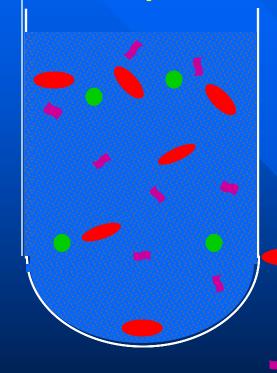
Development of an Immunomagnetic method based on IgG1 Monoclonal antibodies for *Cryptosporidium* and *Giardia*

Philip Scandizzo, Graham Vesey, Mark Gauci, David Baer, James Smith, Duncan Veal*



WATER CONCENTRATE

Concentrated Water Sample



Add Magnetic Beads

GIARDIA CYSTS

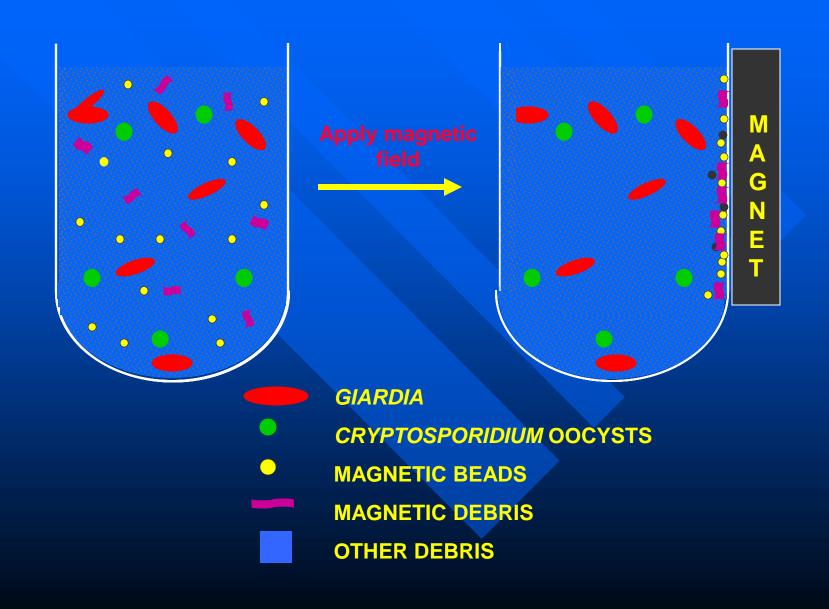
CRYPTOSPORIDIUM OOCYSTS

MAGNETIC BEADS

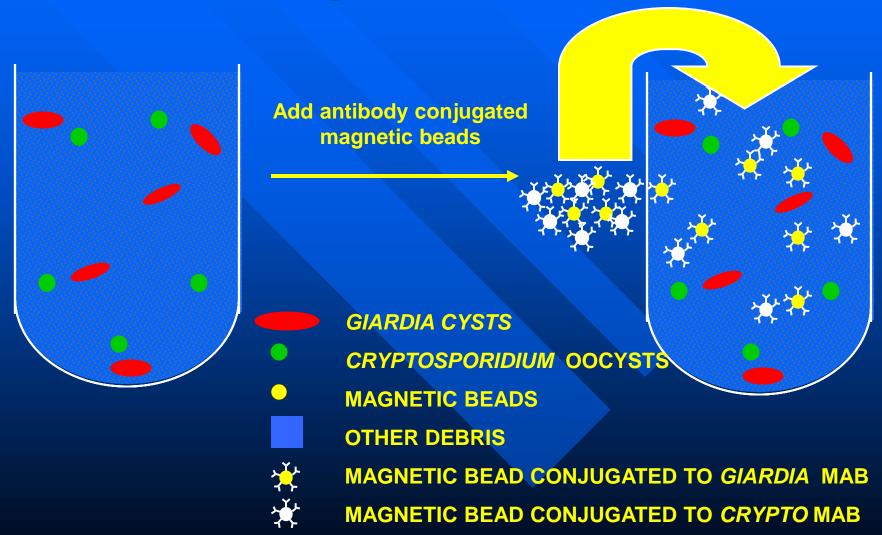
MAGNETIC DEBRIS

OTHER DEBRIS

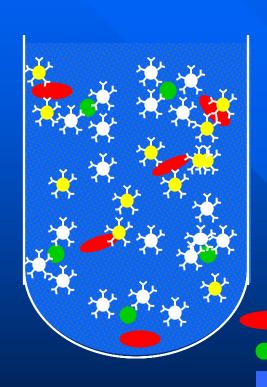
PRECLEAR



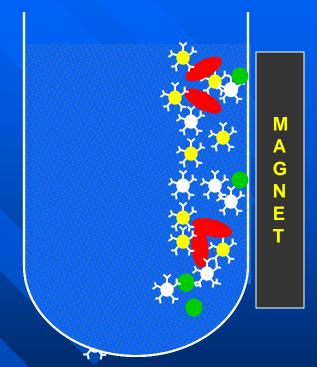
Addition of Conjugated Magnetic Beads



IMS OF CRYPTOSPORIDIUM & GIARDIA



Apply magnetic field



GIARDIA CYSTS

CRYPTOSPORIDIUM OOCYSTS

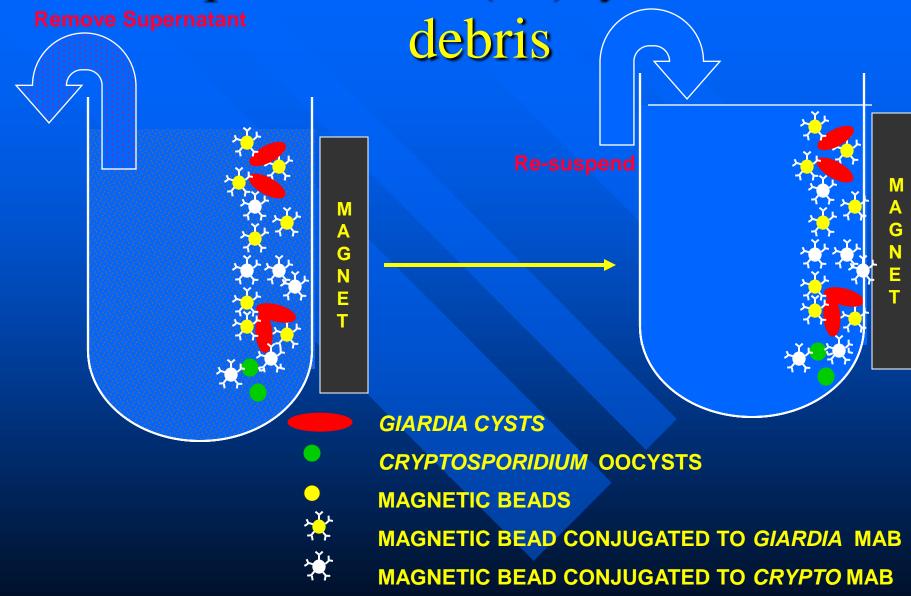
OTHER DEBRIS



MAGNETIC BEAD CONJUGATED TO GIARDIA MAB



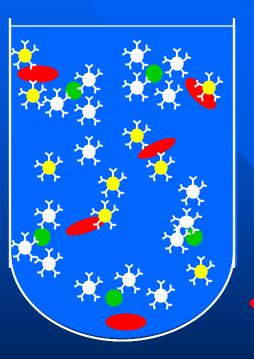
Separation of (oo) cysts from





