

# Equipment and Procedures Overview

## COCVD COBRE Pathology Core



Revised 4/6/17

# Purpose

- To inform you of equipment available for common use and for services provided by the Core.
- To review protocols for use
- To inform you that use of these pieces of equipment and/or services requires that you cite **COBRE grant P20 GM103527** in publishing data that was obtained using any this equipment

# How can you help?

- Notify Wendy (wkatz@uky.edu) of use of common-use equipment by requesting SharePoint reservations (important for maintenance contracts and for general monitoring of equipment)
- Receive training on equipment prior to use to assure that equipment is kept in good operating condition
- Keep things clean
- Be regular monitors yourselves, if you don't like something you see, tell the person using the equipment
- Cite the COBRE grant P20 GM103527 when publishing your exciting findings

# To reserve time on shared equipment (microscopes, microtome)

- Check availability by going to the master list of equipment calendars at <https://cocvd.med.uky.edu/node/58187>
- To access a calendar, log in with your LinkBlue ID, which must be in the MC domain.
- Please notify Wendy of use even if you see something available and jump in at the last minute. Our usage logs help us plan and justify maintenance.
- When you find an available time that works for you, email [wkatz@uky.edu](mailto:wkatz@uky.edu) to request a reservation. Please state:
  - The equipment you want to use
  - The start time you want
  - The end time you want
  - Your PI's name

# Turnaround time

- Because our Core is currently funded by the COBRE grant, projects from COBRE investigators are given first priority.
- Other projects are performed as time permits.
- If a project requires that we cut a very large number of slides, we will alternate between the very long project and small projects so as to keep things moving.

# Processing tissues for paraffin embedding

- A Microm STP120 programmable tissue processor and Leica EG1160 Embedding Center are operated by Core personnel.
- Fixed samples stored in 70% ethanol may be submitted for processing and embedding (see “Preparing and submitting samples for processing,” downloadable from this web page).
- Please submit along with a Core request form, downloadable from the website ([https://cocvd.med.uky.edu/sites/default/files/Pathology%20Core%20Request%20form%202016\\_0.pdf](https://cocvd.med.uky.edu/sites/default/files/Pathology%20Core%20Request%20form%202016_0.pdf)) or emailed to you (request from [wkatz@uky.edu](mailto:wkatz@uky.edu))
- Please note that your IACUC protocol number must be entered on the form.



# Paraffin Sectioning by Core personnel

A Microm HM355S electronic rotary microtome is maintained for use by Core personnel to section your paraffin embedded samples.

- Please indicate on your request form:
  - IACUC protocol number
  - Number of slides desired
  - Desired thickness (5 microns is standard)
  - Number of sections per slide (2 is standard)



Note: if your project requires more than 100 slides, please provide slides and storage boxes for slides in excess of 100.

# Microtomes available for shared use

A Shandon Finesse and Shandon Finesse ME microtome are available for use by approved researchers. Please contact [wkatz@uky.edu](mailto:wkatz@uky.edu) for training and reservations.





# 5<sup>th</sup> floor Wethington microscopes

## General Policies:

- Each microscope has filter controls, etc. in slightly different places. The newer Nikons have a special flip-down or slide-in lens for the 4x objective.
- If you are using a particular microscope for the first time, ask Wendy for an orientation ([wkatz@uky.edu](mailto:wkatz@uky.edu)).
- Please contact Wendy for maintenance, technical or software concerns.



# General microscope policies, cont'd

- Please cite COBRE grant P20 GM103527 when publishing data and images obtained using these microscopes.
- Please move the objective to 4X to insert slides and remove slides from the stage.
- Please do not remove objectives from the microscopes.
- If you use an oil objective (100x or 60x) do NOT use the 20 or 40X objectives afterward as the oil will fog the objectives.
- Use alcohol or Nikon cleaning solution to remove oil. Do not use Windex or any ammonia based cleaner!
- If you aren't sure ASK.....

