

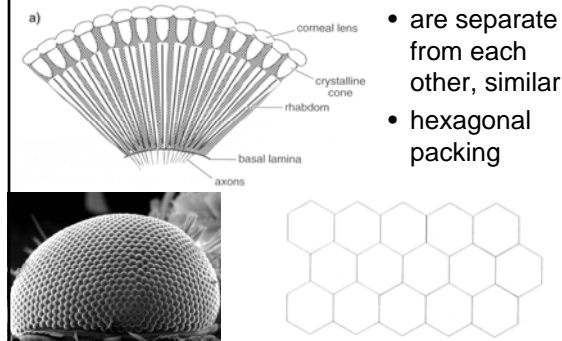
INSECT SENSES

- Vision
- Smell
- Hearing (+insect sounds)

Vision

- General structure and function of eye
- Seeing polarized light
- Adjustments to light levels
- Limits to resolution
- Processing and image formation

ommatidial units in eye



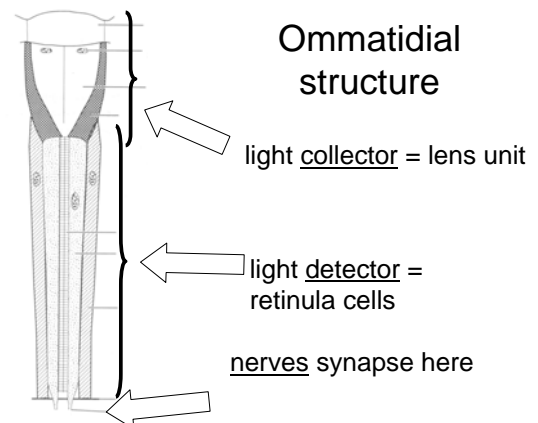
- dragonflies have about 10K ommatidia

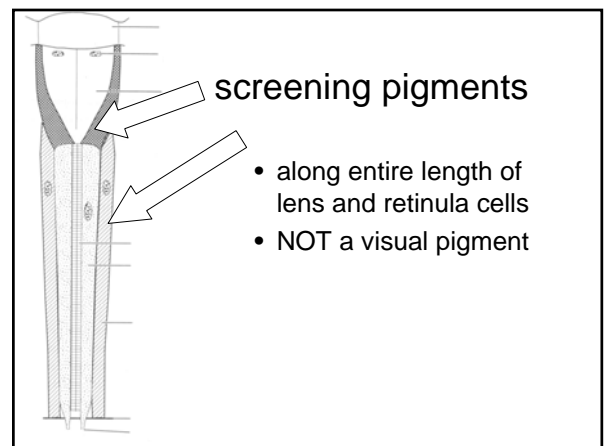
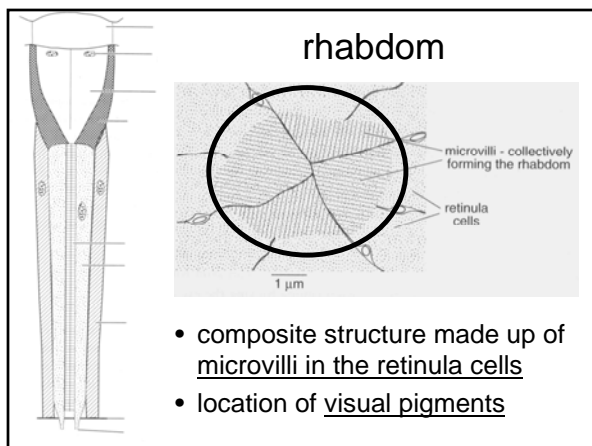
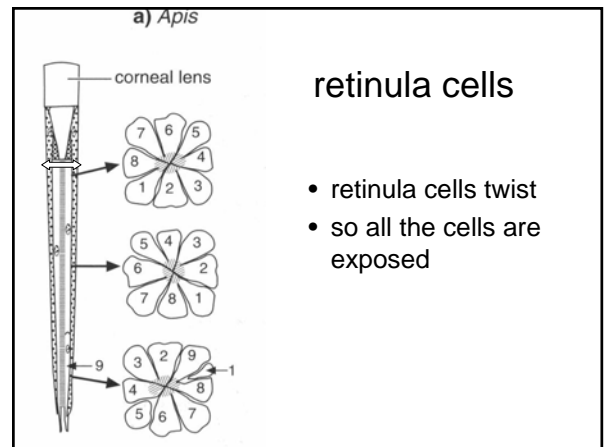
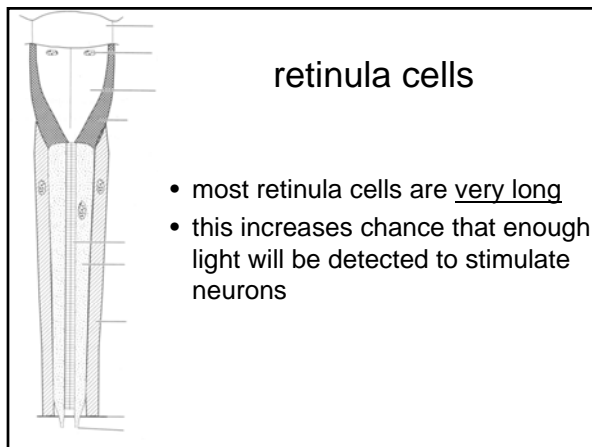
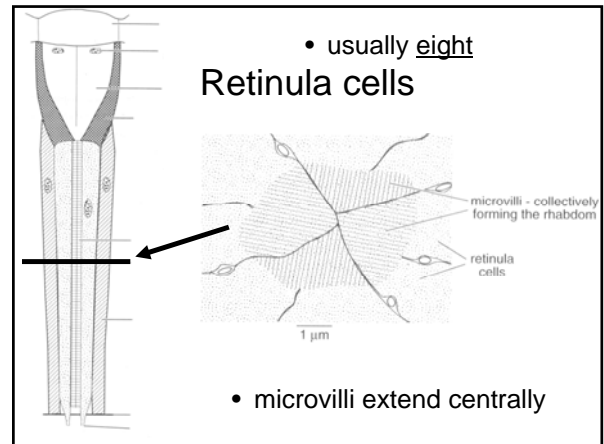
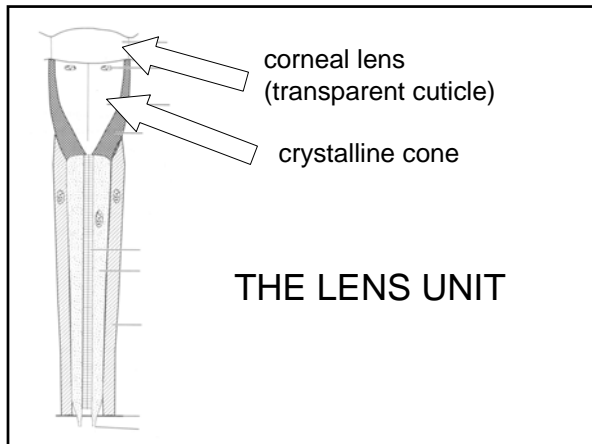
Army ants