1 2 3

Acid Dissociation



Add 0.1M HC



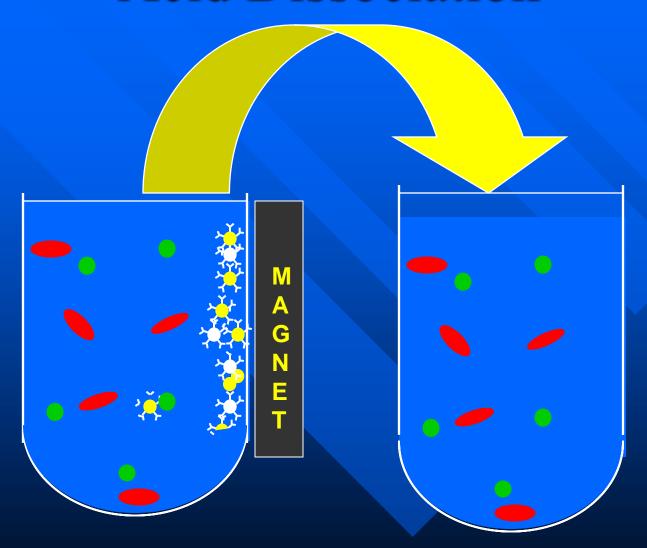
GIARDIA CYSTS





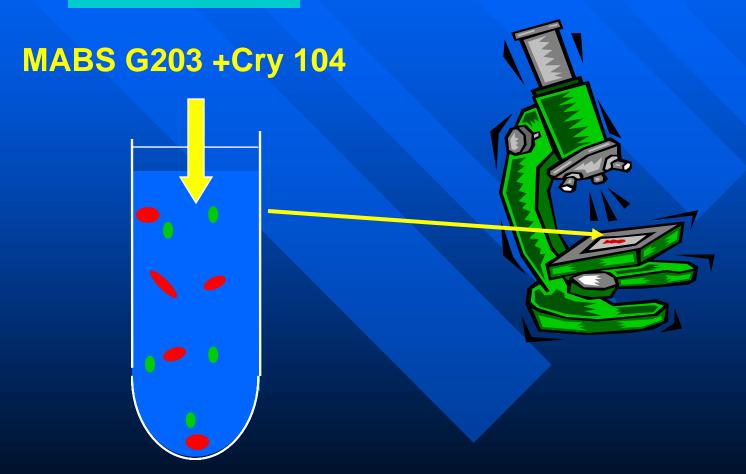
MAGNETIC BEAD CONJUGATED TO GIARDIA MAB
MAGNETIC BEAD CONJUGATED TO CRYPTO MAB

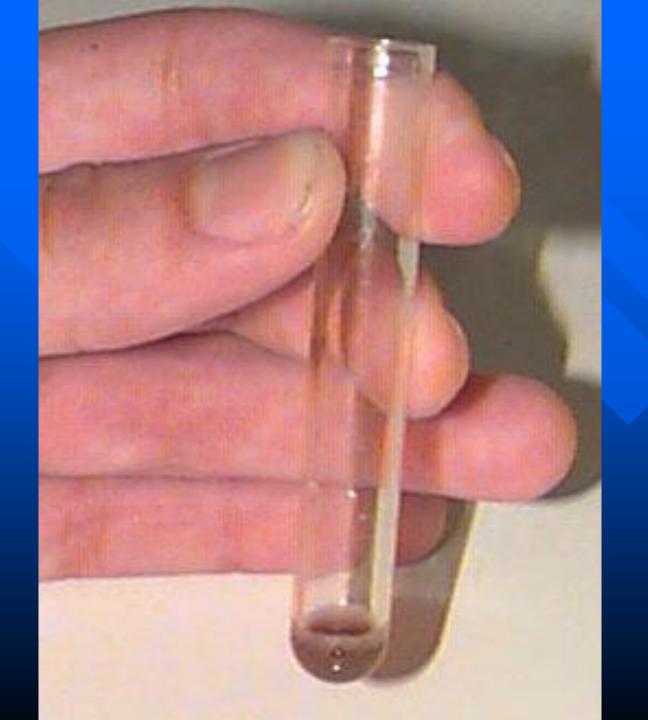
Acid Dissociation



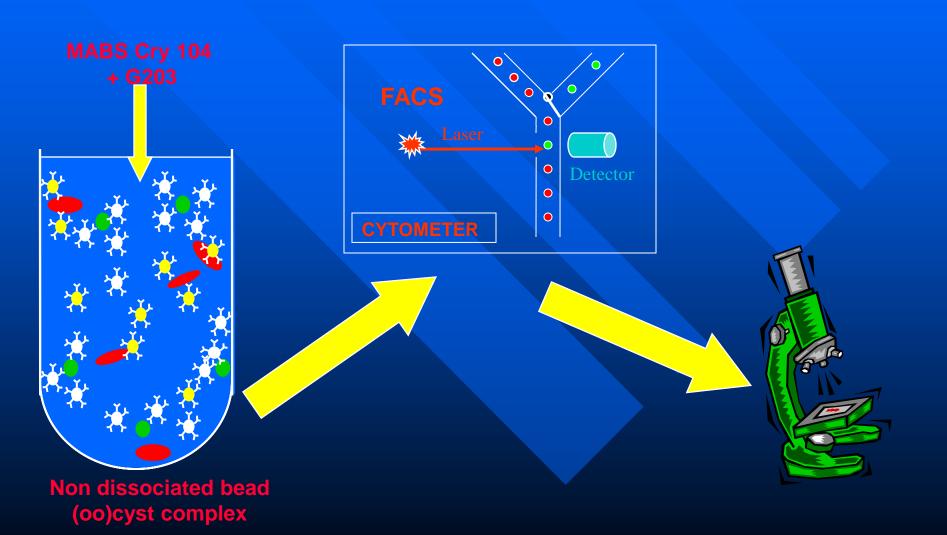
Identification

FLUORESCENT MAB STAINING





Analysis by flow cytometry



Recovery of oocysts from Wimalee Lagoon concentrated water sample and PBS using the Immucell™ IMS Kit.

	PBS	Wimalee Lagoon
Recovery (%)	36 ± 4.6	33 ± 6.4
DAPI Positive (%)	25 ± 4.0	21 ± 2.6
Time Taken microscopy (min)	17 ± 2.1	20 ± 1.5

Recovery of oocysts from Wimalee Lagoon concentrated water sample and PBS using the Dynal® IMS Kit

	PBS	Wimalee
		Lagoon
Recovery (%)	47 ± 6.5	36 ± 3.4
DAPI Positive (%)	28 ± 4.9	25 ± 3.3
Time Taken	19 ± 2.6	20 ± 2.0