# Room 551: Nikon Eclipse 80i Upright with fluorescence

12 MP color camera.

Objectives:

- 4X Plan Fluor
- 10X Plan Fluor
- 20X Plan Apo
- 40X Plan Apo
- 60X\*, 100X\*

\*by prior arrangement



Motorized stage with image stitching software

# Additional objectives for Eclipse 80i

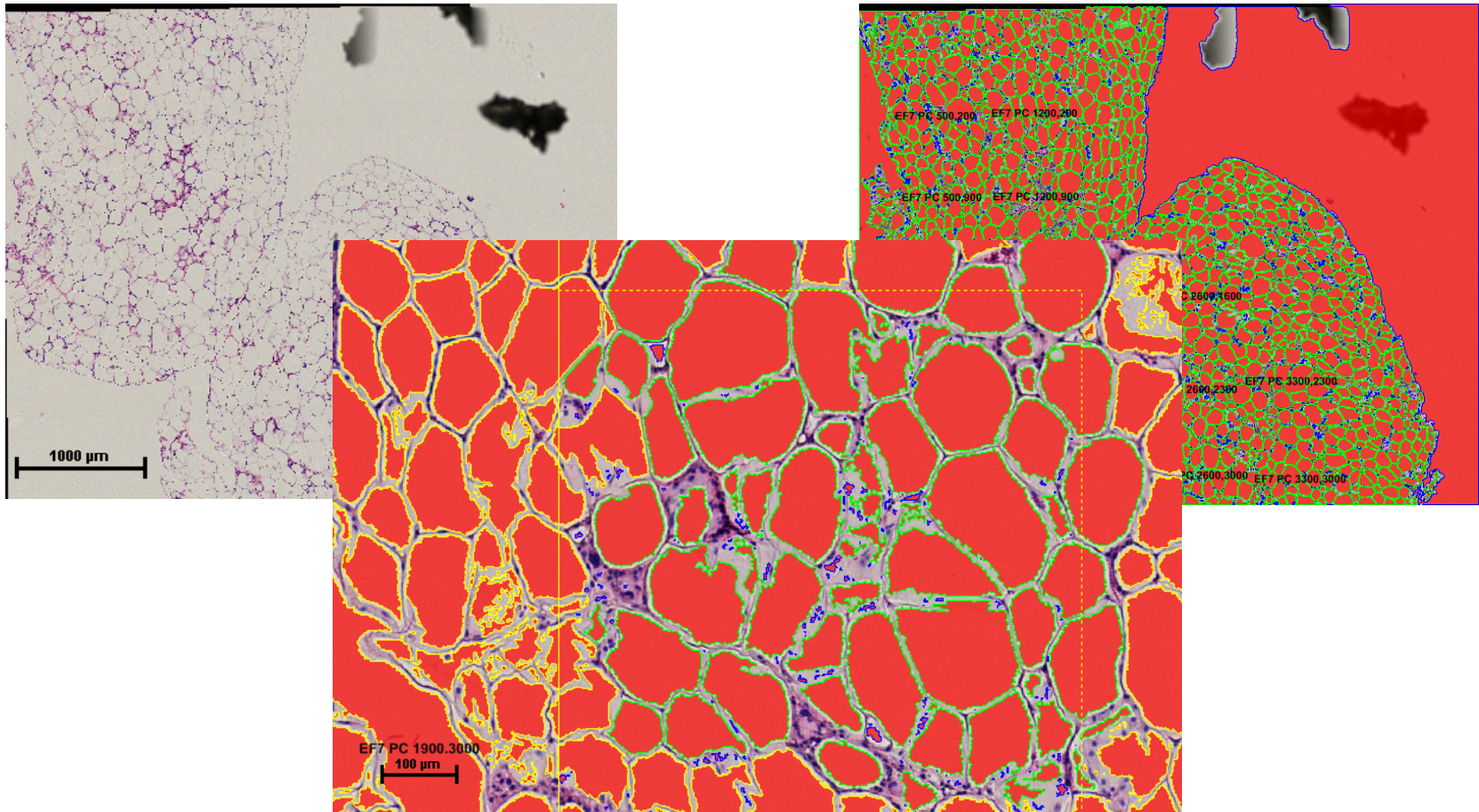
- 60X Plan Apo
  - 100X Plan Fluor
- (both oil objectives)
- For use on *upright* (Eclipse 80i) microscope