have very few ommatidia, sometime just 1

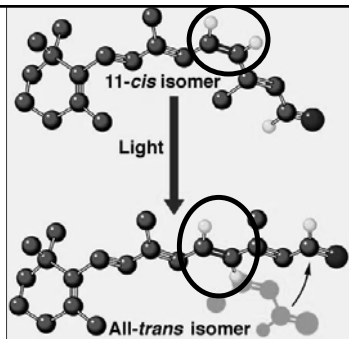
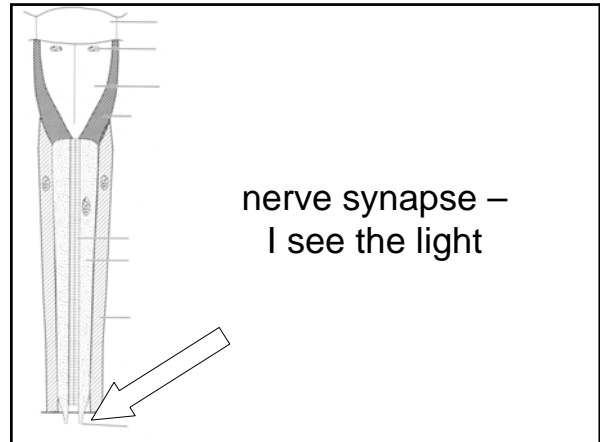


Ommatidial structure



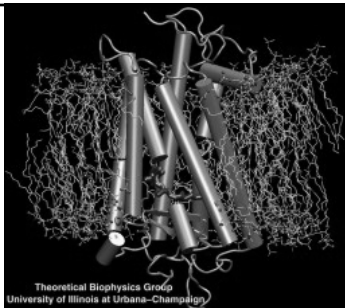


how does light get transduced into neural signal?



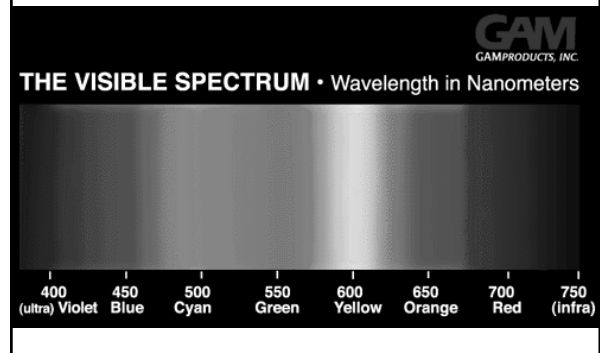
Transduction made possible by RETINAL which has two conformations – one is stable and the other is not.

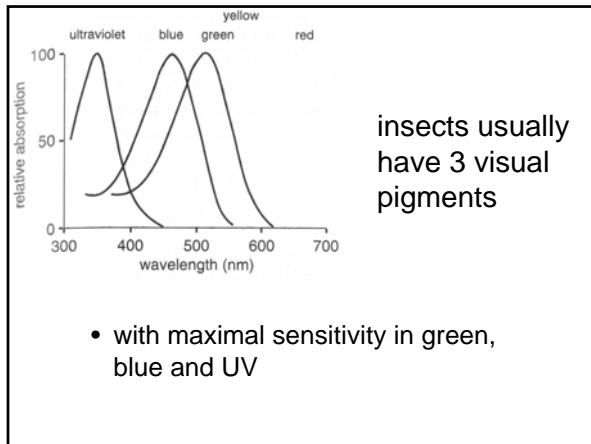
- light converts unstable form to stable trans-retinal
- change in conformation triggers depolarization of the nerve



- Retinal is linked to the visual pigment rhodopsin
- Different rhodopsins responds to different wavelengths

What colors can insects see?



