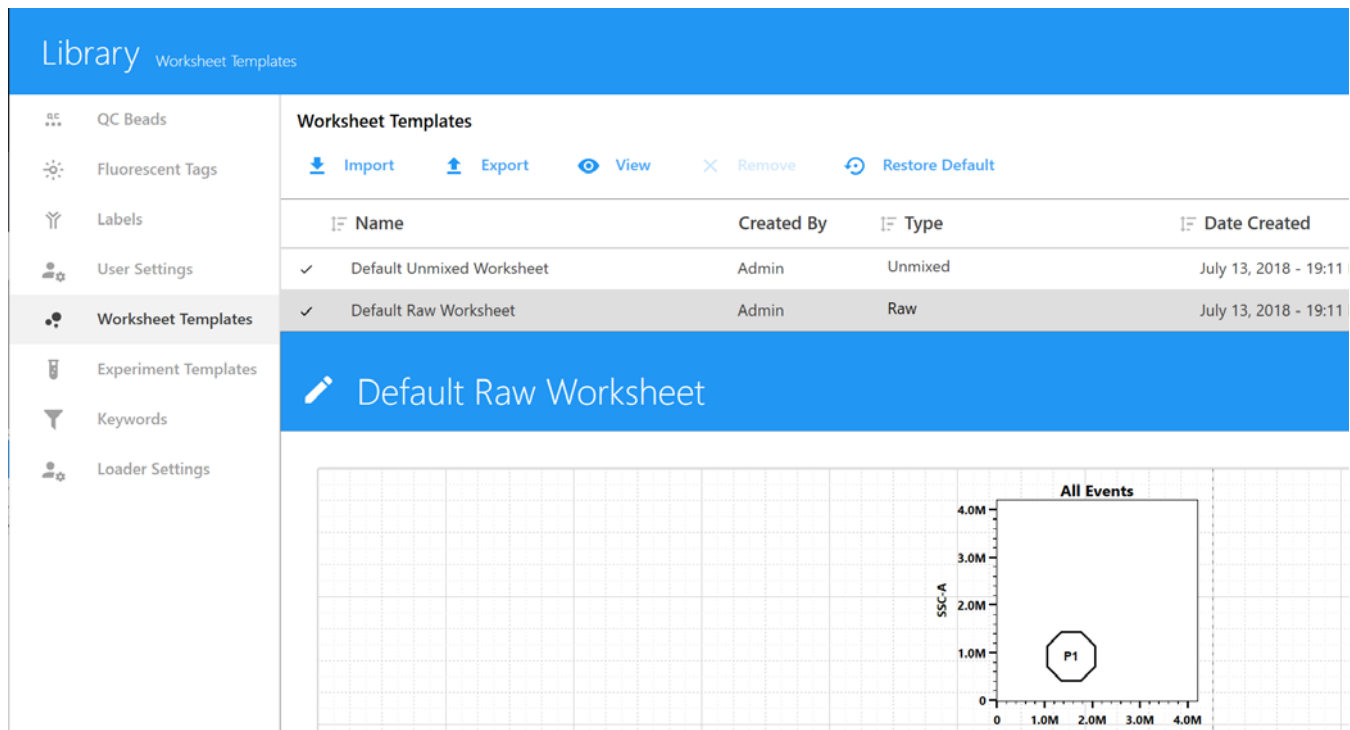
If you wish to reset the system-defined Default user settings to their original settings, select the Default user setting, then click Restore Default.

Library		User Settings			
QC Beads		Import Export Remove Restore Default			
Fluorescent Tags					
Labels					
User Settings		Name	Created By	Date Created	Description
		CytekAssaySetting	Admin	August 15, 2018 - 10:14 AM	Cytek Assay Setting
		Default	Admin	August 15, 2018 - 10:14 AM	Default
Worksheet Templates					

Worksheet Templates

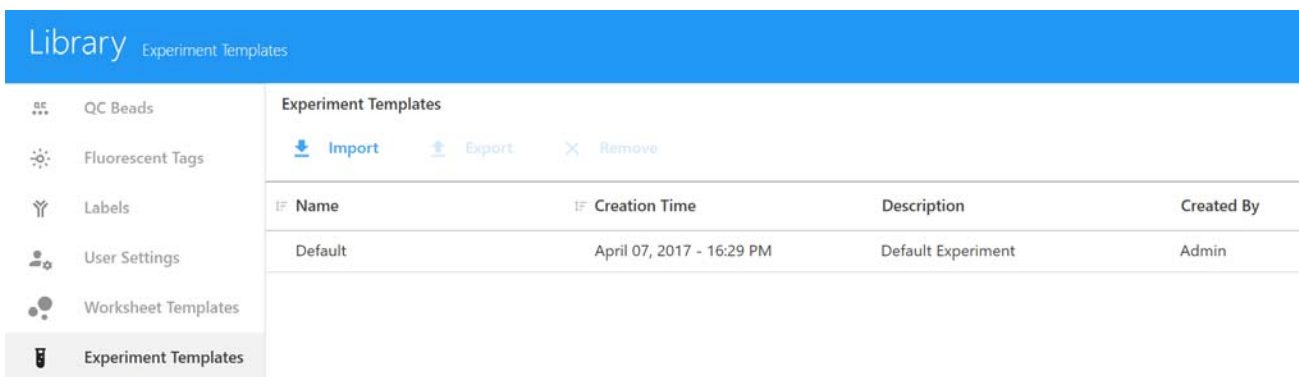
All worksheets created in the Acquisition module are saved in the library and can be accessed through the Worksheet Templates tab. Worksheets can be exported as WTML files and imported for re-use. For more information on worksheets, see [“About Worksheets” on page 20](#).

To view a worksheet, select it, then click View. To remove a worksheet that is no longer needed, select it, then click Remove. You can also set the default raw and unmixed worksheets back to their original defaults by selecting the worksheet, then clicking Restore Default.



Experiment Templates

Experiment templates contain the key elements of an experiment without the data. They can be saved and stored in the library for future use. The name, creation date and time, description, and creator information is displayed. Experiment templates can also be imported and exported from this tab. For more information on experiments, see [“About Experiments” on page 18](#).



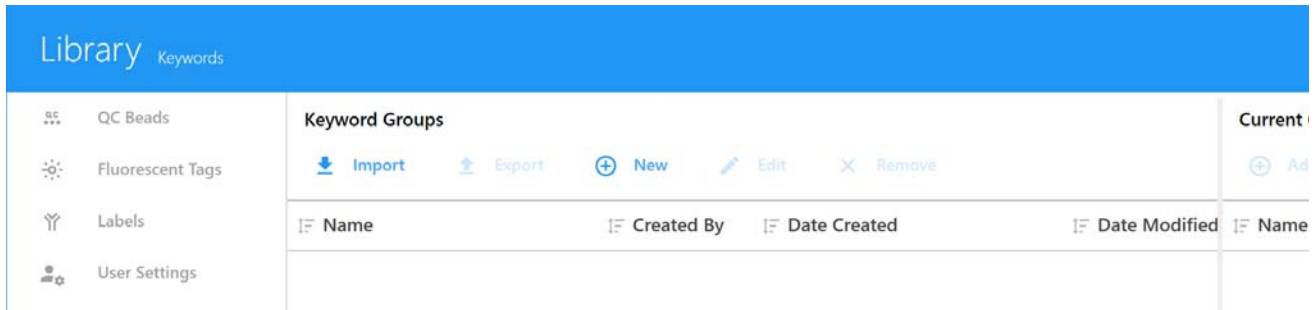
Keywords

Keywords allows you to type, import, or export user-defined, custom FCS keywords by group. These keywords can be assigned in an experiment at the experiment, group, or tube level and are exported with the FCS files.

The following types of keywords can be created:

- Numeric
- String

- Boolean
- Selectable numeric
- Selectable string



Standard Keywords

The following table describes the standard FCS 3.1 keywords found in SpectroFlo FCS files.

Keyword	Description
\$FIL	Tube or well name
\$PLATENAME	Plate name
\$WELLID	Well ID
\$VOL	Total sample volume (microliters)
\$DATE	Date that the FCS data set was acquired
\$BTIM	Beginning time of data acquisition
\$ETIM	End time of data acquisition
\$OP	Operator name
\$INST	Institution name
\$CYT	Name of cytometer (Aurora)
\$CYTSN	Cytometer serial number
\$SPILLOVER	Spillover matrix

Custom Keywords

The following table describes the custom keywords found in SpectroFlo FCS files. Custom keywords are not required as part of the FCS 3.1 standard.

Keyword	Description
CHARSET	Encoding. UTF8.
PLATEROWS	Number of rows of the plate
PLATECOLS	Number of columns of the plate
GROUPNAME	Plate name

Keyword	Description
CREATOR	Software name and version
FSC ASF	FSC area scaling factor
LASER{x}NAME	Name for each laser
LASER{x}DELAY	Laser delay for each laser
LASER{x}ASF	Area scaling factor for each laser
THRESHOLD	Threshold channel and value
WINDOW EXTENSION	Window extension used to record the tube/well
USERSETTINGNAME	The user setting name used to record the tube/well
APPLY COMPENSATION	True or False to indicate whether compensation is enabled when FCS file is loaded into SpectroFlo

Use can change the value of a custom keyword, then select Auto Update for that keyword to update the keyword in subsequent experiments.

Keyword Groups			Current Group: My Keywords				
Import	Export	New	Edit	Remove	Add	Edit	Remove
↑ Name	Created By	Date Created	Name	Suffix	Type	Value	<input type="checkbox"/> Auto Update
My Keywords	Admin	January 21, 2019 - 11:05 AM	Bead Lot		Numeric	0	<input type="checkbox"/>
6C TBNK	Admin	July 12, 2019 - 10:20 AM	PatientID		Numeric	0	<input type="checkbox"/>

Loader Settings

Loader settings can be saved and are stored in the library. The settings name, date the settings were created, as well as the name of user who created them are displayed. Click Description to add information about the settings.

Loader settings include mix time and speed, stage temperature setting, number of SIT flushes, and record data delay time. For more information on Loader settings, see ["Loader Settings" on page 97](#).

Library Loader Settings									
<ul style="list-style-type: none"> QC Beads Fluorescent Tags Labels User Settings Worksheet Templates Experiment Templates 	<p>Loader Settings</p> <p>Import Export Remove Restore Default</p> <table border="1"> <thead> <tr> <th>Name</th> <th>Created By</th> <th>Date Created</th> <th>Description</th> </tr> </thead> <tbody> <tr> <td> </td> <td> </td> <td> </td> <td> </td> </tr> </tbody> </table>	Name	Created By	Date Created	Description				
Name	Created By	Date Created	Description						

Preferences

The Preferences module allows you to change various functionality and display elements of the software user interface. The following section describes the options that can be changed in the Preferences module. Each section within the Preferences can be restored to its default settings by selecting Restore Default Preferences.



Restore Default Preferences

Acquisition Preferences

In the Acquisition tab, you can change the number of events displayed on plots during acquisition, the number of SIT flushes, the preview time before recording begins, the value that the gain settings increment when you click the spin-box arrow during acquisition, and the SIT Lift Distance. You can also enter default prefixes for tube, group, plate, and experiment names. A label/lot specific unmixing feature allows you to choose specific reference controls for unmixing. The Sample Delivery Mode options allow you to choose tube or plate for the acquisition method.

Acquisition

Number of Events to Display on Plots:	2,000
Number of Automatic SIT Flush After Tube Unplugged:	1
Recorded Tube Preview Time:	0
Gain Spinbox Up/Down Increment (Ctrl key Hold):	5
SIT Lift Distance:	0.75

Experiment

Default Tube Name Start With:	Tube_
Default Group Name Start With:	Group_
Default Plate Name Start With:	Plate_
Default Experiment Name Start With:	Experiment_

Unmixing Feature

Label/Lot Specific Unmixing

Sample Delivery Mode

Tube

Plate

Fluidics

Skip Fluidics Boost when Starting Acquisition

Enable Bubble/Clog Detection

In Case of Bubble

Move to next target after SIT flush

Stop immediately after SIT flush

In Case of Clog

Move to next target after SIT flush

Stop immediately after SIT flush

The following table describes the options in the Acquisition preferences.

Item	Description
Number of Events to Display on Plots	The number of events displayed in the pseudocolor plots, dot plots, and histograms. The default is 2,000 events.
Number of Automatic SIT Flush After Tube Unplugged	Choose the number (0–2) of SIT Flushes that will be performed once a tube is removed from the SIP.
Recorded Tube Preview Time	The number of seconds that elapse before the tube pointer moves to the next tube after the current tube is finished recording.
Gain Spinbox Up/Down Increment (Ctrl key Hold)	Increments the gain for each detector channel by the amount indicated when you hold the Ctrl key and select the up and down arrows of the Gain spinbox.
SIT Lift Distance	The distance in millimeters from the tip of the SIT to bottom of the well. The recommended starting point is 1.5 for U-bottom plates and slightly higher for V-bottom plates.
Experiment (Default Names)	Enter the default name for tubes, groups, plates, and experiments. Edit the default name and/or append to the names.
Label/Lot Specific Unmixing	Allows you to use lot-specific reference controls for unmixing when appropriate. If this option is selected, the software will search the library and experiment reference groups for reference controls that have the same fluorochrome, label, and lot information in order to use the corresponding control for unmixing.
Sample Delivery Method	Choose the delivery method—Tube for manual sample delivery, or Plate if you are using a Loader.
Fluidics	<p>Select Skip Fluidics Boost if you wish to skip the boost that quickly delivers sample from a tube/well to the flow cell at the start of acquisition.</p> <p>NOTE: The Skip Fluidics Boost feature works only for Medium and High flow rates.</p> <p>Enable Bubble/Clog Detection allows you to choose how you want to proceed in the event of a bubble or a clog when running a plate. You can either move to the next well or stop immediately after a SIT flush.</p> <ul style="list-style-type: none"> • If two consecutive clogs are detected when acquiring from a plate, acquisition stops immediately, regardless of how the Bubble/Clog Detection option is set. • Disabling SIT Flush (see <i>Number of Automatic SIT Flush After Tube Unplugged</i> above) also disables the SIT flush step during clog detection. For proper functionality, ensure the automatic SIT Flush is enabled when Bubble/Clog Detection is enabled.

Worksheet Preferences

The Worksheet tab allows you to change the way elements are displayed in the worksheet. Header and footer properties are also adjusted in the Worksheet tab.