- **If you use oil on your slide do NOT use the 20 or 40X objectives afterward as the oil will fog the objectives.**
- Use alcohol or Nikon cleaning solution to remove oil from objectives. **Do not use Windex or any ammonia based cleaner!**
- Wendy has them in a locked cabinet. To use them, make arrangements with Wendy at least 24 hrs in advance

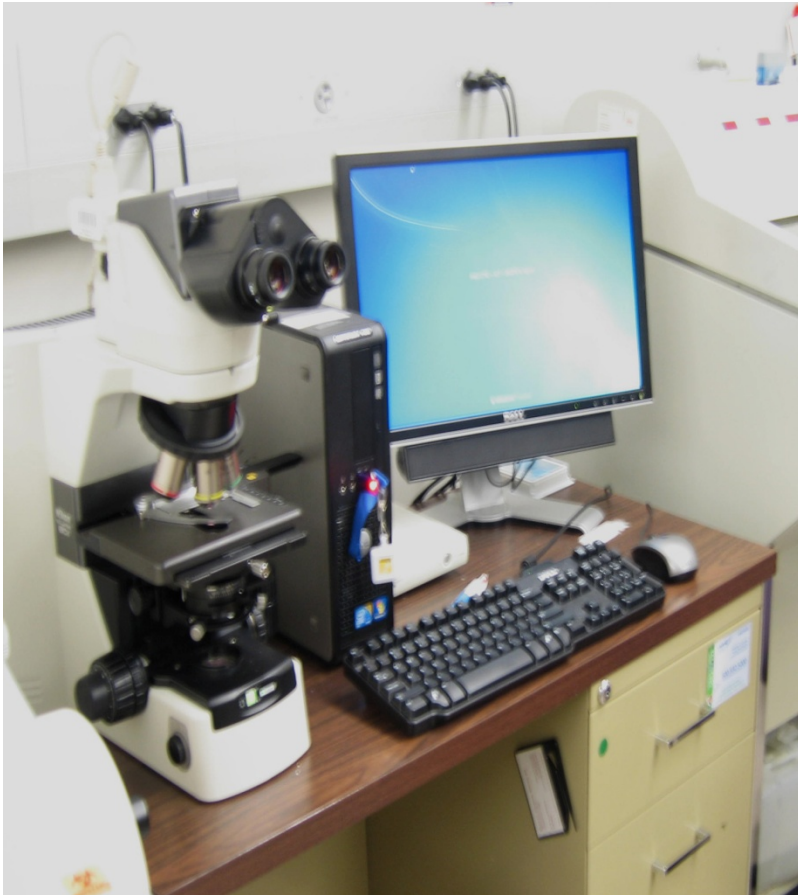
# Motorized stage on Nikon 80i in 551

- Joystick control
- Rapid navigation options in software
- Permits automated stitching of multiple fields
- Please contact Wendy for training and technical issues



# Room 564 Nikon Eclipse 80i

Located to the left of the cryostat



- Color camera
- Objectives:
  - 4X Plan Fluor
  - 10X plan Fluor
  - 20X Plan Fluor
  - 40X Plan Fluor