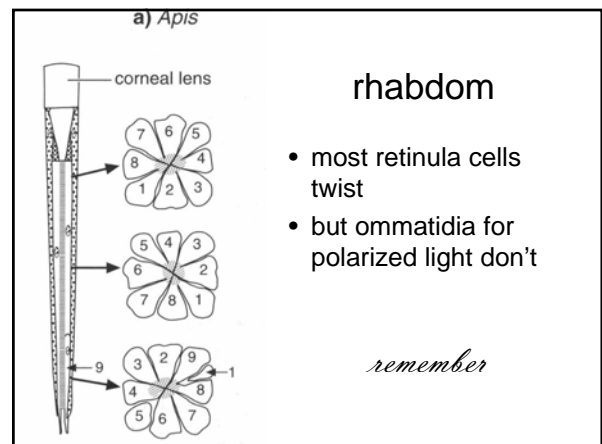
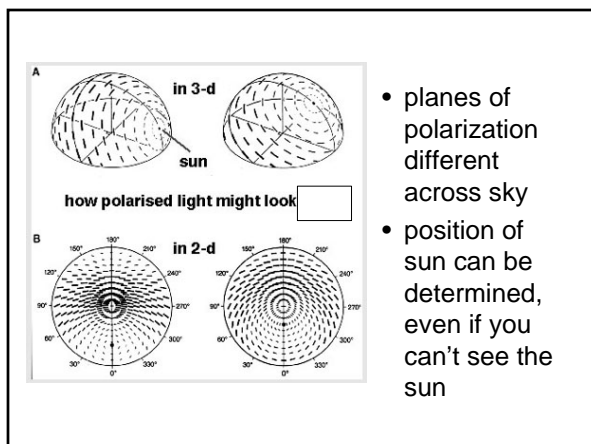
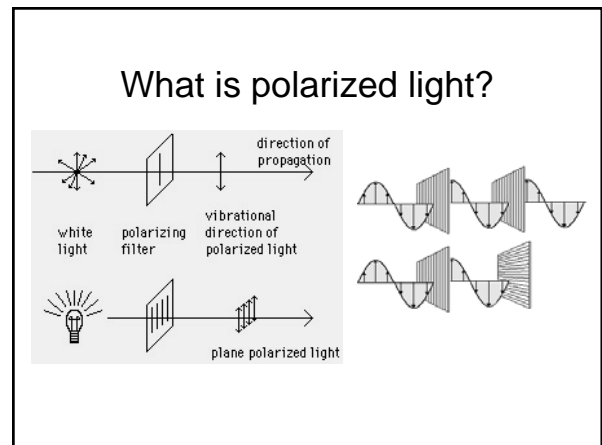


different pigments in different cells

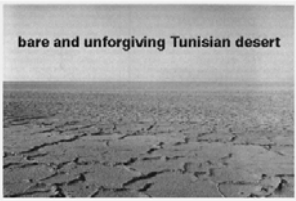
- it is important that different pigments be in different retinula cells
- why?

Many insects detect polarized light

- most studied in social insects which use it as a navigational aid


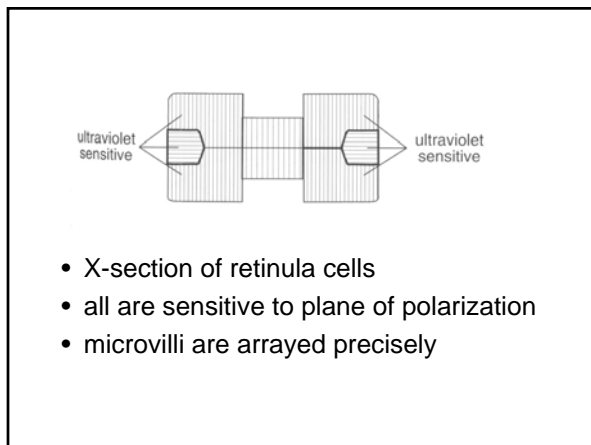
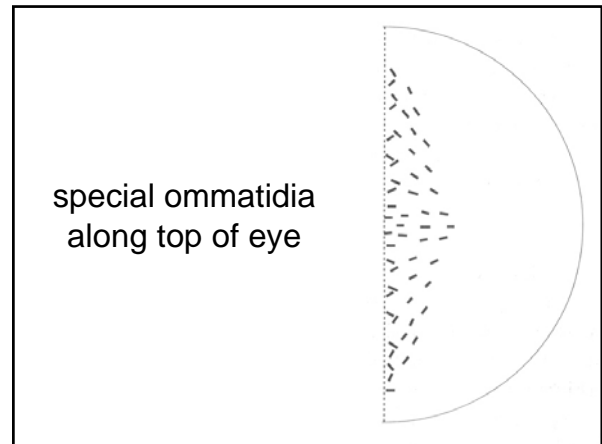


bare and unforgiving Tunisian desert

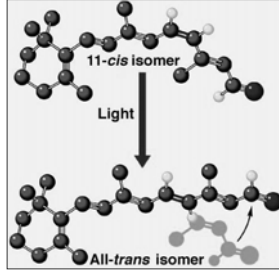
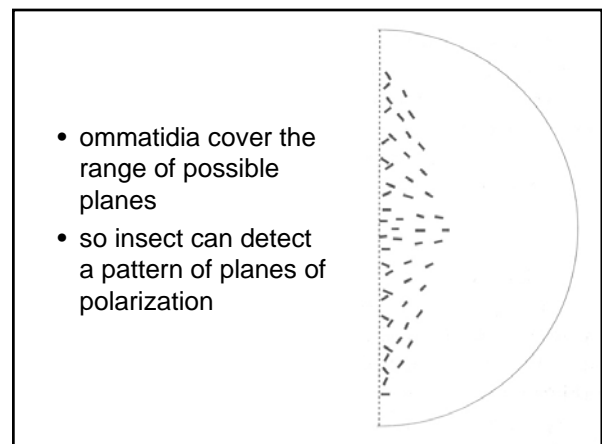
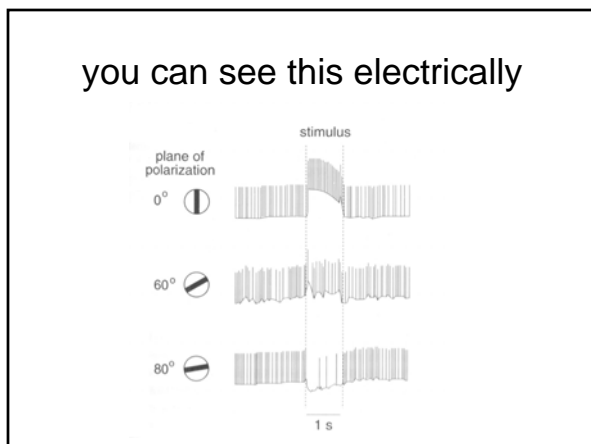