The screenshot shows the 'Worksheet' preferences dialog. The left sidebar has 'Worksheet' selected. The main content area includes:

- Population Hierarchy Window Size:** Height: 300, Width: 300
- Statistics Table Window Size:** Height: 300, Width: 400
- Grid:**
 - Display Grid
 - Display Page Line
 - Grid Size: 1/8" (dropdown)
 - Snap to Grid
- Page Setup:**
 - Show & Print Page Number
 - Print Header & Footer
- Print Grid:**
 - Yes
 - No
 - Use Page Setting
- Page Orientation:**
 - Landscape
 - Portrait
- Page Margin:**
 - Narrow
 - Normal
 - Wide
- Title:** (dropdown)
- Page Size:** A4 8" x 12" (dropdown)
- Headers:**
 - Left: Not Selected (dropdown)
 - Right: Not Selected (dropdown)
- Footers:**
 - Left: Not Selected (dropdown)
 - Right: Not Selected (dropdown)

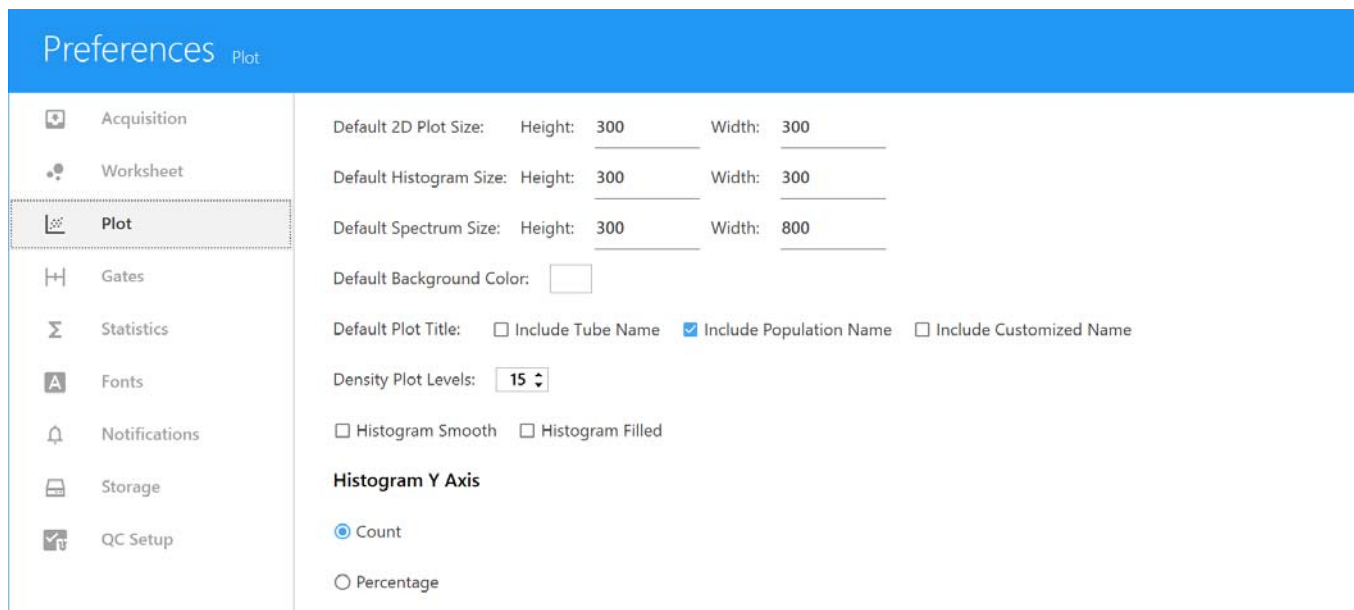
The following table describes the options in the Worksheet preferences.

Item	Description
Population Hierarchy Window Size	Sets the default height and width for the population hierarchy experiment element.
Statistics Table Window Size	Sets the default height and width for the statistics table.

Item	Description
Grid	<p>Display Grid – Toggles on/off the display of grid lines in the worksheet.</p> <p>Display Page Line – Toggles on/off the page break line in the worksheet.</p> <p>Grid Size – Modifies the size of the grid squares. Options include 1", 1/2", 1/4", and 1/8".</p> <p>Snap to Grid – Toggles on/off the ability for the worksheet elements to snap to and line up with the grid lines on the worksheet.</p>
Page Setup	<p>Show & Print Page Number – Toggles whether the page number is shown and printed.</p> <p>Print Header & Footer – Toggles whether the header and footer are printed.</p>
Print Grid	Toggles whether the grid is printed. Can also be set to use Page Setting.
Page Orientation	Toggles between landscape and portrait.
Page Margin	<p>Sets the margins of the page to Narrow, Normal, or Wide.</p> <p>Title allows you to select the text displayed as the worksheet's title. The title is shown when the worksheet is printed or exported as a PDF.</p>
Page Size	Sets the page width according to standard paper sizes.
Headers	Sets the text that is displayed in the left and right headers.
Footers	Sets the text that is displayed in the left and right footers.

Plot Preferences

The display properties of pseudocolor, dot, and histograms plots can be adjusted in the Plot tab.



Preferences Plot

- Acquisition
- Worksheet
- Plot**
- Gates
- Statistics
- Fonts
- Notifications
- Storage
- QC Setup

Default 2D Plot Size: Height: 300 Width: 300

Default Histogram Size: Height: 300 Width: 300

Default Spectrum Size: Height: 300 Width: 800

Default Background Color:

Default Plot Title: Include Tube Name Include Population Name Include Customized Name

Density Plot Levels: 15

Histogram Smooth Histogram Filled

Histogram Y Axis

Count

Percentage

The following table describes the options in the Plot preferences.

Item	Description
Default 2D Plot Size	Set the default height and width of the pseudocolor plots and dot plots in pixels.
Default Histogram Size	Set the default height and width of histograms in pixels.
Default Spectrum Size	Set the default height and width of spectrum plots in pixels.
Default Background Color	Set the default background color for all plots. Click to select from the available colors.
Default Plot Title	Customize the title of all plots to include the tube name, population name, and/or a custom name.
Density Plot Levels	Increase or decrease the number of density levels displayed in the pseudocolor plot.
Histogram Smooth	Set whether histogram distributions are smoothed.
Histogram Filled	Set whether histogram distributions are filled.
Histogram Y Axis	Set the scale of the histogram y-axis to a count or a percentage.

Gates Preferences

Gate properties can be adjusted in the Gates tab.

The screenshot shows the 'Preferences' dialog box with the 'Gates' tab selected. The left sidebar contains navigation options: Acquisition, Worksheet, Plot, Gates (selected), Statistics, Fonts, Notifications, Storage, and QC Setup. The main area displays the following settings:

- Default Name Location: **Inside** (dropdown)
- Show % of Parent together with Gate Name
- Show Count together with Gate Name
- Gate Boundary Line Weight: **Normal** (dropdown)
- Default Colors for First: **10** (dropdown) Gates:

Under 'Interval Gate Default Color':

- No Color
- Has Color

Under 'Quadrant Gate Default Color':

- No Color
- Same Color
- Different Color in each Quad

The 'Default Colors for First' dropdown is set to '10', which opens a table showing 10 gates with their respective colors:

Gate	Color
1	Red
2	Blue
3	Green
4	Purple
5	Orange
6	Cyan
7	Olive
8	Magenta
9	Brown
10	Grey

The following table describes the options in the Gates preferences.

Item	Description
Default Name Location	Select where the gate name is displayed with respect to the gate itself.
Show % of Parent together with Gate Name	Toggles on/off the display of the % of Parent with the gate name.
Show Count together with Gate Name	Toggles on/off the display of the population count with the gate name.
Gate Boundary Line Weight	Sets the thickness of the line drawn by the gate.
Interval Gate Default Color	Toggles on/off whether the population captured by the interval gate has a default color.
Quadrant Gate Default Color	Select whether the population captured by the quadrant gate has a default color.
Default Colors for First X Gates	Set the number of gates that will follow the color scheme detailed in the gate color table. The order in which the colors appear can be changed.

Statistics Preferences

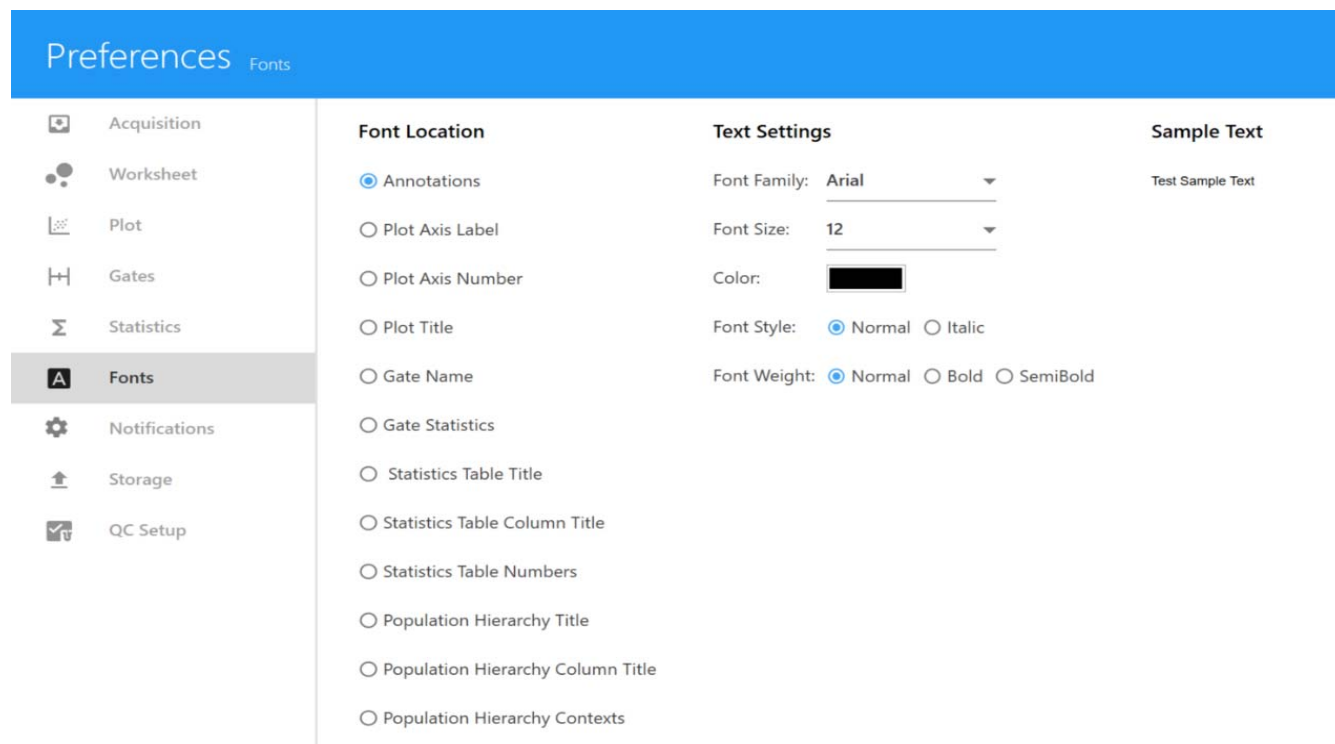
The default degree of precision (number of decimal places) of the statistics displayed in the worksheet can be modified in the Statistics tab.

The precision for the following statistics can be adjusted: Mean, rSD, % rCV, Mean, Max, Min, SD, % CV, % Total, % Parent, and % Grand Parent.

Decimal Places	
Statistics Variable	Decimal Places
Median	0
rSD	0
% rCV	2
Mean	0
Max	0
Min	0
SD	0
% CV	2
% Total	2
% Parent	2
% Grand Parent	2

Fonts Preferences

The Fonts tab allows you to change the font properties of each display element.

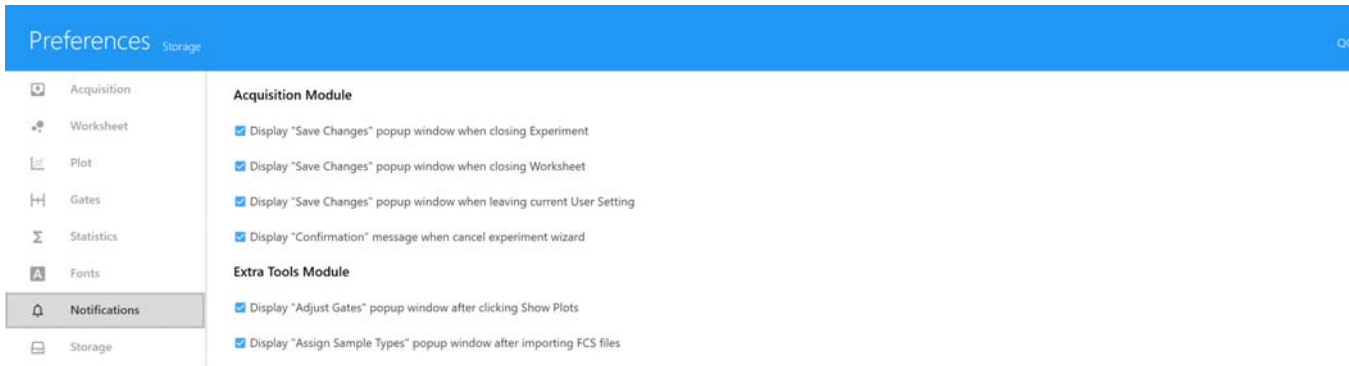


The following table describes the options in the Fonts preferences.

Item	Description
Font Locations	Select which display element's font to modify.
Text Settings	Font Family – Select the font family. Font Size – Select the font size. Color – Select the font color. Font Style – Toggles between normal and italic. Font Weight – Select normal, bold, or semibold.
Sample Text	Example preview text with the properties set in the Text Settings.

Notifications Preferences

The Notifications tab allows you to change certain notification settings in the Acquisition and Extra Tools modules.



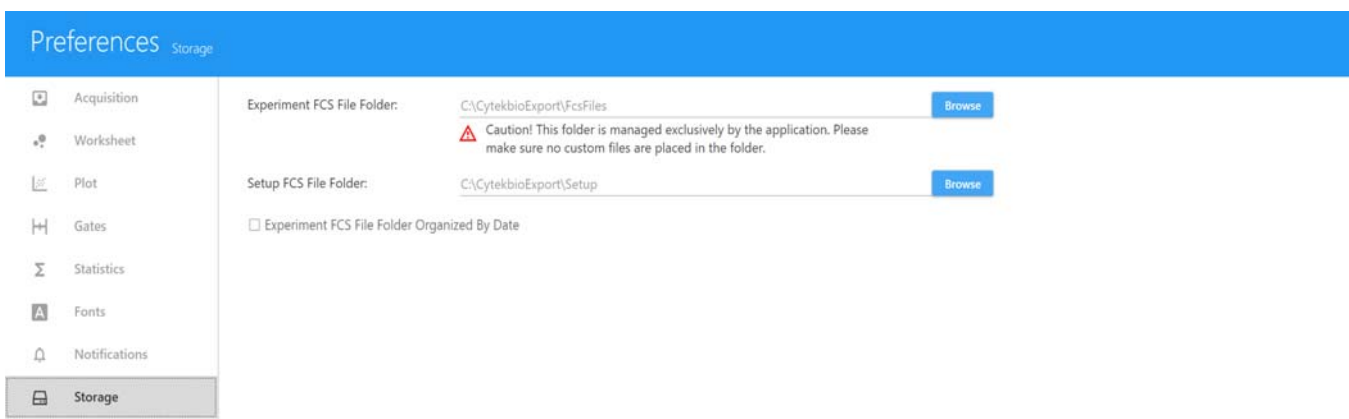
The following table describes the options in the Notifications preferences.

Item	Description
Acquisition Module	Toggles whether to display the Save Changes pop-up window when closing an Experiment, Worksheet, or User Settings, as well as whether to display a confirmation message when canceling the experiment wizard.
Extra Tools Module	Toggles whether to display the instructional dialog boxes in the Extra Tools module.