microscopy (min)		

Beads and antibodies

Beads	Size (µm)	Supplier	Cost beads / ml	Cryptosporidium and Giardia-specific mAb
Dynal® IgG	4.5	Dynal®,	\$145	CRY 104 (Mac Uni) OW50 (Uni Arizona); Giardia-specific mAb G203 (Mac Uni)
Biomag	1	Perspective Diagnostics	\$6	CRY 104 (Macquarie University)
Spherotech	4-4.5	Spherotech,	\$20	CRY 104 (MacUni)
Dynal® IgM	4.5	Dynal®,	\$145	CRY 26 (Mac Uni)

Recovery of oocysts from Wimalee Lagoon using 0W50-coated anti-IgG Dynal® beads and CRY 26-coated anti-IgM Dynal® beads.

	0W50-coated anti-	CRY 26-coated anti-IgM	
	IgG Dynal® beads	Dynal® beads.	
Recovery (%)	71 (± 12.8)	56 (± 17.5)	
Time Taken FACS	$5.2 (\pm 0.4)$	5.9 (± 0.6)	
(min)			
Time Taken	8.8 (± 1.2)	$7.7 (\pm 0.4)$	
microscopy (min)			

Comparison of recovery of oocysts from Wimalee Lagoon performing IMS with CRY 104-coated Spherotech™, Biomag® and Dynal®-anti-IgG beads.