Cataglyphis

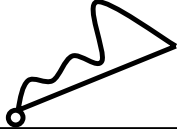
- navigates using polarized light
- few landmarks in environment
- it's @#\$%& hot
- speed essential
- return home in straight line

- orientation of microvilli important because molecules of retinal are oriented along their long axis AND
- pigment is stimulated only if struck by light vibrating in the plane of its long axis

Cataglyphis



- somehow uses polarization map for navigation
- remembers it on way out
- goes straight home
- =path integration

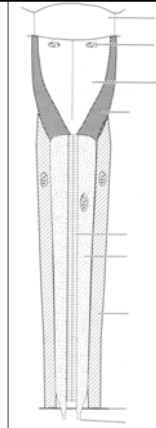
Pedometer



Light Control

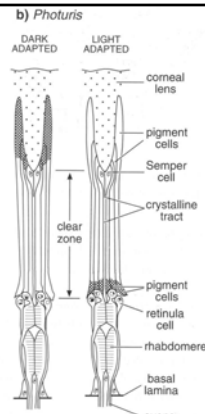
apposition eye

- found in day-flying insects
- best suited for high light levels
- light entering om. can only trigger its own nerves
- screening pigments prevent stray light



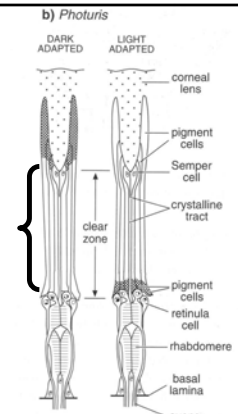
superposition eyes

- best for low light levels – nocturnal insects
- works in two modes
- light and dark adapted



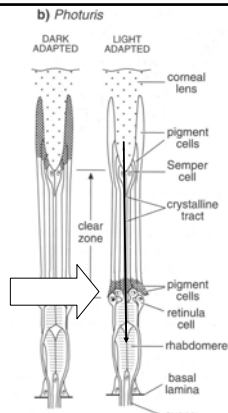
superposition eyes

- lens
- retinula cells
- screening pigment
- CLEAR ZONE



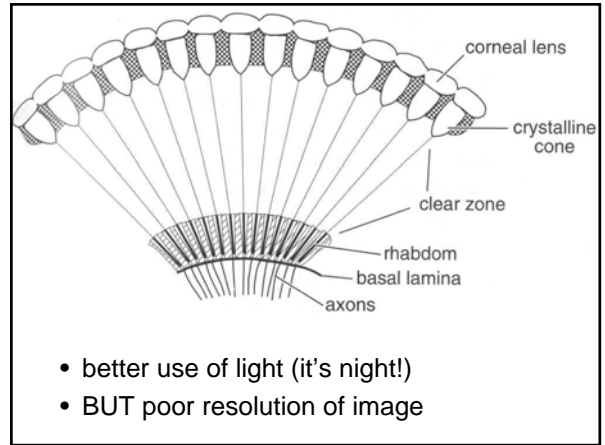
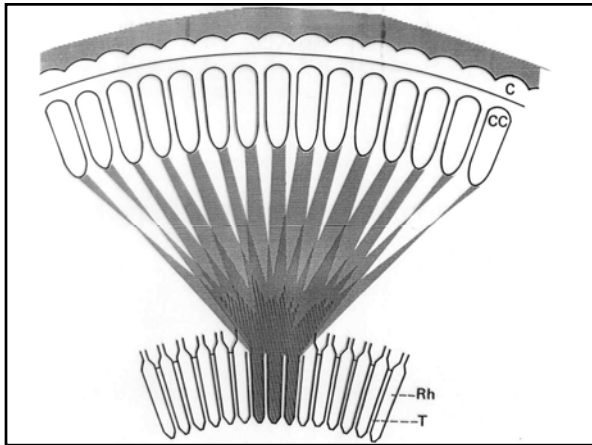
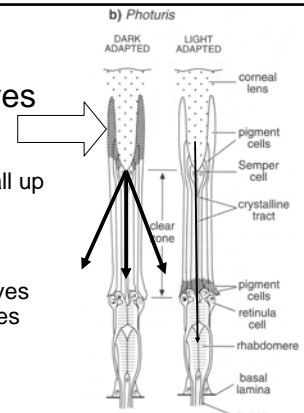
Light adapted superposition eyes

- In the day, pigment is near retinula cells
- light can pass only through lens to its own rhabdome



Dark adapted superposition eyes

- at night, pigment is all up around lens
- light is free to move throughout eye
- each rhabdom receives light from many lenses



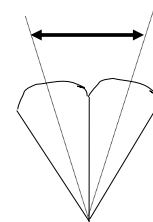
- better use of light (it's night!)
- BUT poor resolution of image

What determines the quality of the image formed by an insect's eye?



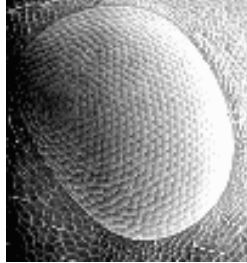
What determines resolution?