Storage Preferences

The Storage tab allows administrators to set the default storage locations for the experiment FCS files and setup FCS files. This option is available only for administrators.

■ **NOTE:** Do not move files stored in these folders.

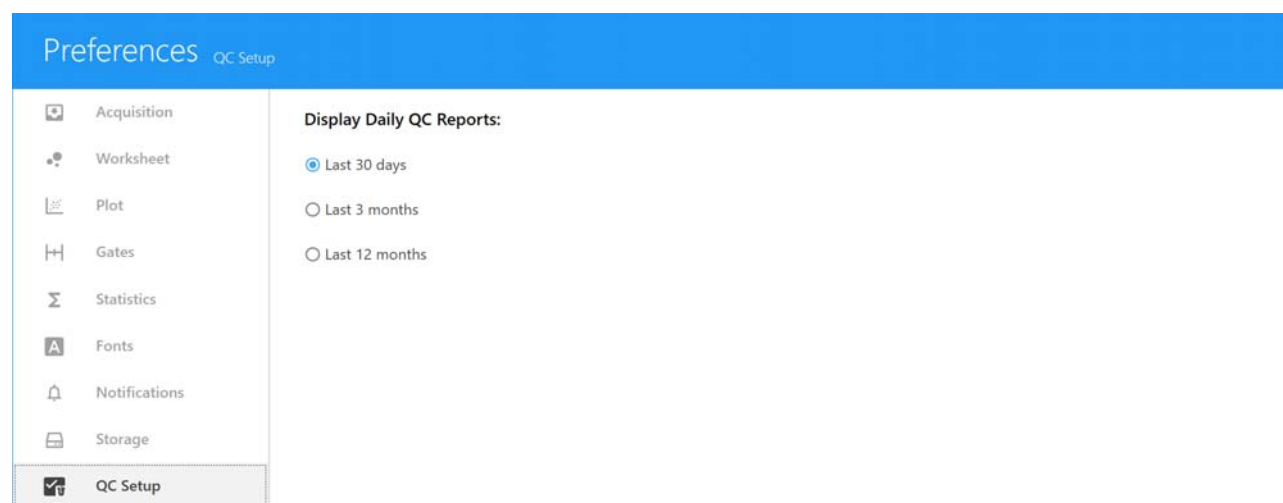


The following table describes the options in the Storage preferences.

Item	Description
Experiment FCS Files Folder	Select the folder where Experiment FCS files are saved. FCS files from experiments where only raw data was acquired, as well as FCS files from experiments where live unmixing was performed are stored here. For experiments where live unmixing is performed, both raw and unmixed FCS files are saved.
Setup FCS Files Folder	Select the folder where FCS files generated by QC & Setup procedures are saved.

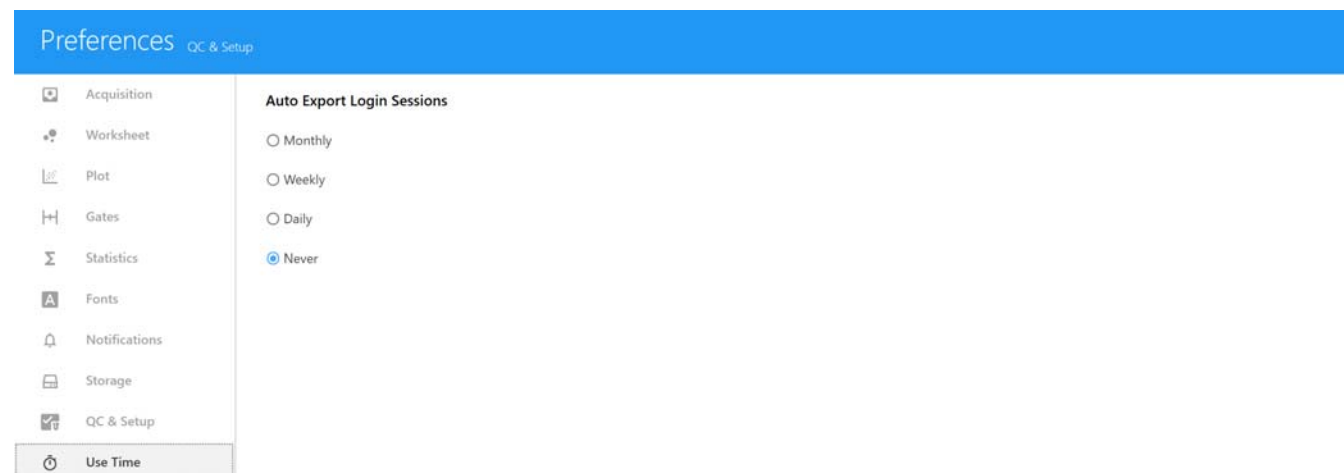
QC Setup Preferences

The QC Setup tab allows you to select the days/months of QC reports to display in the Reports section of the Cytometer QC menu in the QC & Setup module.



Use Time Preferences

The Use Time tab allows you to select if and when (daily, weekly, monthly) to automatically export the login sessions. The auto-exported use time files are exported to C:\CytekbioExport\LoginSessions. For more information on exporting login sessions, see [“Use Time” on page 92](#).



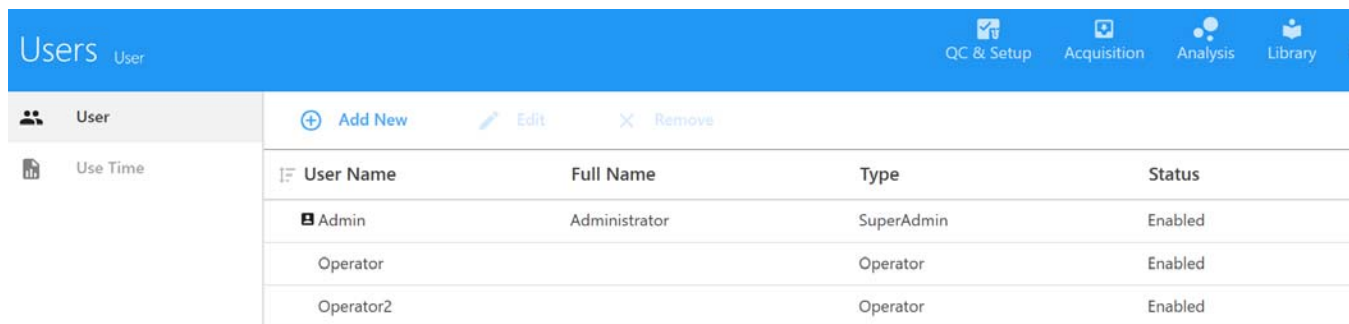
Users

User accounts can be managed in the Users module. User account information and use time are stored in the module.

There are two types of user accounts—administrator and operator. Only administrators can manage user accounts.

Managing Users


Administrators can add, remove, edit, and disable user accounts from the User tab. User passwords can also be changed/reset. The User tab lists all users and displays the type and status of each.



The screenshot shows the 'Users' module interface. At the top, there are navigation tabs for 'QC & Setup', 'Acquisition', 'Analysis', and 'Library'. Below the navigation, there are three buttons: 'Add New', 'Edit', and 'Remove'. The main area displays a table with the following columns: 'User Name', 'Full Name', 'Type', and 'Status'. The table contains three rows of user data.

User Name	Full Name	Type	Status
Admin	Administrator	SuperAdmin	Enabled
Operator		Operator	Enabled
Operator2		Operator	Enabled

Adding a New User Account

- 1 Click  Add New in the User tab of the Users module. This option is available only for administrators.

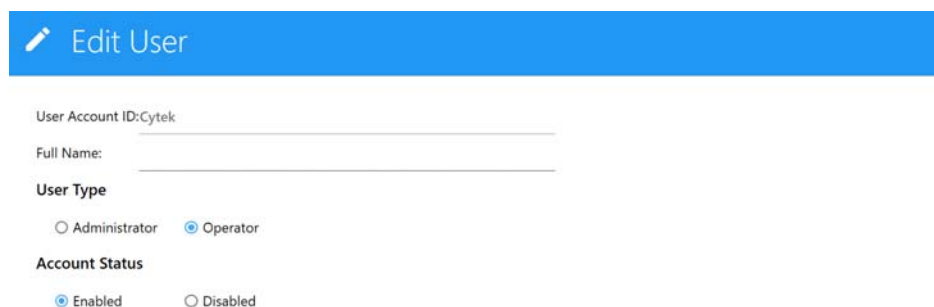


The screenshot shows the 'Create User' form. It has a blue header with a pencil icon and the text 'Create User'. Below the header, there are several input fields and radio buttons. The 'User Account ID' field contains the text 'Cyttek'. The 'Password' and 'Confirm Password' fields are masked with dots. The 'Full Name' field is empty. Under 'User Type', the 'Operator' radio button is selected. Under 'Account Status', the 'Enabled' radio button is selected.

- 2 Enter a user account ID and password, then enter the password again to confirm. The user account ID appears in the user name field in the User's tab.
- 3 (Optional) Enter the user's full name.
- 4 Select the user type—administrator or operator.
- 5 Select the user status—enabled or disabled.
- 6 Click Save.

Editing a User Account

- 1 Select the user from the User tab of the Users module, then click Edit.



User Account ID:Cytek

Full Name:

User Type

Administrator Operator

Account Status

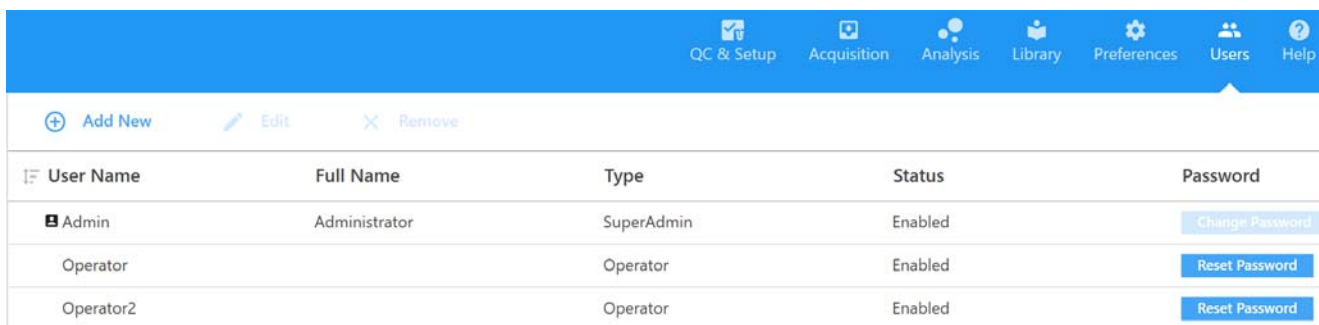
Enabled Disabled

- 2 You can edit or add a user name. You can also change the user type and/or account status.
- 3 Click Save.

Resetting a User Password

An administrator can change their or another administrator's password or reset an operator's password.

- 1 Select the user from the User tab of the Users module.
- 2 Click Reset Password.



User Name	Full Name	Type	Status	Password
Admin	Administrator	SuperAdmin	Enabled	Change Password
Operator		Operator	Enabled	Reset Password
Operator2		Operator	Enabled	Reset Password

- 3 A dialog opens displaying the new password.
- 4 Make note of the password and click OK.

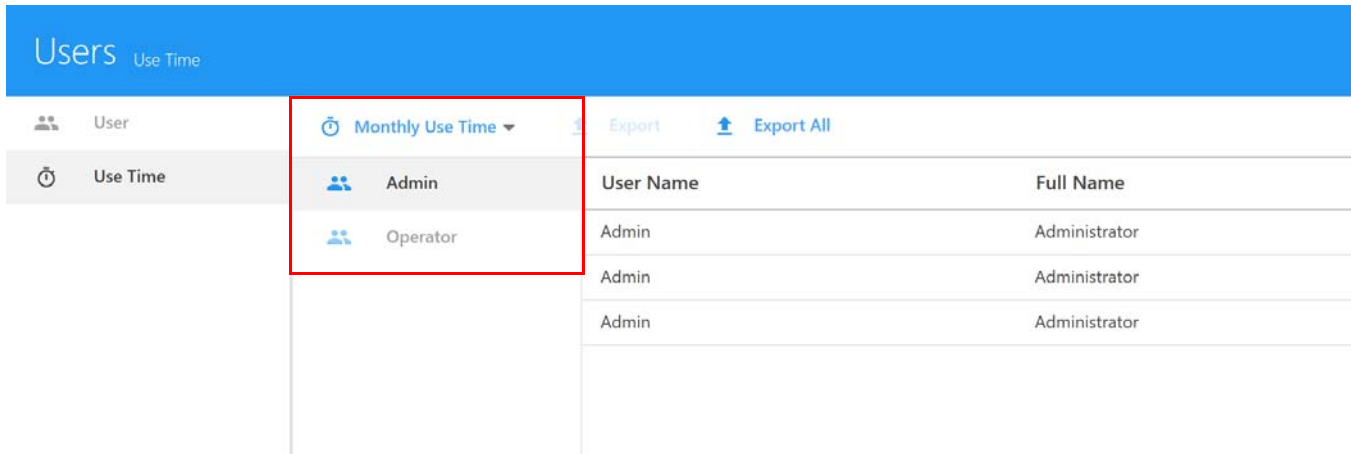
Removing a User Account

To remove a user account, select the user from the User tab of the Users module, then click Remove. When you remove a user account all the account information and that data acquired by that user is deleted. The FCS files saved by the user are retained.

Use Time

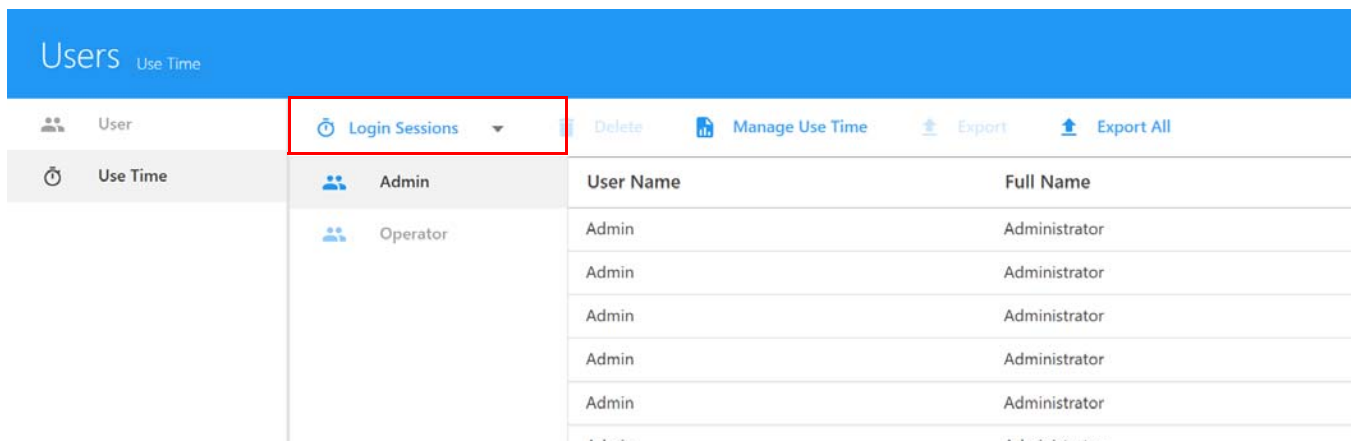
Select the Use Time tab of the Users module.