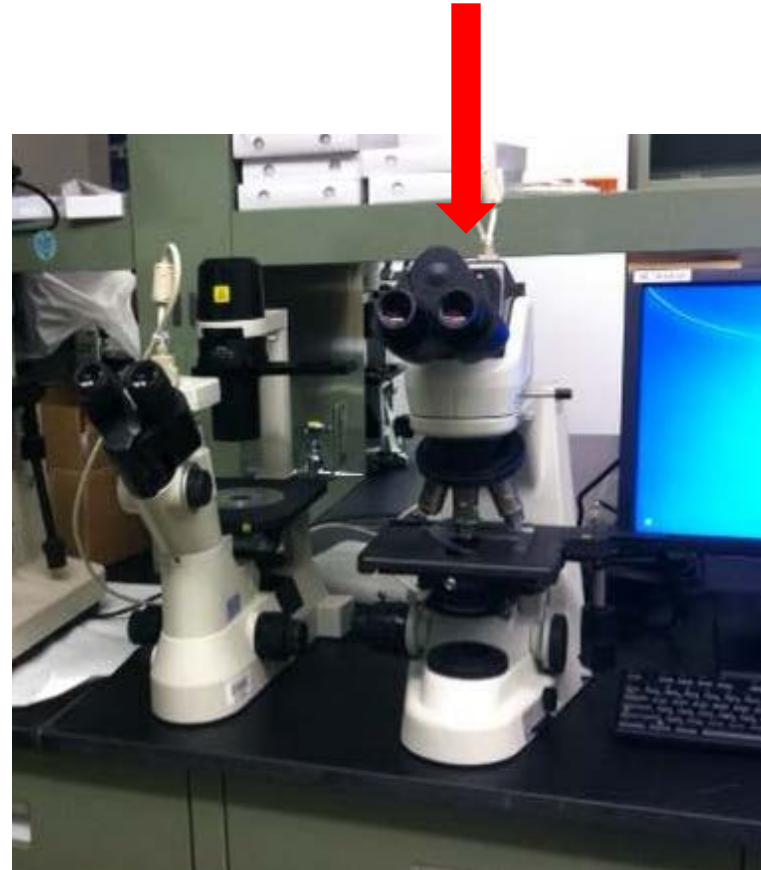
# Room 564: Nikon Eclipse 55iUpright

Located between embedding center and microtome.

12 MP Color camera.

Objectives:

- 4X Plan
- 10X Plan
- 20X Plan
- 40X Plan
- 100X Plan (Oil)



**If you use oil on your slide do NOT use the 20 or 40X objectives afterward as the oil will fog the objectives.**

**Use alcohol or Nikon cleaning solution to remove oil from objectives. Do not use Windex or any ammonia based cleaner!**

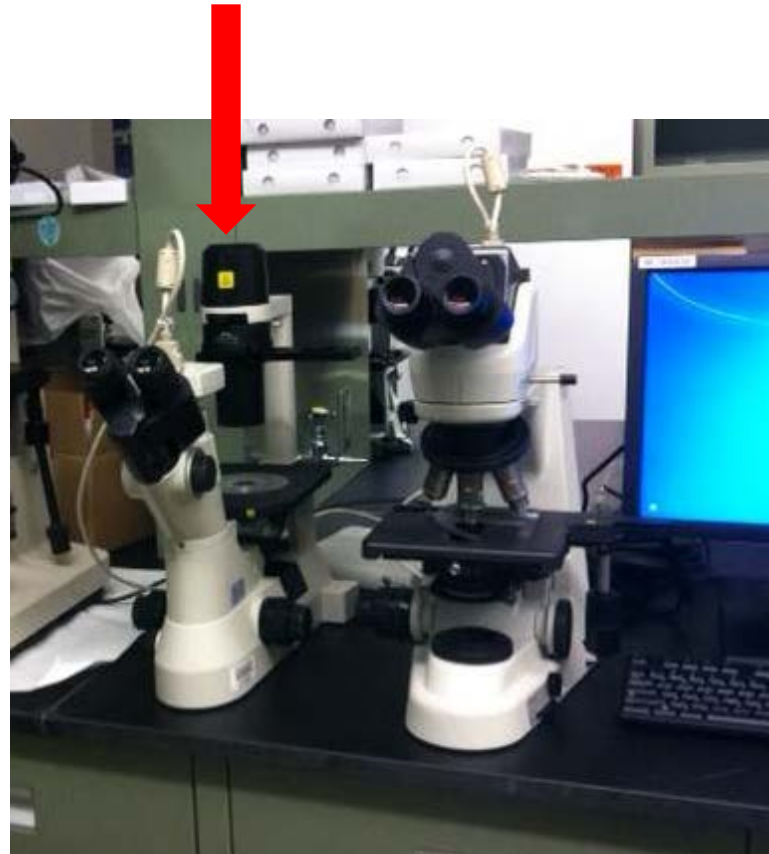
# Room 564: Nikon TS100 Inverted microscope

Located between embedding center and microtome.

Color camera.

Objectives:

- 2X Plan
- 4X Plan Fluor Phase
- 10X Plan Fluor Phase
- 20X Plan Fluor Phase
- 40X Plan Fluor Phase





# 564 TS100 Inverted Microscope (cont'd)

- Please note that there is a slider having different positions (marked on the slider) for the different powers of phase contrast objectives
- This microscope is unique among all of ours in having a knob (on the right, below the binoc) to control whether the slide is visible through the eyepieces, or through the camera.