- interommatidial angle



for an eye of a particular curvature,

- smaller facets will result in smaller angle and more facets



10 (5k)

25 (33k) 50 pixels/inch

(135k)

BUT, there is a lower limit to diameter

- increased diffraction causes loss of useful light



facet diameter varies

- between species
- between sexes
- on a single eye

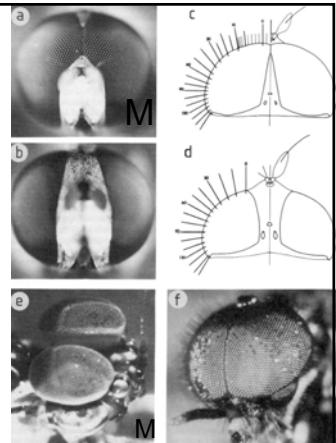
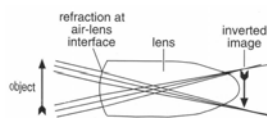
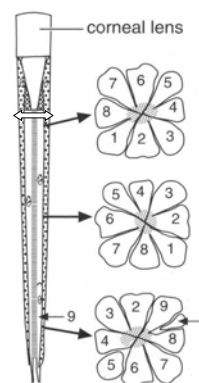


IMAGE PROCESSING How it all works



- inverted image forms just behind lens

a) *Apis*

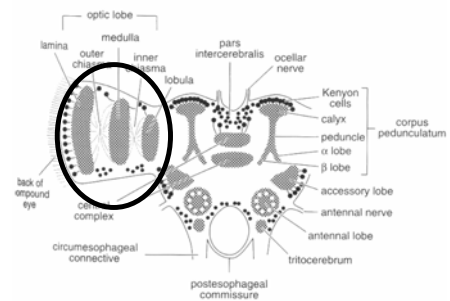


retinula cells

- retinula cells twist as they extend towards nerve
- so all the cells are exposed to the light and image is not preserved

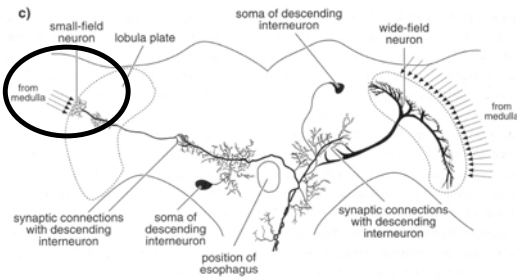
How is the image put together?

Processing, processing, processing

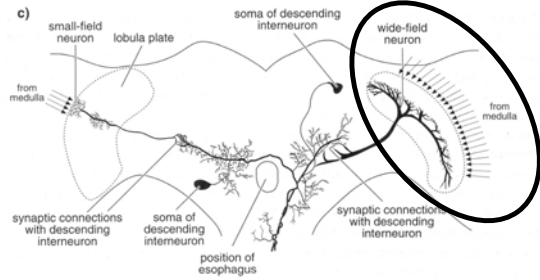


example in the fly lobula

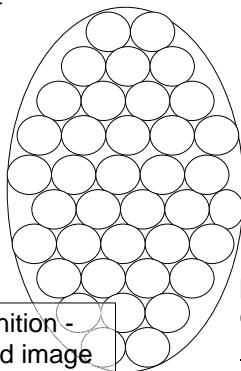
- small field neurons connect with relatively few columns (20-100)