Use Time shows the total daily, weekly, or monthly use time (duration) and number of sessions that each user is on the system. Choose Admin or an operator to view and export the use times for the respective user account.



The screenshot shows the 'Users Use Time' interface. The left sidebar has 'User' and 'Use Time' tabs. The 'Use Time' tab is active. A dropdown menu is open, showing 'Monthly Use Time' selected, with 'Admin' and 'Operator' options below it. The main area displays a table with columns 'User Name' and 'Full Name'. The table contains three rows, all with 'Admin' in the 'User Name' column and 'Administrator' in the 'Full Name' column. Buttons for 'Export' and 'Export All' are visible at the top right of the table area.

To see the individual login sessions for each day, select Login Sessions. Choose Admin or Operator to view and export the login sessions for the respective user account. The log on and log off times for each session, as well as the session duration are displayed. Administrators can manage the list of login sessions by deleting all sessions before a specified date. To delete sessions, click Manage Use Time, select the date, and click Delete.



The screenshot shows the 'Users Use Time' interface. The left sidebar has 'User' and 'Use Time' tabs. The 'Use Time' tab is active. A dropdown menu is open, showing 'Login Sessions' selected, with 'Admin' and 'Operator' options below it. The main area displays a table with columns 'User Name' and 'Full Name'. The table contains six rows, all with 'Admin' in the 'User Name' column and 'Administrator' in the 'Full Name' column. Buttons for 'Delete', 'Manage Use Time', 'Export', and 'Export All' are visible at the top right of the table area.

Administrators can export daily, weekly, and monthly use times and login session to .csv files by selecting the user(s) and session dates and clicking Export or selecting Admin or user(s) and clicking Export All to export all use times or login sessions.

Loader

Loader Overview

The new Automatic Micro-Sampling System (AMS), referred to as Loader throughout this guide, is an optional sample loading accessory that delivers a standard 96-well plate to the cytometer for sample acquisition. The Loader plate stage houses a metal element that can cool/heat from 4°–30°C, if you choose. A wash station, located at the back of the plate stage is used for cleaning the mixing probe.

Loader settings can be adjusted for the duration and speed of probe mixes, the number of SIT flushes, and the stage temperature. Pre-defined Loader settings include the default settings for high throughput mode, standard mode, and low carryover mode.



When the Loader is powered on and successfully connected to the software, the status indicator in the lower-right corner of the screen shows a green checkmark. A red X appears if the Loader is not powered on or not connected to the software.

✓ Sheath ✓ Waste ✓ Cytometer ✓ Loader

Using the Loader

The Loader must be calibrated for the plate type you will be using. For information on calibrating the Loader for the plate, see “Calibrating a Plate” on page 101.

When using the Loader, place fluidics tanks on the floor beneath the instrument, otherwise fluid can backflush into the wash well. For systems with a Loader, we recommend using the 20-L cubitainers instead of the 4-L tanks.

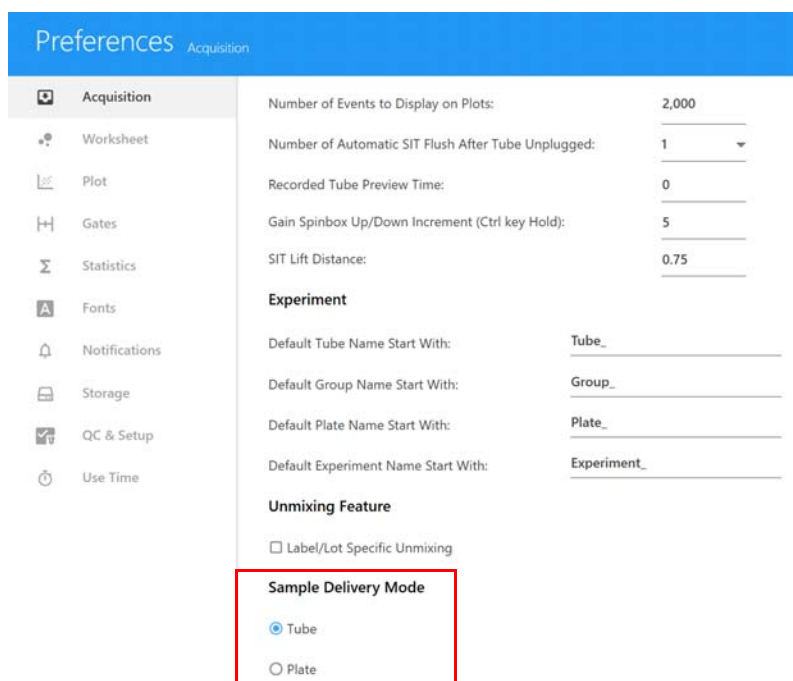
Enabling the Loader

- 1 Turn on the power to the Loader. The power switch is at the back of the Loader where the fluidic lines connect.
- 2 If there is a tube on the SIP, remove it.
- 3 Pull the lever towards you to bring the mixer forward.



power switch

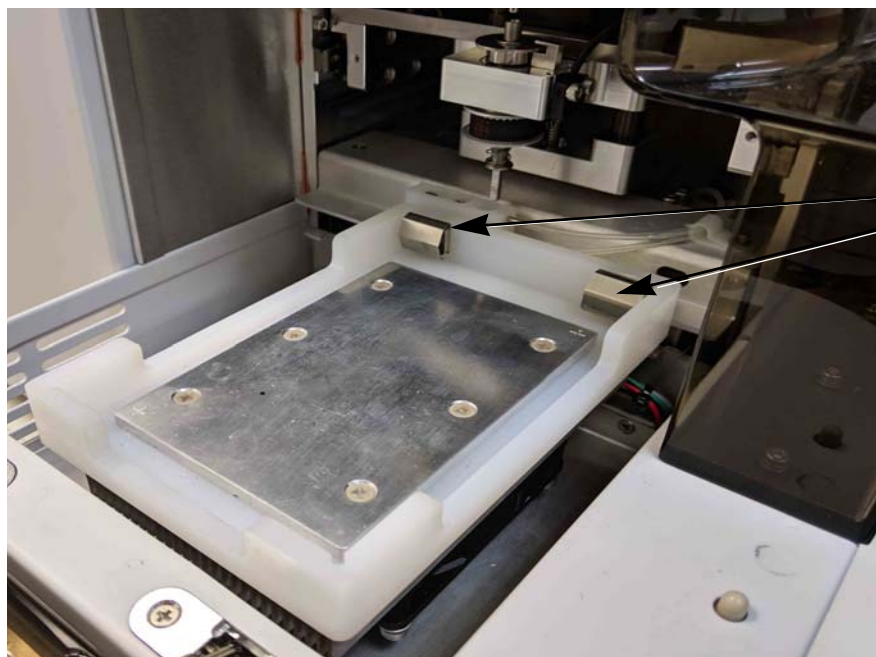
- 4 Set the Sample Delivery Mode in Acquisition Preferences to Plate.



- 5 Before running your experiment, you must perform a SIT calibration to set the proper SIT depth within the well. See [“Calibrating the SIT”](#) on page 103.

Loading a 96-Well Plate

- 1 If necessary, press Eject from either the QC & Setup or Acquisition module to eject the stage, bringing it forward.
- 2 Load a plate on the plate stage so that position A1 is located in the front-left corner. Insert the edge of the plate behind the metal tabs at the front of the plate stage, then press down on the back edge of the plate to secure it in the holder.



Insert back edge of plate in front of metal tabs, then press down on front edge of plate to secure.

- 3 Press Load from the QC & Setup or Acquisition module to load the plate, followed by Start to begin acquisition.
Or press Start to load the plate and begin acquisition.
- 4 To switch back to tube mode, set the Sample Delivery Mode in the Acquisition Preferences back to Tube and slide the Loader lever back.

Minimum Well Volume

When acquiring from a well, a boost is applied to deliver the sample to the flow cell as quickly as possible. This process, as well as mixing, results in the loss (consumption) of up to 75 μL of sample, as the sample is neither acquired nor recorded. We recommend a minimum volume of 150 μL of sample per well, keeping in mind that approximately 75 μL will be lost.

■ **NOTE:** With the Loader v1.5, sample consumption is reduced to <35 μL . This can be reduced further by selecting to skip the fluidics boost. For information, see [“Acquisition Preferences”](#) on page 81.

You can disable the boost to save sample. For information see [“Acquisition Preferences”](#) on page 81.

Loader Settings in the Experiment

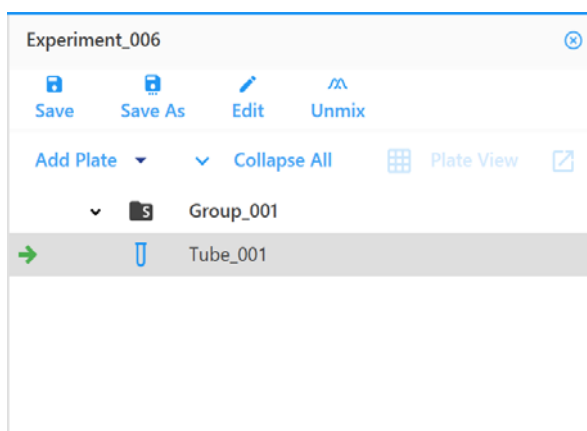
This section lists the differences in the Acquisition module when using a Loader. The following panes are described:

- Group Hierarchy (“Group Hierarchy/Plate Display” on page 96)
- Acquisition Controls (“Loader Acquisition Controls” on page 96)
- Loader Settings (“Loader Settings” on page 97)

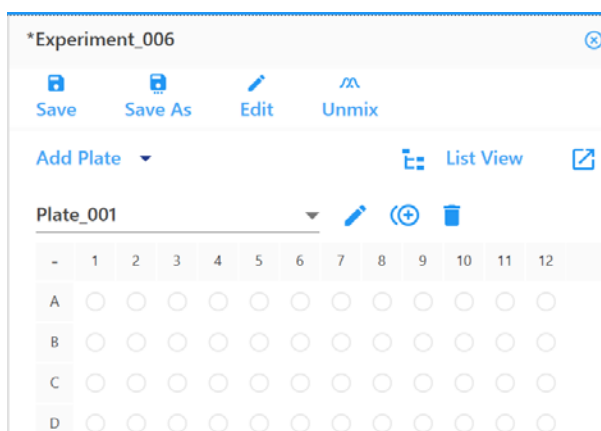
Group Hierarchy/Plate Display

The Group and Tube hierarchy pane when using a Loader defaults to one group with one tube. It also provides an option to add a plate. You can view either a hierarchical list (List View), or once a plate is added, a graphical representation of the plate (Plate View).

Default Experiment List View

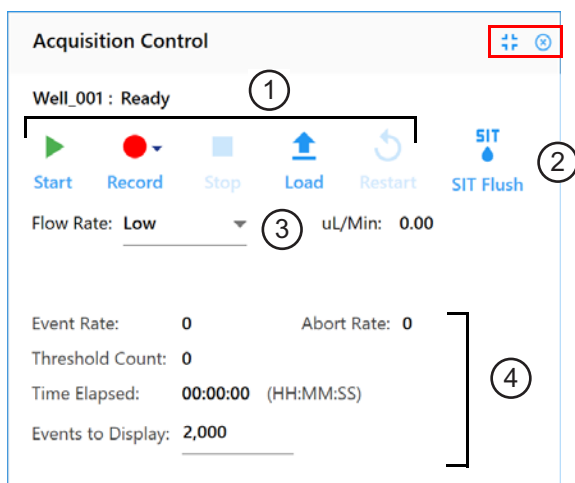


Default Experiment Plate View Once Plate is Added



Loader Acquisition Controls

The Acquisition Control pane allows you to start, stop, and pause acquisition, record data, and restart acquisition counters. To show, hide, or undock (float) this pane from the experiment panel, use the dock/undock and hide icons in the top-right corner.

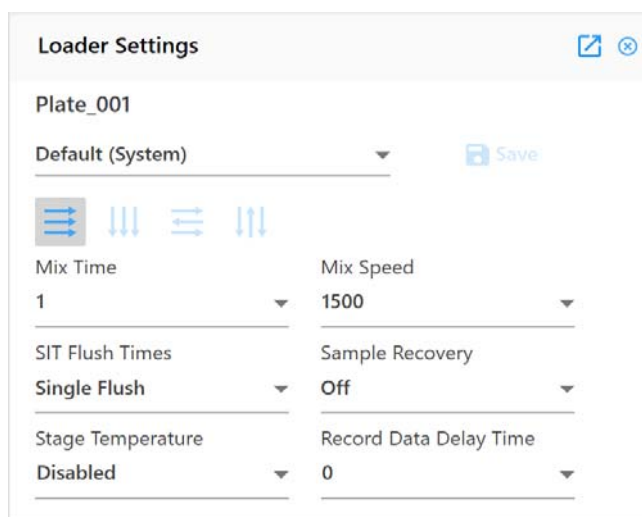


The following table describes the controls in the Acquisition Control pane.

No.	Control	Description
1	Start/Record/Stop/Load/Eject/Restart	Start and Record are enabled when the Loader is on, the lever is pulled forward, and the Sample Delivery Mode in Acquisition preferences is set to Plate. Select Start to load the plate and start acquisition Select Record to record data. Record can also start acquisition. Select Stop to stop acquisition. Select Load or Eject to load or eject the plate. Select Restart to restart the acquisition counters. All events and results displayed are refreshed. Stop and Restart are enabled once Start is selected. Stop and Pause are enabled once Record is selected.
2	SIT Flush	Select to perform a SIT Flush
3	Flow Rate	Select Low (15 μ L/min), Medium (30 μ L/min), or High (100 μ L/min). The exact flow rate is displayed.
4	Event Rate, Abort Rate, Threshold Count, Time Elapsed	Displays the real-time counts during acquisition.
5	Events to Display	Enter the number of events to display during acquisition.

Loader Settings

Several default Loader settings are available. You can select the Default, High Throughput, or Low Carryover Loader Setting, depending on your application. You can also create custom settings. Loader settings are enabled once a plate is added. Adjust the settings before acquisition.



The following table describes the Loader settings.

Setting	Description
Selected Settings	Three Loader settings are available—default, high throughput, and low carryover. You can create your own custom settings.
Acquisition order	Select the order that you want the plate to run. Wells are acquired by: <ul style="list-style-type: none"> • row from left to right (A1-A12, B1-B12, etc) • column from top to bottom (1A-1H, 2A-2H, etc) • row from left to right, then right to left (A1-A12, B12-B1, C1-C12, etc) • column from top to bottom, then bottom to top (1A-1H, 2H-2A, etc)
Mix Time	Select the time (in seconds) each well gets mixed before acquisition. You can also disable the mix time.
Mix Speed	Select the speed at which the mixer spins (in RPM).
SIT Flush Times	A SIT flush is performed over the wash station after each acquisition. Choose single flush, double flush, or disabled if you do not wish to perform a SIT flush.
Sample Recovery	Allows any remaining sample that is left in the SIT after acquisition is complete to be deposited back into the well.
Stage Temperature	Select the plate stage temperature (4°–30°C).
Record Data Delay Time	Select the time in seconds you wish to preview data from a well before recording begins once you click Record.