	Biomag®	Dynal®	Spherotech™
Recovery (%)	91 (± 3.1)	94 (± 3.4)	79 (± 7.5)
Time Taken FACS (min)	11.4 (± 2.3)	10.1 (± 1.4)	12.7 (± 1.2)
Time Taken microscopy (min)	12 (± 1.0)	13 (± 1.5)	12.3 (± 1.5)

Recovery of oocysts from different water types using CRY 104-coated Dynal® beads.

Water Sample	Original sample volume (L)	Recovery (%)	Time Taken FACS (min)	Time Taken microscopy (min)
Wimalee Lagoon	10	90 (± 3)	10.3 (± 2.2)	14.3 (± 2.5)
Drinking Water	100	78.3 (± 5.5)	4.5 (± 1.0)	19.3 (± 5)
River Water	100	92.7 (± 5.8)	5.9 (± 2.1)	14.3 (± 2.5)

Recovery of *Giardia* cysts from different water types using G203-coated Dynal® anti-IgG beads

Water Type	Recovery (%)	Time Taken FACS (min)	Time Taken microscopy (min)
Wimalee Lagoon	86.3 (± 5.5)	5.1 (± 0.5)	3.8 (± 0.2)
Drinking Water	87 (± 7.9)	5.5 (± 0.5)	5.4 (± 0.2)
River Water	78.3 (± 11.9)	5.6 (± 0.7)	4.4 (± 0.5)