- wide field neurons connect with many columns: 3 run one way, 9 the other

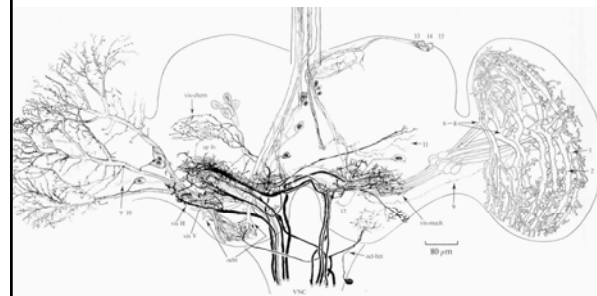


coverage of small field neurons in medulla

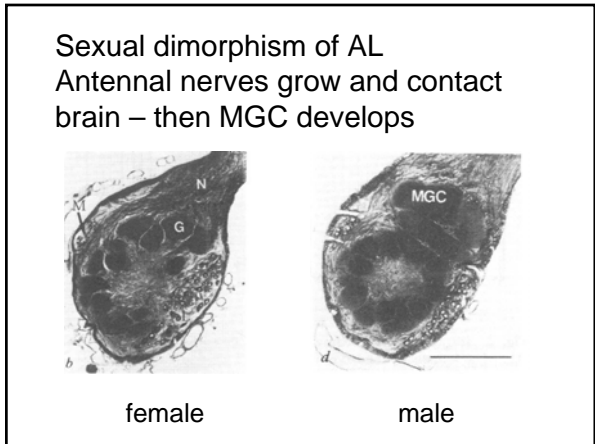
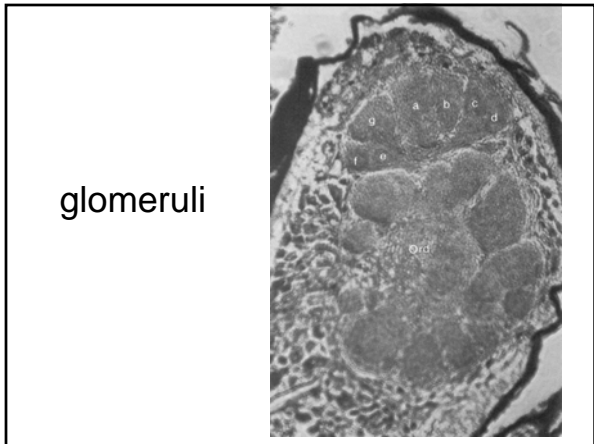
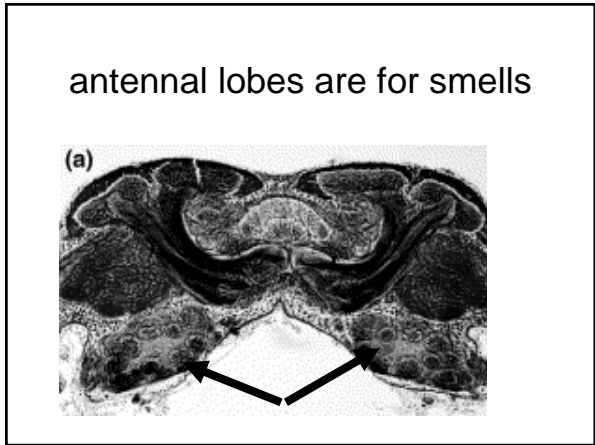
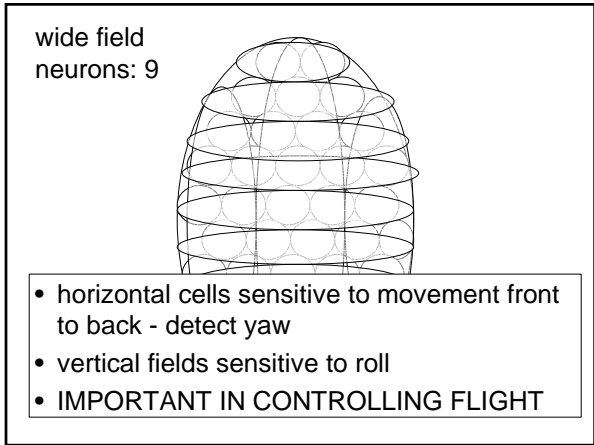
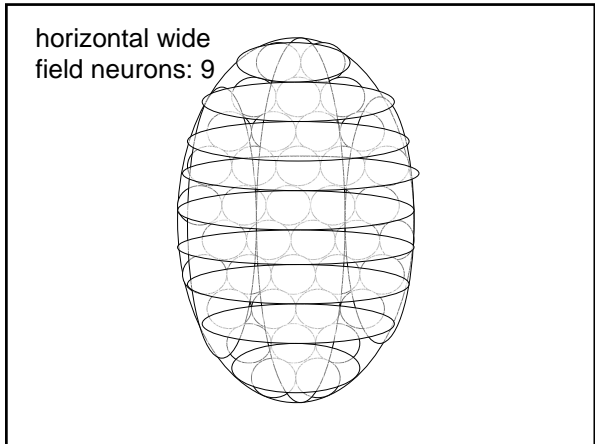
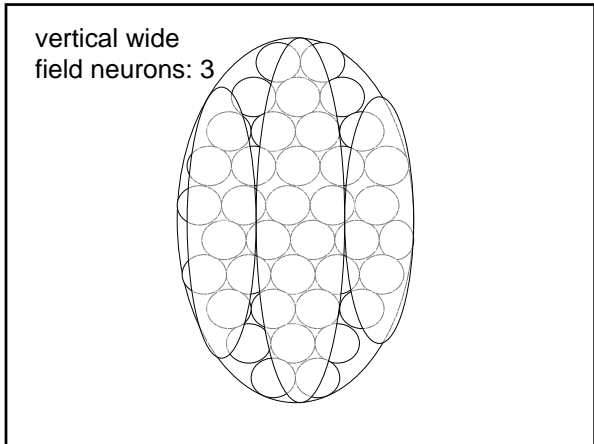


pattern recognition - coarse grained image

retains general patterns of ommatidia - retinotopic mapping



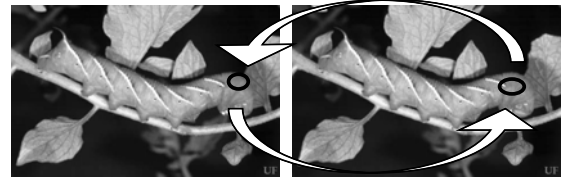
wide field neurons



How does the AL know to develop the MGC?

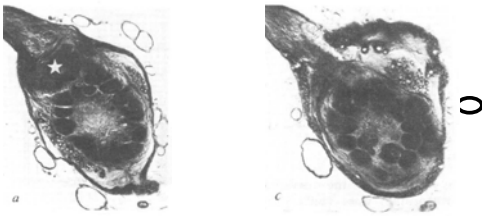
- Because the brain tissue is male?
- Or because the developing antennae induces it?

transplant experiment with antennal imaginal discs



- female larva
- male larva

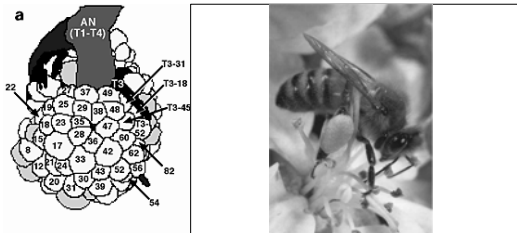
male antenna tissue induces male type brain development in AL