Experiments in Plate Mode

Follow the instructions “Creating a New Experiment” on page 50 to create a new experiment. The steps to create a new experiment when running in plate mode are the same as when running in Tube mode, except for the following tasks—creating groups and defining Loader settings—outlined below.

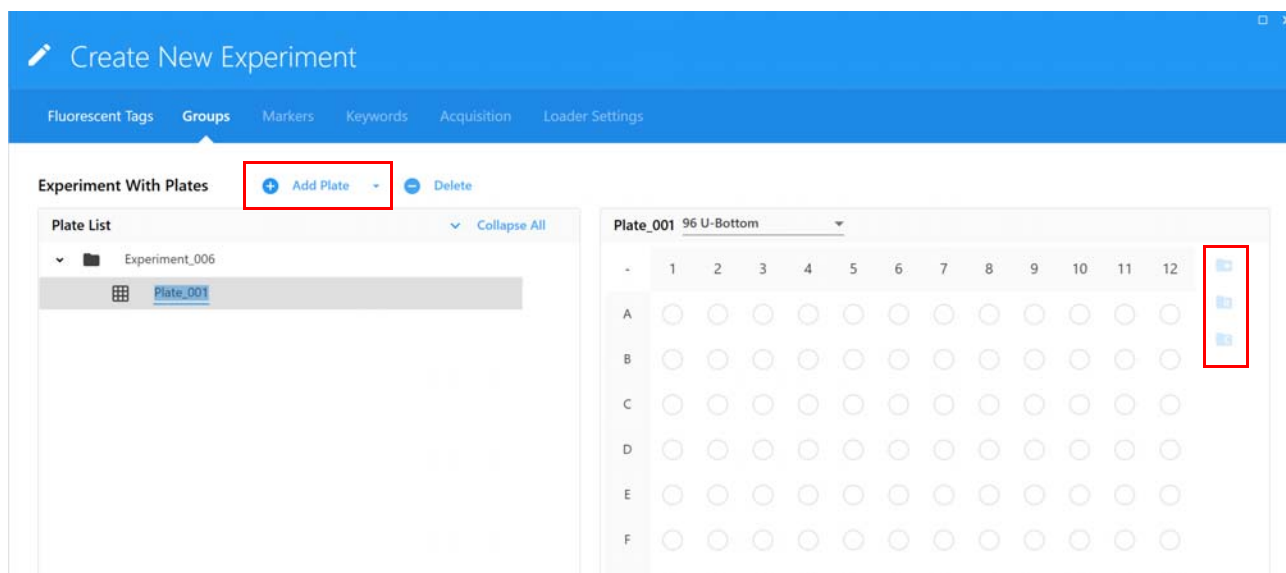
Creating Groups When Using the Loader

■ **NOTE:** When running sticky samples, such as lyse, no wash (LNW) samples, we recommend adding cleaning wells between samples to thoroughly clean the mixing probe. For example, add two wells, one with 10% bleach and the other with DI water. At the end of a plate, consider adding a group of four wells, two with 300 μ L of 10% bleach and two with 300 μ L of DI water. Program a long mix (15 seconds at 1500 rpm) to thoroughly clean the mixing probe.

Before you can add groups when creating a new experiment, you must add a plate.

- 1 To add groups when running in plate mode, click  Add Plate, then select the plate type (96 U-, V-, or Flat-Bottom)

A plate image appears on the right. Three icons at the far right allow you to add groups.



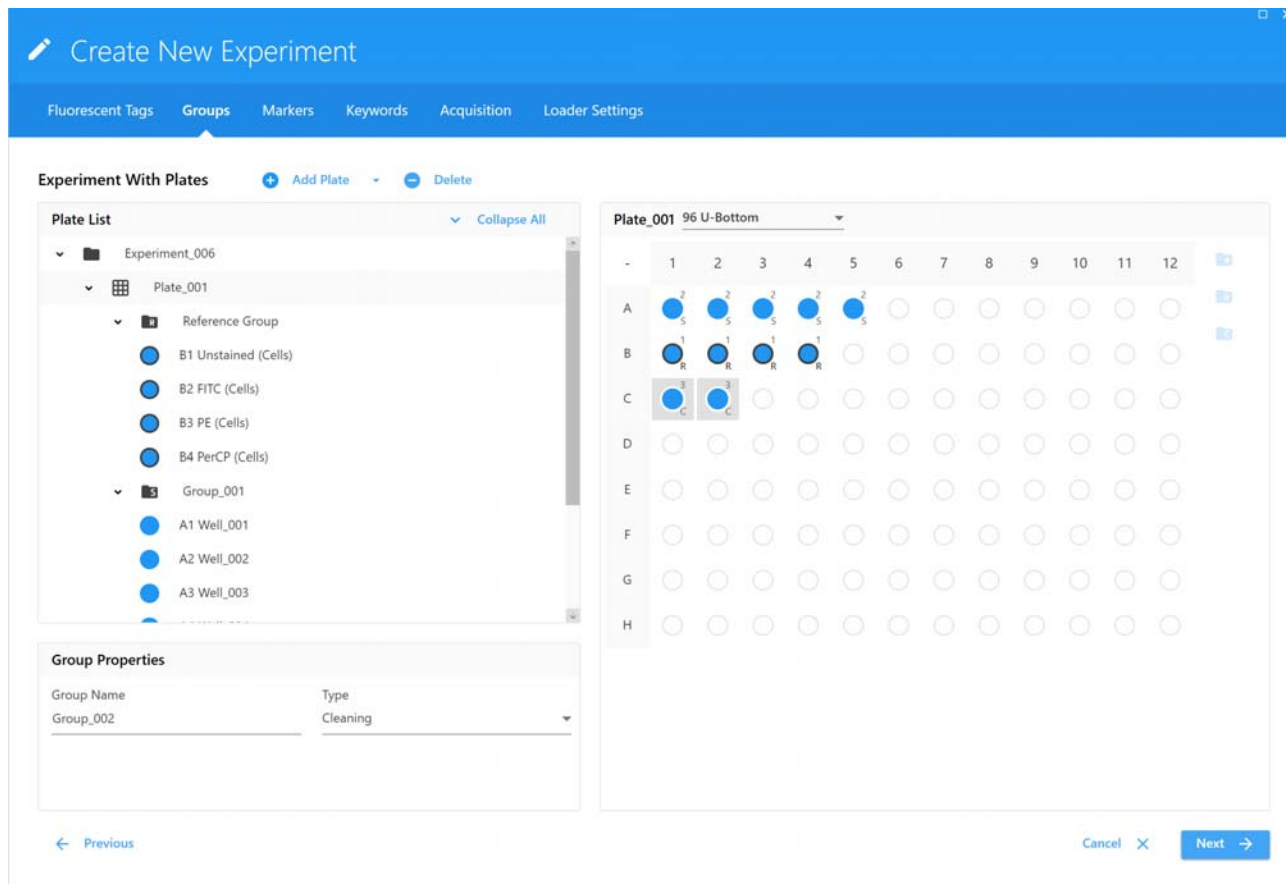
- 2 Click in the plate image to select a well, or click and drag to select multiple wells corresponding to the wells in the group you wish to add, then click the appropriate icon to the right of the plate to define the sample types in the group:

- To add a group for samples, click +.
- To add a group for reference controls, click R.

If you are intending to unmix with controls acquired in this experiment you must add a reference group. When you add a reference group you will be prompted to define the reference controls. See [step 7 on page 53](#) for details on defining the unstained and fluorescence controls.

- To add a group for cleaning well(s), click C. For example, you can add DI water to wells to rinse the SIT and clean the mixing probe to prevent carryover.

Wells will be marked as sample (S), reference control (R), or cleaning (C).

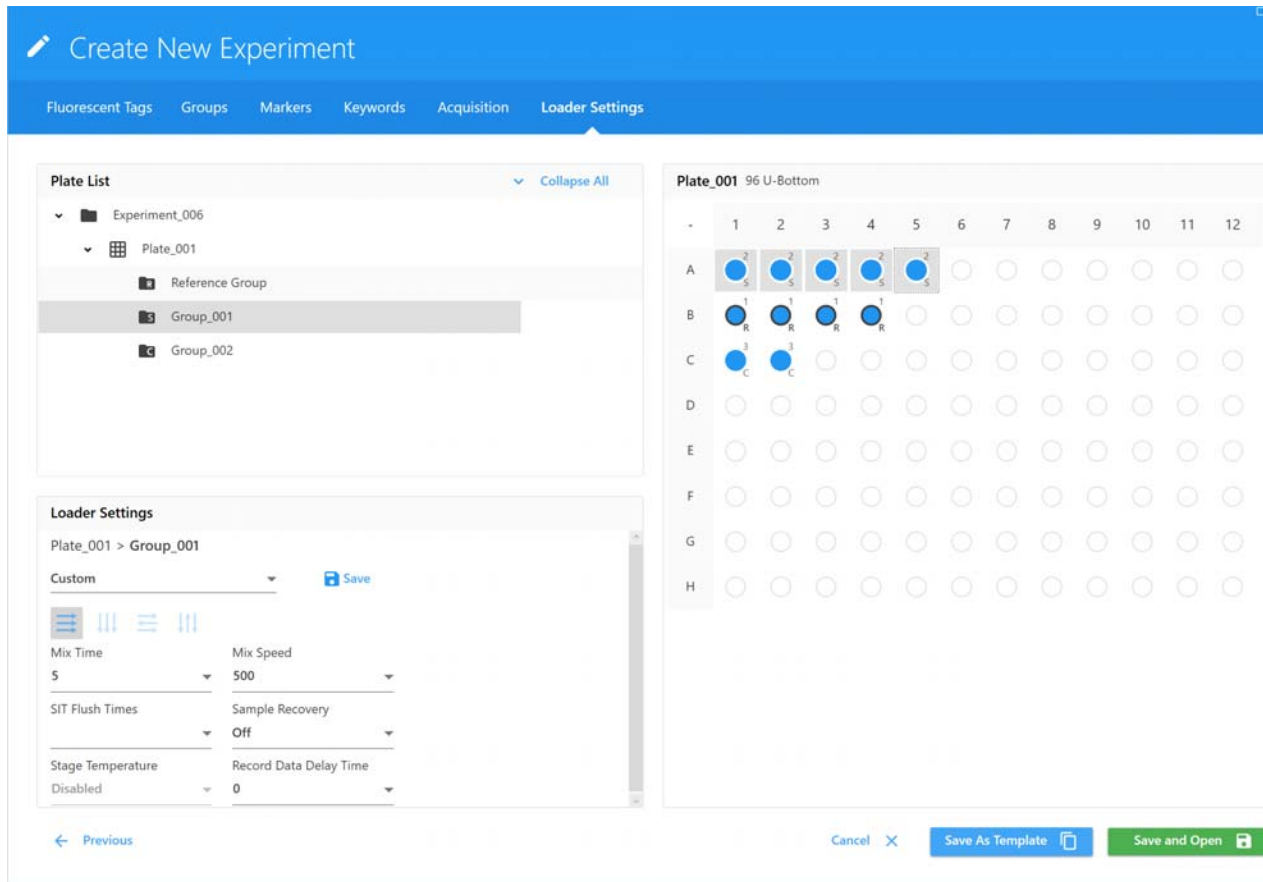


- 3 When all groups have been defined, click Next.
- 4 Follow steps 14 through 16 starting on [page 54](#) to continue creating the experiment.

Defining Loader Settings

- 1 The last step when creating an experiment is to select the Loader settings. Loader setting can be applied to all wells in the plate or at the group level. See [“Loader Settings” on page 97](#) for information.

We recommend using with the Default settings for most immunophenotyping applications.



- 2 Once the worksheet and stopping criteria have been defined, click Save and Open to open the new experiment.

To make any changes to the experiment, click Edit above the group/well hierarchy.

- 3 To acquire controls and samples and perform live unmixing, see [“Live Unmixing” on page 60](#).

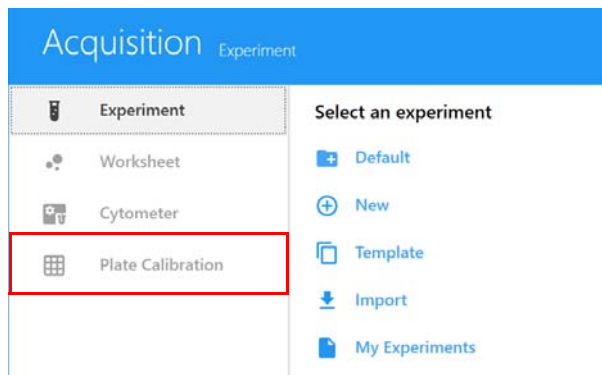
Calibrating a Plate

The plate calibration feature allows administrators to calibrate the stage for each of the compatible 96-well plates. Use the controls to align the SIT within well A1 and adjust the depth of the mixer in the well. The settings are saved automatically. Use the plate calibration feature if you change plate types.

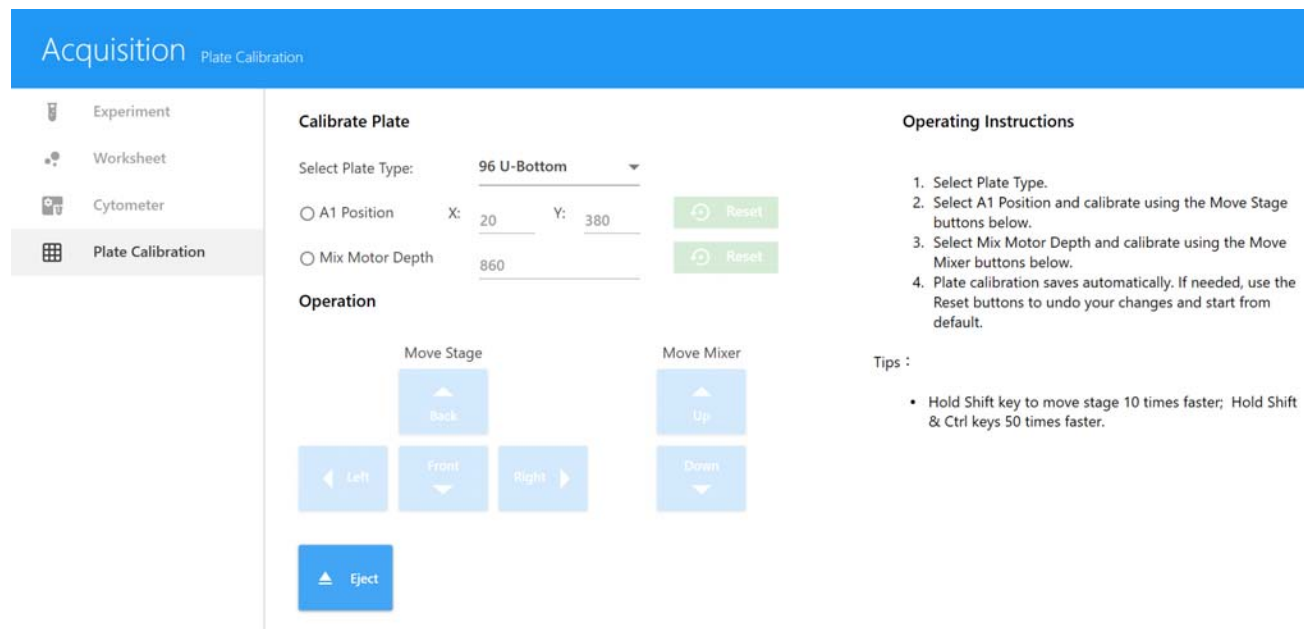
For a list of compatible plates, see [“Supported 96-Well Plates” on page 126](#).