- female larva

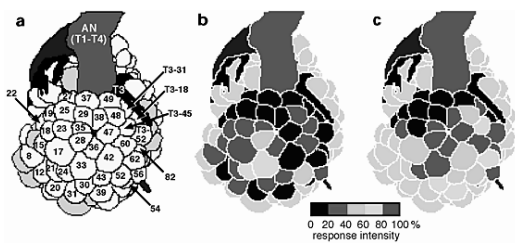
male larva

honey bee - odor specific code



38 most active glomeruli

honey bee - odor specific code

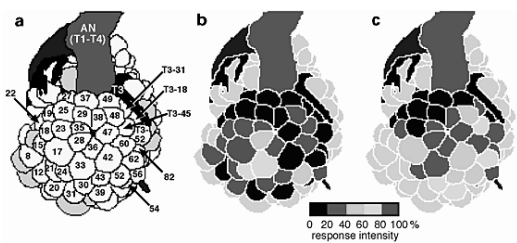


38 most active glomeruli

1-octanol, 21 individuals

clove oil, 5 individuals

honey bee - odor specific code

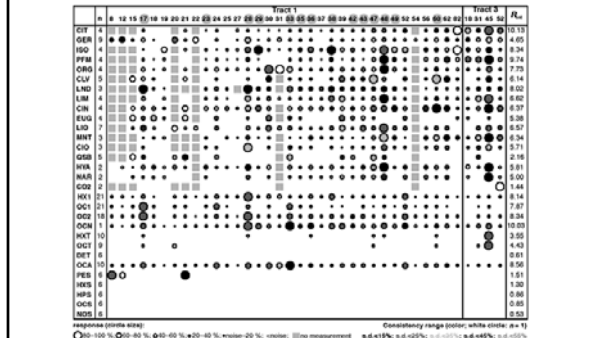


38 most active glomeruli

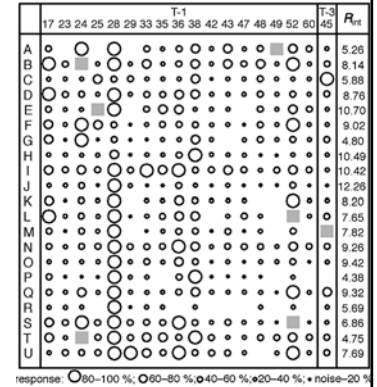
1-octanol, 21 individuals

clove oil, 5 individuals

patterns produced by 30 compounds
= avg. differences in brain response



individual
variability



Points

- same glomerulus can respond to many odors in varying degrees
- combination of glomeruli responses yields an odor specific response
- individual variability in glomerular responses

Insect Hearing

- I. Hearing
 - A. Chordotonal ears
 - B. Tympanal ear

Insect hearing is an extension of mechanoreception

- hearing is detecting vibrations - usually air or substrate
- sensory detector attached to something that amplifies vibration - cuticle

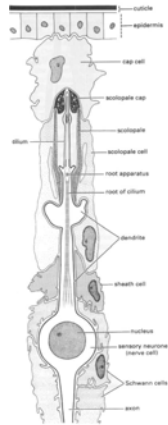
chordotonal organs

- made up of scolopidia
- one scolopidium = neuron, scolopale cell, attachment cell
- under the cuticle
- attached to cuticle on at least one end

scolopidium

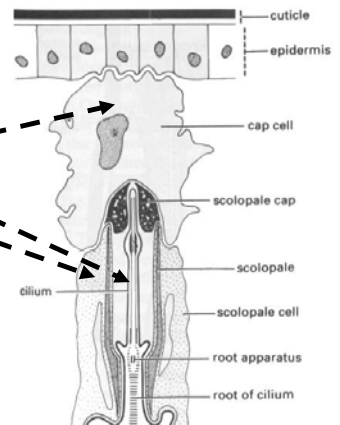
[mod.L., coined in Ger. (F. Eggers 1923, *Zool. Anzeiger* LVII. 239), f. Gr. skolop, skolof, **spike**, after OMMATIDIUM.]

- 1939 V. B. WIGGLESWORTH *Princ. Insect Physiol.* vii. 135
Chordotonal sensilla or scolopidia
These sensilla are generally believed to be derived from sensilla becoming elongated and deeply sunk within the body.



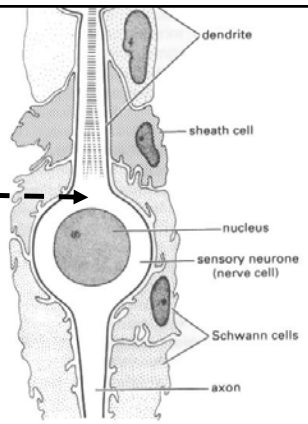
scolopidium

- attachment cell
- neuron
- scolopale cell



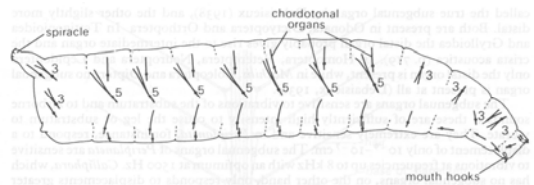
scolopidium

- neuron

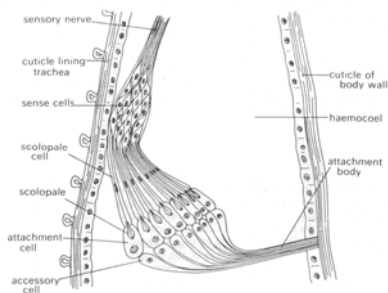


simple chordotonal organs in *Drosophila* larva

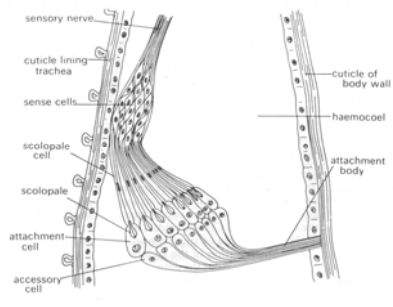
- group in clusters and give general sense of pressure, deformation of cuticle



larger groups of scolopidia placed in particular locations can detect air and substrate vibration

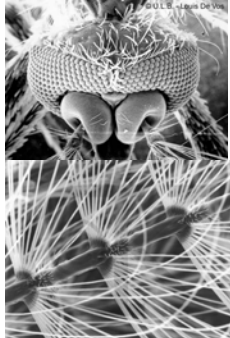


subgenual organ “below the knee”



chordotonal organ extraordinaire

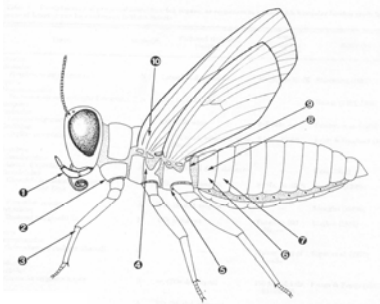
- Johnston's organ in mosquitoes
- second antennal segment
- the most complex mechanosensory organ known in insects
- what does it detect?