- 1 Click the Plate Calibration icon in the Acquisition module.

The Plate Calibration option is available only to administrators.



The plate calibration screen appears.



- 2 Load the plate that you wish to calibrate.
- 3 Select the plate type from the drop-down list.
- 4 Select A1 Position to calibrate the X and Y directions.
 - Use the Move Stage buttons to align the SIT within the well.
 - Click Reset if you wish to revert to the default settings.
- 5 Select the Mix Motor Depth to calibrate the Z direction.
 - Ensure the SIT is positioned above well A1 and click Continue.
 - Use the Move Mixer Down button to move the mixing probe deeper into the well. Use the Move Mixer Up button to move the mixing probe to a shallower position.
 - Click Reset if you wish to revert to the default settings.

■ **NOTE:** A Reset All button reverts all settings to the defaults.

The settings are saved automatically and should not need to be changed unless you change the plate type.

Calibrating the SIT

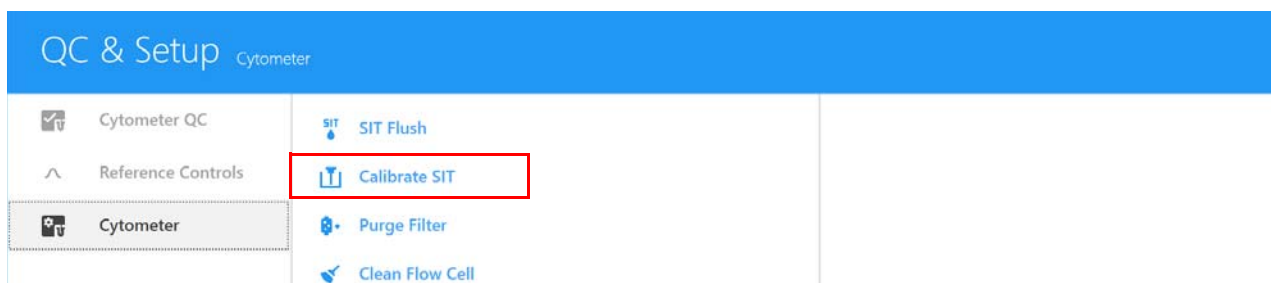
Use the Calibrate SIT feature to set the proper SIT depth within a plate well. You will need to perform a SIT calibration:

- prior to running your first experiment of the day in plate mode
- every time you switch from tube mode to plate mode
- if you change plate types

The calibration value applies only to your individual user account. Once calibration is complete, the settings apply to all plates of the same type.

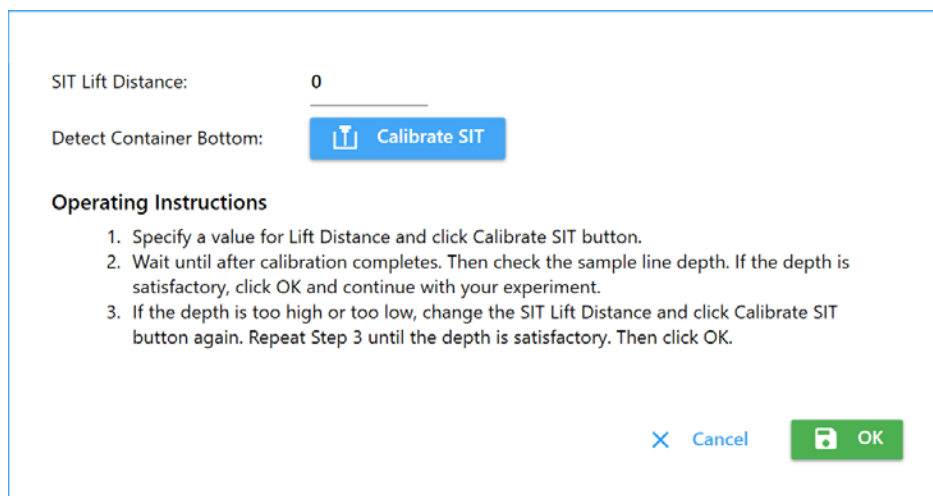
■ **NOTE:** A SIT calibration is performed automatically in the tube that is loaded on the SIP when the system is turned on.

- 1 Load a 96-well plate.
- 2 In the Cytometer tab, from either the QC & Setup or Acquisition module, select Calibrate SIT.



- 3 A dialog appears prompting you to enter the SIT Lift Distance. Enter the distance, then click Calibrate SIT. The lift distance is the distance in millimeters from the tip of the SIT to bottom of the well.

The recommended starting point for the SIT Lift Distance is 1.5 for U-bottom plates. Set the value higher for V-bottom plates. If you experience clogs, try setting the SIT LIFT Distance higher. For information on changing the default SIT Lift Distance see [“Acquisition Preferences”](#) on page 81.



- 4 Check the depth of the sample line in the well. If it's satisfactory, click OK. If it needs to be raised or lowered, adjust the SIT Lift Distance and click Calibrate SIT.

- 5 Repeat step 4 until the sample line depth is satisfactory. Click OK.
- 6 Once SIT calibration is complete, you can run your experiment.

Maintenance

Maintenance Schedule



Any instrument surface in contact with biological specimens can transmit potentially fatal disease. Use universal precautions when cleaning the instrument or replacing parts. Wear suitable laboratory attire such as protective gloves, eyewear, and lab coat.

The 10% bleach solution used throughout the maintenance procedures is prepared by adding 1 part household bleach to 9 parts DI water.

Routine maintenance of the Aurora cytometer includes periodic replacement of parts. For part numbers, see [“Supplies and Replacement Parts”](#) on page 127.

Scheduled Maintenance

The following table describes the scheduled maintenance procedures for your cytometer.

Maintenance Procedure	Description	Frequency
Replace sheath filter	Ensures debris-free sheath fluid	Every 6 months, or as needed
Long Clean	Cleans the fluidic lines with 10% bleach solution	Once a month and prior to service calls

Unscheduled Maintenance

The following table describes the unscheduled maintenance procedures for your cytometer.

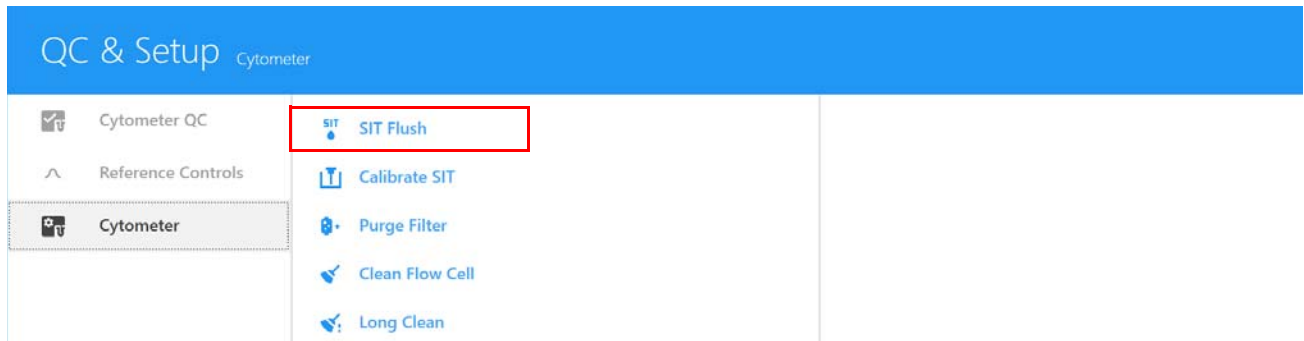
Maintenance Procedure	Description	Frequency
SIT Flush	Backflushes the SIT	As needed, if SIT clogs or after running sticky dyes
Purge Filter	Removes bubbles from the sheath filter	If bubbles are present in the sheath filter, or if the plenum or sheath tank runs dry
Clean Flow Cell	Runs 10% bleach solution followed by DI water through the flow cell	As needed, or after running sticky dyes
Clean external surfaces	Keeps surfaces free from salt buildup	As needed

SIT Flush

A sample line backflush is performed whenever a tube is removed from the SIP after sample acquisition. If the sample line exhibits signs of carryover or becomes clogged after completing an experiment with a sticky dye such as propidium iodide, acridine orange, or thiazole orange, the sample line should be manually backflushed.

If you are using a Loader, a SIT flush occurs over the wash well, located at the back of the Loader plate stage.

- 1 In the Cytometer tab, from either the QC & Setup or Acquisition module, select SIT Flush.



- 2 If carryover or a clog persists, place a tube of 10% bleach on the SIP and acquire at High flow rate for 5 minutes. Afterwards, acquire a tube of DI water at High flow rate for 5 minutes.

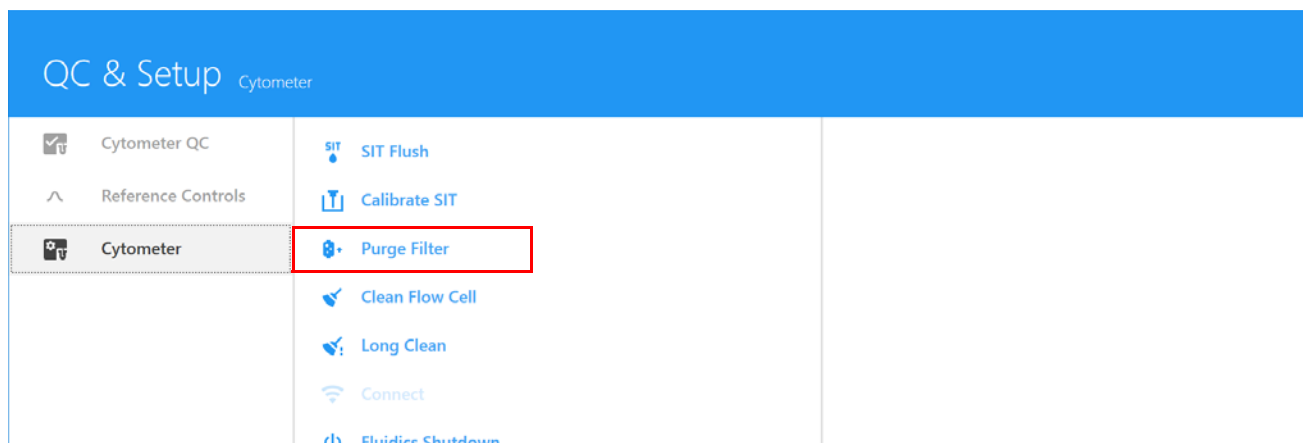
■ **NOTE:** If running large quantities of beads or large cells, we recommend running a tube of 10% bleach followed by a tube of DI water, each for 5 minutes, between experiments.

Purge Filter



Perform this procedure if air bubbles are visible in the sheath filter, or if the plenum or sheath tank have run dry and air is present in the fluidics system.

- 1 In the Cytometer tab, from either the QC & Setup or Acquisition module, select Purge Filter.



The vent valve connected to the sheath filter will open releasing any air bubbles trapped inside the sheath filter.

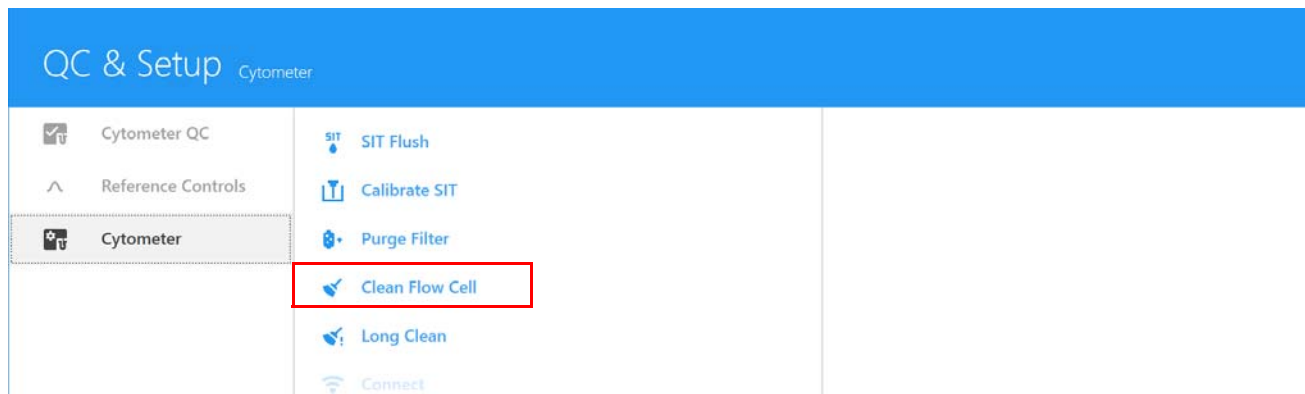
- 2 Repeat the Purge Filter fluidic mode until there are no visible bubbles inside the sheath filter.

Clean Flow Cell

Clean the flow cell after completing an experiment with a sticky dye such as propidium iodide, acridine orange, or thiazole orange. Cleaning the flow cell is also recommended after acquiring large quantities of highly concentrated bead solutions or if you suspect a clog.

If running sticky samples, perform this procedure using a 50% Contrad 70 solution instead of 10% bleach between experiments.

- 1 In the Cytometer tab, from either the QC & Setup or Acquisition module, select Clean Flow Cell.



- 2 Follow the instructions that appear. Load a tube containing 3 mL of 10% bleach solution on the SIP and click Continue.
- 3 When prompted, load a tube containing 3 mL of DI water on the SIP and click Continue.
- 4 Click Done when the procedure is complete.

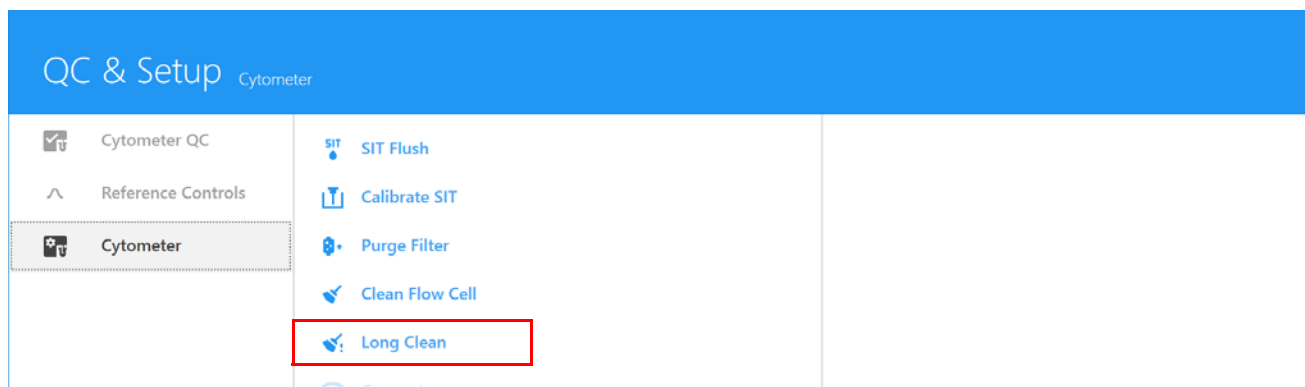
Long Clean

Decontaminate the fluidics system monthly by running the Long Clean fluidics mode. Run the Long Clean just prior to service calls, or if you run high volumes of unwashed samples or samples stained with propidium iodide, acridine orange, or thiazole orange.

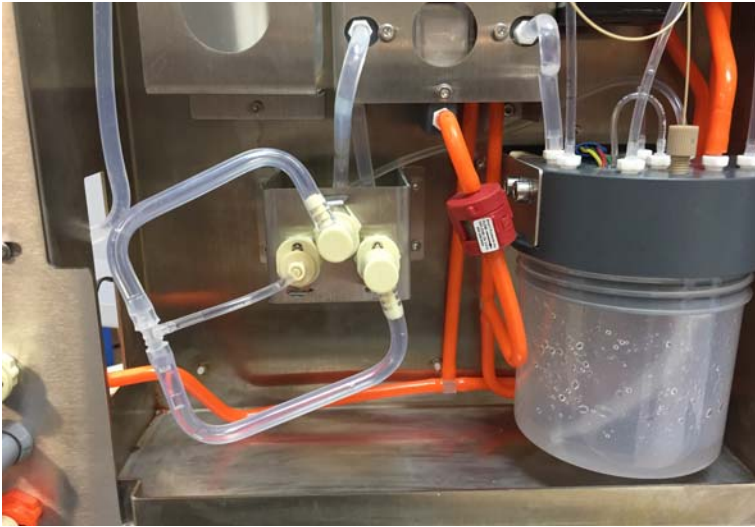


Do not run bleach or detergent through the sheath filter. It is difficult to remove cleaning solutions from the sheath filter.

- 1 In the Cytometer tab, from either the QC & Setup or Acquisition module, select Long Clean.



- 2 Follow the instructions that appear. Prepare the appropriate cleaning tubes and fluidics tanks.
- 3 Empty the waste tank. Replace the sheath filter with the sheath filter bypass (long clean tubing) assembly.



- 4 Detach the sheath tank and replace it with a tank containing a 10% bleach solution.
- 5 Load a tube containing 3 mL of a 10% bleach solution on the SIP.
- 6 Proceed with the Long Clean in the software.
- 7 Once the bleach cleaning cycle is complete, reattach the sheath tank.
- 8 Remove the tube of 10% bleach from the SIP and replace with a tube of 3 mL of DI water.
- 9 Proceed with the Long Clean in the software.
- 10 When prompted, remove the long clean tubing assembly and re-install the sheath filter.

Fluidics Shutdown

Perform fluidic shutdown at the end of each day that you use the instrument. The shutdown procedure thoroughly cleans the fluidics system.

■ **NOTE:** If you are operating in plate mode, you must switch to tube mode before performing Fluidics Shutdown.

- 1 In the Cytometer menu from either the QC & Setup or Acquisition module, select Fluidics Shutdown.
- 2 Load a tube containing 3 mL of 10% bleach on the SIP and click Continue.
A 10% bleach solution is prepared by adding 1 part household bleach to 9 parts DI water.
- 3 Load a tube containing 3 mL of DI water and click Continue.
- 4 Load a tube containing 3 mL of 50% Contrad 70 and click Continue.
- 5 Load a tube containing 3 mL of DI water and click Continue.

- 6 Allow the shutdown procedure to complete, then click Done and turn off the cytometer. Make sure the SIT is submerged in the DI water at the end of the procedure.

The following day when you turn on the system, the startup procedure begins, using the tube of DI water on the SIP.

Cleaning the External Surfaces

Periodically check for saline residue.

- 1 Dampen a cloth with a mild cleaning solution and wipe the surfaces of the instrument.
- 2 Dampen a cloth with DI water and wipe the surfaces again to remove residual cleaning solution.
- 3 Dry the surfaces with a clean, dry cloth.

Inspecting the Fluidics Lines



Check the cytometer periodically for fluid leaks. If any evidence of a leak is detected, contact Cytek Technical Support immediately. Do not attempt to repair the instrument.

- 1 Visually inspect for fluid leaks by looking for small pools of liquid near any of the quick-connects.
- 2 Visually inspect for dried residue or slight discoloration in the spaces surrounding the cytometer.

Replacing the Sheath Filter

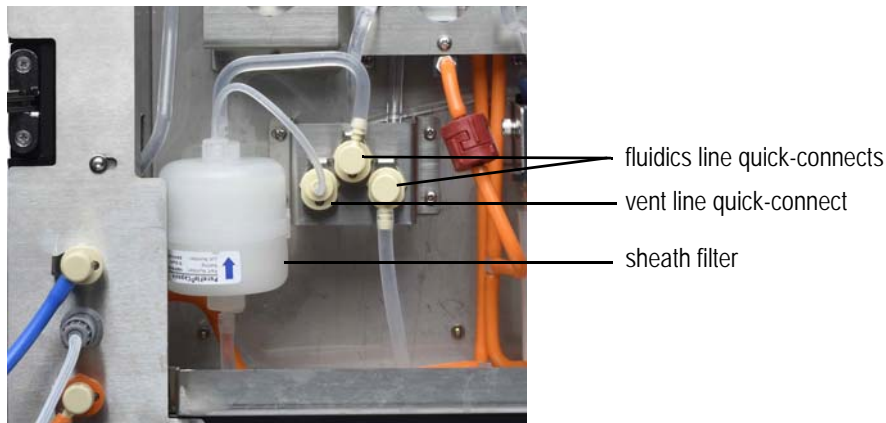
The sheath filter traps debris and air bubbles before they reach the flow cell. Replace the filter assembly every 6 months, or when you see increased sample flow rates or debris in an FSC vs SSC plot.



Wear appropriate safety attire such as protective gloves, eyewear, and lab coat while performing this procedure.

- 1 Turn off the cytometer.
- 2 Open the front cytometer panel.

- 3 Press the two fluidics line quick-connects and the vent line quick-connect to the right of the sheath filter.



- 4 Discard the sheath filter according to standard laboratory protocol and local regulations.
- 5 Install a new sheath filter with the arrow pointing up.
- 6 Restart the cytometer and run the Purge Filter fluidic mode to remove air bubbles (see [“Purge Filter” on page 106](#)). Repeat this step until all air bubbles are purged from the filter.
- 7 Close the front panel.

Replacing the SIT

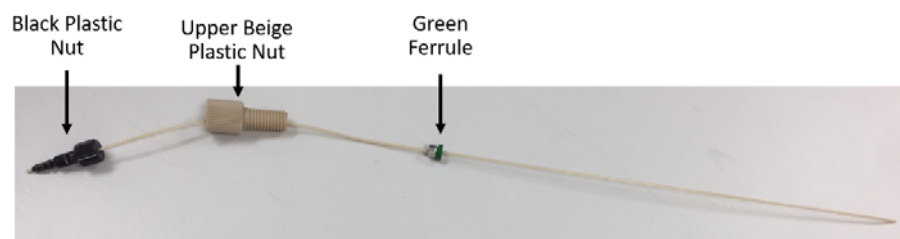
Replace the SIT if the tubing is clogged even after repeatedly cleaning and flushing the SIT.

- 1 Ensure the SIT is extended in a tube of DI water and the cytometer is turned off.

If the cytometer was shut down properly using the Fluidics Shutdown procedure, the SIT will already be extended and left in water.

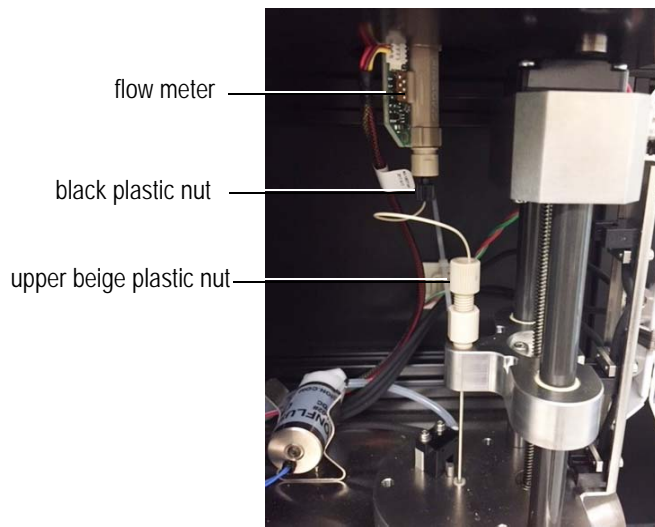
■ **NOTE:** If the SIT is not extended, turn on the cytometer and run Fluidics Shutdown. Then turn off the cytometer.

- 2 Obtain a SIT tubing assembly.

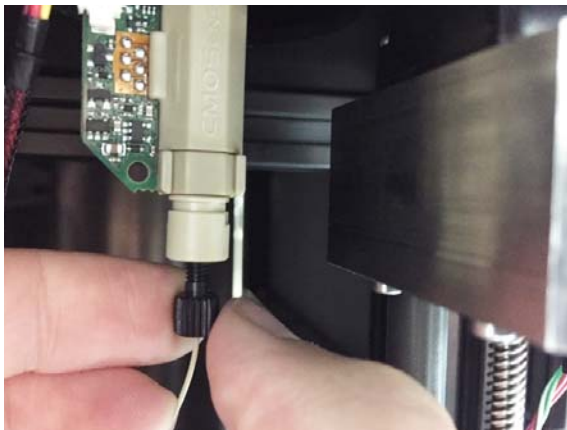


- 3 Open the SIT door. Identify the three components shown in the following figure.

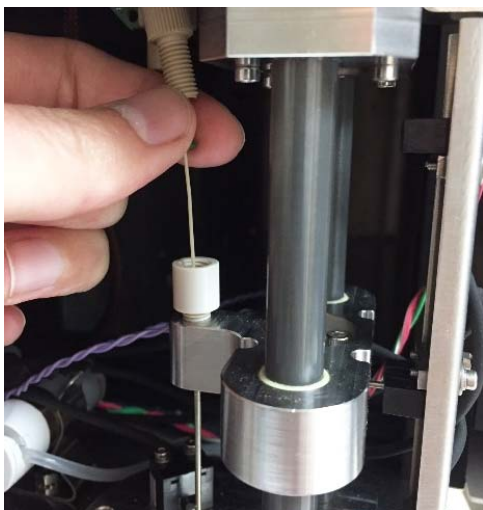
Open the front panel, then open the SIT door. The SIT door is located above the SIP. See “[Front of Cytometer](#)” on page 12.



- 4 Twist off and carefully remove the black plastic nut from the bottom of the flow meter.

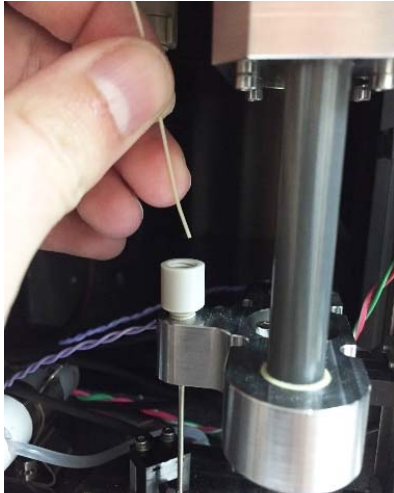


- 5 Follow the tubing from the black nut down to the beige plastic nut. Twist off the beige nut and gently pull it and tubing out from the SIP.

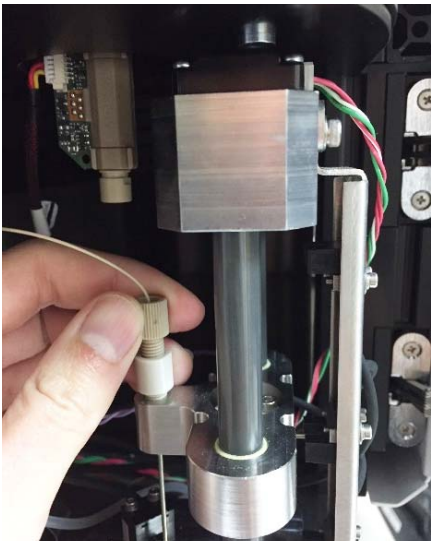


- 6 Discard the SIT tubing assembly.

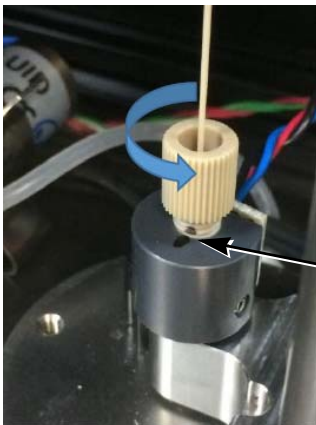
7 Insert the new sample tubing through the hole.



8 Slide the beige nut down into the hole and turn the nut to secure, but do not tighten.

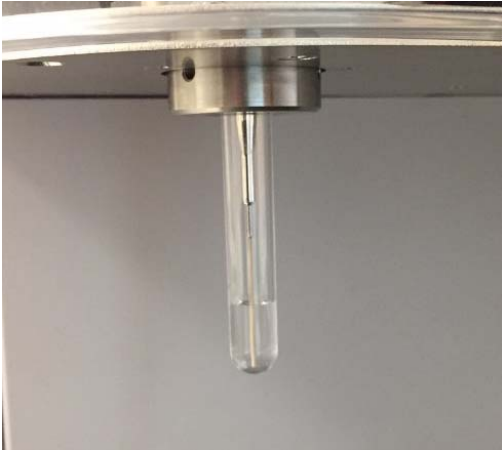


9 Mark the fully screwed in position at the base and on the nut, as shown. Then unscrew the nut 1½ revolutions from the marked position.

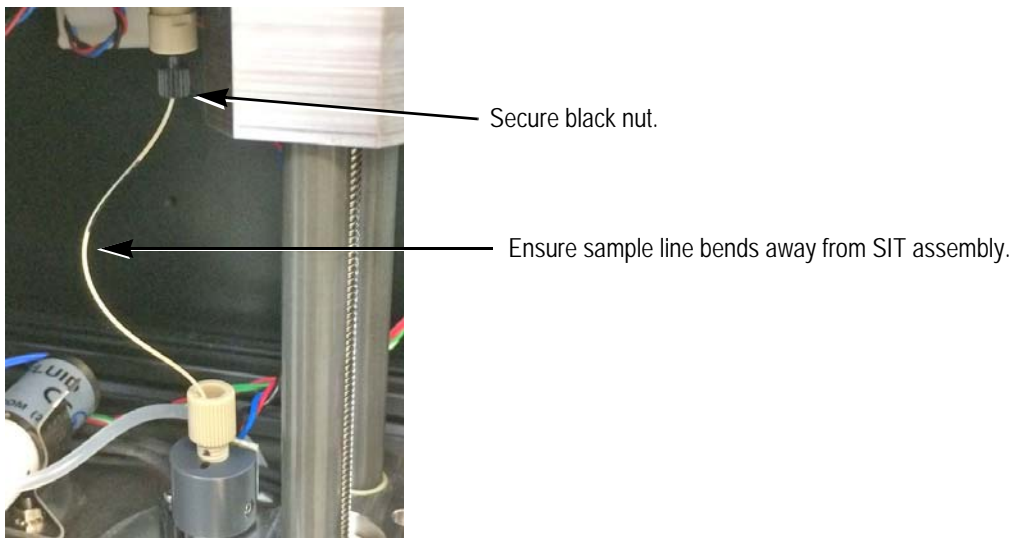


Mark position of screwed in nut.

10 Verify that the tubing contacts the bottom of the tube on the SIP.



11 Secure the black nut to the bottom of the flow meter so it is firmly attached. Make sure the sample line bends away from the SIT assembly when the SIT stage rises.



12 Close the SIT door.

13 Turn on the system. During initialization the system automatically calibrates the SIT depth in the tube loaded on the SIP.

Troubleshooting

This section provides tips to help you identify and resolve issues that might occur on your flow cytometer. If additional assistance is required, contact Cytek Biosciences. Please have the following information available: serial number, error messages, and details of recent performance.

For instrument support within the US, call 1-877-92-CYTEK. Visit our website, www.cytekbio.com, for up-to-date contact information.

Observations

Observation	Possible Causes	Recommended Solutions
Daily QC does not complete	Wrong QC bead sample	Ensure you are running SpectroFlo QC beads.
	Bead sample not properly mixed	Mix the bead sample.
	Bead sample too dilute	Concentrate the bead sample or prepare a fresh bead sample.
	Air bubble in sample line	Run a SIT Flush.
Daily QC failed	Air bubble in fluidics	Run a Purge Filter.
	Dirty flow cell	Run a Clean Flow Cell. If the problem persists, run a Clean Flow Cell using 25%–50% Contrad 70, followed by DI water.
	Questionable sample prep	Verify the sample prep technique.
	Air in sheath filter	Run a Purge Filter.
	Sample not diluted in same fluid as sheath	Dilute the sample in the same fluid as the sheath solution.
Air in sheath filter	Cytometer was not in use for a prolonged period	Run a Purge Filter. Check that all sheath connectors are securely attached. Check for leaks or cracks in the sheath plenum. Replace, if needed.
	Empty sheath tank	Fill the sheath tank. Run a Purge Filter.

Observation	Possible Causes	Recommended Solutions
No events displayed (flow rate lower than expected)	No sample in tube	Add sample or install a new sample tube.
	Sample not properly mixed	Mix the sample to suspend cells/particles.
	Clogged SIT	Run a SIT Flush. Then run a Clean Flow Cell with 10% bleach, followed by a Clean Flow Cell with DI water. If the clog persists, replace the sample line.
	For loaders, the SIT Lift Distance set too low (touching bottom of tube)	Increase the SIT Lift Distance. See “Calibrating the SIT” on page 103.
No events displayed (flow rate normal)	Insufficient gain for threshold parameter	Increase the gain for the threshold parameter.
	Threshold too high	Lower the threshold.
	Laser delay not correct	Ensure the laser delay values match those from the latest Daily QC run. See “Instrument Control” on page 44 for the laser delay location. If the values do not match, rerun Daily QC.
	Threshold set to incorrect parameter	Set the threshold to the appropriate parameter for the application (usually FSC).
	Gated plot with no data in gate	Delete or move the gate.
Low sample event rate	Threshold too high	Lower the threshold.
	Insufficient gain for threshold parameter	Increase the gain for the threshold parameter.
	Sample not properly mixed	Mix the sample to suspend cells/particles.
	Sample too dilute	Concentrate the sample. Set the flow rate to Medium or High.
	Clogged SIT	Run a SIT Flush. Then run a Clean Flow Cell with 10% bleach, followed by a Clean Flow Cell with DI water. If the clog persists, replace the sample line.
Erratic event rate	Partially blocked SIT	Run a SIT Flush. Then run a Clean Flow Cell with 10% bleach, followed by a Clean Flow Cell with DI water.
	Clumpy sample	Vortex, filter, or disaggregate the sample.

Observation	Possible Causes	Recommended Solutions
Data in scatter parameters appear distorted	Air bubble in flow cell	Run a SIT Flush.
	Air in sheath filter	Run a Purge Filter.
	Dirty flow cell	Run a Clean Flow Cell.
	Poor sample health	Check the viability of the cells.
	Hypertonic buffers	Check the pH of the buffers and fixative.
	Incorrect instrument settings	Optimize the instrument settings.
High CVs	Air bubble in fluidics	Run a SIT Flush and a Purge Filter.
	Sample flow rate set to High	Set the sample flow rate to Low or Medium.
	Dirty flow cell	Run a Clean Flow Cell. If the problem persists, run a Clean Flow Cell using 25%–50% Contrad 70, followed by DI water.
	Questionable sample prep	Verify the sample prep technique.
	Air in sheath filter	Run a Purge Filter.
	Sample not diluted in same fluid as sheath	Dilute the sample in the same fluid as the sheath solution.
SIT hitting bottom of well/tube	SIT Lift Distance set too low	Set the SIT Lift Distance to at least 1.5. See “Calibrating the SIT” on page 103.

Glossary

APD	Avalanche photodiode (APD) is a highly sensitive semiconductor electronic device for measuring light intensity. APDs convert light to electricity and can be thought of as photodetectors.
auto-fluorescence	The inherent fluorescence arising primarily from cell structures such as mitochondria and lysosomes. Auto-fluorescence can hinder detection of dim fluorescent signals.
compensation	The process by which spillover fluorescence from secondary parameters is accounted for so that fluorescence values for a parameter represent only the fluorescence of the primary fluorophore.
data file	A collection of measured values from a single sample combined with text describing the data that has been stored as a flow cytometry standard (.fcs) file to disk.
deconvolve	An algorithm-based process used to reverse the effects of convolution (or overlapping) on recorded data.
detector	A device that responds to a specific stimulus. Photodiodes and APDs tubes are two types of detectors in cytometers. They convert light signals into electronic signals.
dot plot	A graphical representation of two-parameter data. Each axis of a plot displays values of one parameter.
electronic noise	Random fluctuation in electronic signals, a characteristic of all electronic circuits.
event rate	The rate at which cells or particles are acquired.
FCS	Flow cytometry standard, a standard format for flow cytometry data files.

filter	An optical device that blocks the passage of part of the incident light, allowing the rest to pass virtually unchanged.
flow cell	The flow cell enables hydrodynamic focusing of the sample so that the individual cells or particles of interest can be interrogated by the laser(s) sequentially.
flow cytometry	A technology that simultaneously measures and analyzes multiple characteristics of single cells or particles as they pass through a laser beam.
flow rate	The amount of fluid passing through a point per unit of time. The Aurora's flow rate is measured by the flow meter.
fluorescence	The emission of light of longer wavelengths that occurs when a substance absorbs light of shorter wavelengths.
fluorophore	A fluorescent dye. A molecule capable of absorbing light energy, then emitting light at a longer wavelength (fluorescence) as it releases this energy.
gain	Amplification of a signal. Increasing gain results in a larger output signal for a given input signal.
gate	A numerical or graphical boundary (region) that defines a subset of data. Gates can be single- or multi-dimensional.
laser	Light Amplification by Stimulated Emission of Radiation. A light source that is highly directional, monochromatic, coherent, and bright. The emitted light is in one or more narrow spectral bands, and with most lasers, is concentrated in an intense, narrow beam.
laser delay	Amount of time between signals from different laser intercepts.
photodiode	A device for measuring light intensity. A photodiode generates an output current proportional to the incident light intensity.
rSD	Robust standard deviation. The robust SD is based on the deviation of individual data points to the median of the population.
reference	Spectral profile of a fluorescent tag in all detectors for all lasers.

resolution	A measure of a cytometer's ability to distinguish between two populations with differing fluorescence or light scatter intensities.
sensitivity	A measure of a cytometer's ability to distinguish particles from background noise. It is often expressed in terms of a minimum number of fluorescent molecules per particle required to distinguish a stained particle from an unstained particle. Sensitivity depends on the instrument, the dye, and the preparation method.
SIP	Sample injection port. The area of the cytometer where the sample is placed.
SIT	Sample injection tube. The probe that pulls sample from the sample tube to the flow cell.
spectral overlap	The phenomenon of different fluorophores emitting light within the same detection range. In multi-color experiments, compensation must be performed to correct for spectral overlap.
spectral unmixing	The mathematical method of deconvolving signals from multiple fluorophores/dyes to differentiate them.
spillover	Emitted light from a fluorophore entering the detector of another fluorophore.
voltage	Measure of electric potential. The voltage applied to a detector affects its amplification gain.

Specifications

Cytometer

Optics

Item	Specification
Optical platform	Fixed optical assembly configured with up to five spatially separated laser beams. Laser delays are automatically adjusted during instrument QC.
Lasers (up to five lasers are available)	355 nm: 20 mW (ultraviolet) 405 nm: 100 mW (violet) 488 nm: 50 mW (blue) 561 nm: 50 mW (yellow green) 640 nm: 80 mW (red) NOTE: Ultraviolet and yellow-green lasers are available for Aurora cytometers only. They are not available on Northern Lights standard configurations.
Beam geometry	Flat-top laser beam profile with narrow vertical beam height optimized for small particle detection.
Emission collection	Fused silica cuvette coupled to high NA lens for optimum collection efficiency to optical fibers.
Forward scatter detector and filter	High-performance semiconductor detector with 488-nm bandpass filter.
Side scatter detectors and filters	Two high-performance semiconductor detectors with 405- and 408-nm bandpass filters.
Fluorescence detectors	High-sensitivity Coarse Wavelength Division Multiplexing (CWDM), 16-channel semiconductor detector array per laser, enabling more efficient spectrum capture for dyes emitting in the 420-nm to 830-nm range. No filter changes required for any fluorophore excited by the on-board lasers. NOTE: See the table on page 71 for the detector bandwidth specifications.

Fluidics

Item	Specification
General operation	Vacuum driven fluidics with the following fluidics modes: Long Clean, SIT Flush, Purge Filter, Clean Flow Cell, Fluidics Shutdown.
Compatible tubes	12 x 75-mm polystyrene and polypropylene tubes
Fluidic reservoirs	4-L fluid tanks with level-sensing provided. Compatible with 20-L sheath and waste cubitainers.
Sample flow rates	Three preset flow rates measured by the flow meter: Tube mode: <ul style="list-style-type: none"> • Low: 15 µL/min • Medium: 30 µL/min • High: 60 µL/min Plate mode: <ul style="list-style-type: none"> • Low: 15 µL/min • Medium: 30 µL/min • High: 100 µL/min
Data acquisition rate	Up to 35,000 events/s (for three-laser systems)

Fluorescence and Scatter Sensitivity

Item	Specification
Fluorescence sensitivity	FITC: <110 molecules of equivalent soluble fluorophore (MEFL) PE: <35 MEFL APC: <15 MEFL Pacific Blue: <190 MEFL NOTE: Measurements performed using SPHERO Rainbow Calibration Particle (RCP-30-5A) based on its single-peak detection channel.
Fluorescence linearity	FITC $R^2 \geq 0.995$ / PE $R^2 \geq 0.995$
Forward and side scatter sensitivity	Enables separation of fixed platelets from noise.
Forward and side scatter resolution	Performance is optimized for resolving lymphocytes, monocytes, and granulocytes.
Side scatter resolution	Capable of resolving 0.2-µm beads from noise.

Workstation

Item	Specification
Operating system	Microsoft® Windows® 10 Pro, 64-bit
Processor	Intel® Core™ i7 processor, 3.6 GHz
RAM	32 GB
Hard drive	500 GB SSD/1TB SATA
Video processor	530 NVIDIA® GeForce
Monitor	32-in UHD 4K Monitor

Installation Requirements

Item	Specification
Dimensions (W x D x H)	54 x 52 x 52 cm (21.3 x 20.5 x 20.5 in) 58.4 x 62 x 52 cm (23.0 x 24.4 x 20.5 in) with the Loader
Weight	61 kg (134.5 lb) to 71 kg (156.5 lb)
Workstation	3.5 x 18.3 x 17.9 cm (1.4 x 7.2 x 7.0 in)
Recommended workspace	152.4 x 61 x 132 cm (60 x 24 x 52 in)
Power	For the five-laser system: 3.4 A at 120 VAC 1.7 A at 240 VAC
Fuse rating	250 VAC, 5 A, size 5 x 20-mm
Heat dissipation	500 W with all solid-state lasers
Temperature	17–28°C
Environmental conditions	Indoor use
Altitude	Up to 2,000 m (6,562 ft)
Pollution degree	2
Humidity	20% to 85% relative non-condensing
Air filtering	No excessive dust or smoke
Lighting	No special requirements

Loader

Item	Specification
Dimensions (W x D x H)	23 x 47 x 28 cm (9.1 x 18.5 x 11 in)
Weight	13 kg (28.7 lb)
Power	160 W (24 V, 6.67 A)
Connection	USB A to USB A
Throughput modes	Default Low Carryover High Throughput
Throughput time	22 min for High Throughput mode (100 µL/min) sampling each well for 2 s.
Dead volume	0 µL for all supported plate types
Carryover	Standard mode: ≤ 0.3% Low Carryover mode: ≤ 0.1% High Throughput mode: ≤ 1%
Plate stage temperature	4–30°C

Supported 96-Well Plates

The system supports the following plates:

Plate Type	Material	Part Number
96-well flat-bottom	Polypropylene	VWR 29444-100
	Polystyrene	VWR 29442-070
96-well U-bottom	Polypropylene	VWR 29444-104
	Polystyrene	VWR 29442-396
96-well V-bottom	Polypropylene	VWR 29444-102
	Polystyrene	VWR 29442-402

Supplies and Replacement Parts

Item	Part Number	Description
4-L Tank	60-30060-00	4-L tank for sheath or waste
Lid for sheath tank	02-62001-00	Lid fits 4-L sheath tank and includes liquid level sensor
Lid for waste tank	02-62002-00	Lid fits 4-L waste tank and includes liquid level sensor
Cubitainer waste cap	N7-32014-0A	Cap fits 20-L waste cubitainer
Cubitainer sheath cap	N7-32015-0A	Cap fits 20-L sheath cubitainer
Reservoir holder	18-22008-00	Holds the sheath and waste tanks
SpectroFlo QC Beads	97-30310-00	2 mL of beads at 10^7 /mL concentration. Beads provide a single peak fluorescence intensity for use with Daily QC.
	N7-97355-0A	2 mL of beads at 10^7 /mL concentration. Beads for use with Aurora systems equipped with a UV laser.
Sheath filter assembly (0.2 μ m)	N7-22006-0A	0.2- μ m sheath filter assembly with quick-connect fittings and manifold
Sheath filter assembly (0.4 μ m)	N7-22016-0A	0.4- μ m sheath filter assembly with quick-connect fittings and manifold
Sample line	N7-22007-0A	SIT tubing assembly
Sheath filter bypass (Long Clean tubing)	N7-22010-0A	Replaces the sheath filter during a Long Clean
25-mm disk filter (20 μ m)	N4-00047-0A	Filter at sheath tubing intake for cubitainer and 4-L sheath tank cap

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