tympanal organs

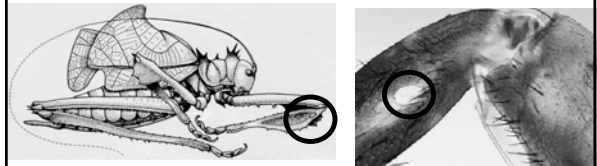
- use principle of a vibrating membrane - really thin cuticle
- usually backed by an air sac to allow free vibration
- to it are attached 1-1000 scolopidia

tympanal organs can be just about anywhere



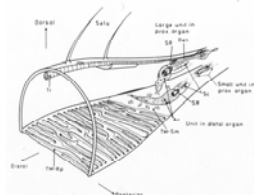
tympanal organs can be just about anywhere

- prothoracic legs - crickets and katydids

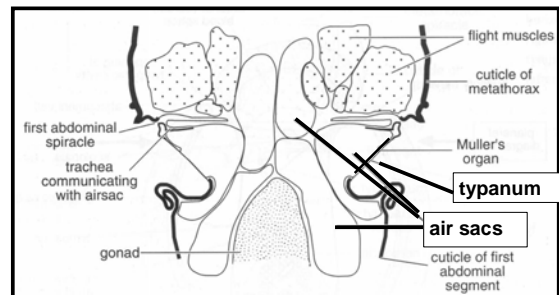


tympanal organs can be just about anywhere

- wing vein - lace wing



grasshopper ear - abdomen

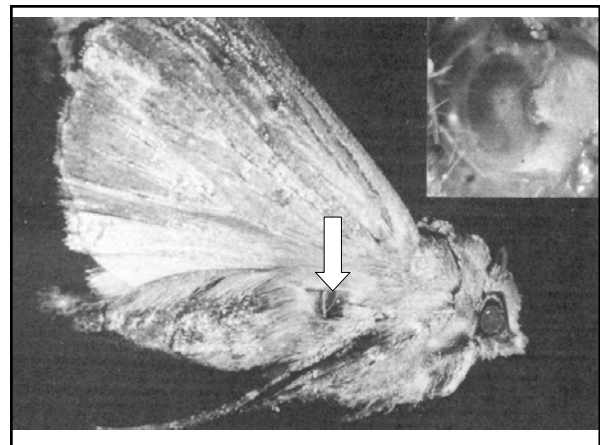


- ~80 scolopidia attached to tympanum
- different directions, attachment sites, shapes give different nerves different sensitivities
- remind you of anything?

b) Muller's organ

katydids and crickets

- acoustic spiracle stays open
- acoustic trachea
- directional hearing



- tympanum
- scolopidium

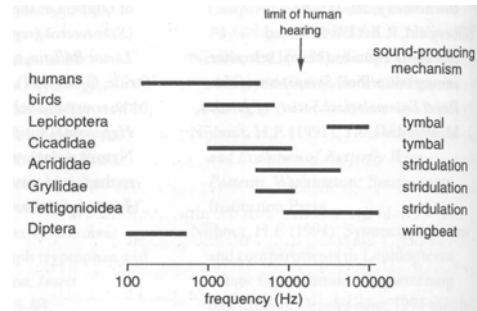
simple moth ears

moth dives

Insect Sounds

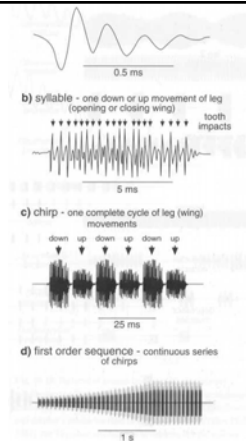
- stridulation (file and scraper)
- tymbal - vibrating membrane
- percussion (eg. striking head on substrate)
- vibrations produced by wing muscles
- air expulsion (eg. hissing cockroach)

frequencies

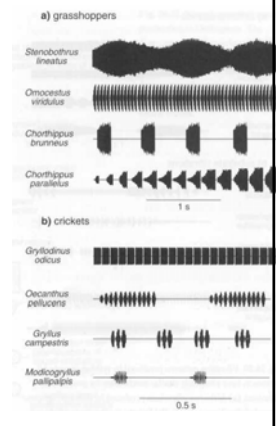


sonograms

- syllable - one stroke
- chirp - full cycle
- sequence - <100 ms between chirps

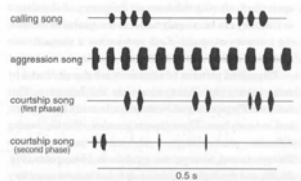


- grasshopper (leg)
and cricket (wing) songs
-different songs for
different species

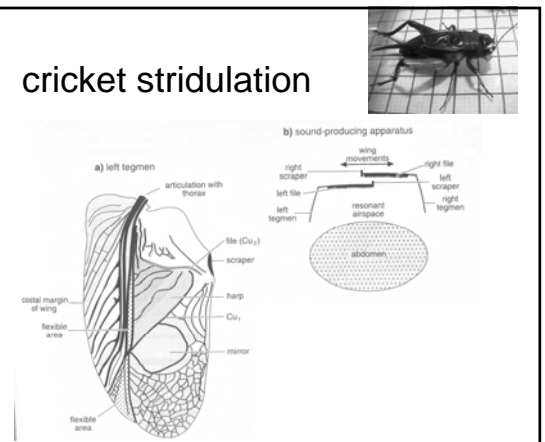


crickets

- different songs made by the same individual

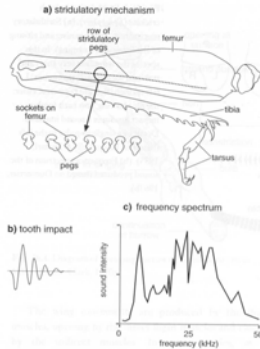


cricket stridulation

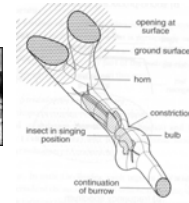


grasshopper stridulation

- pegs are pulled across wing vein



mole cricket



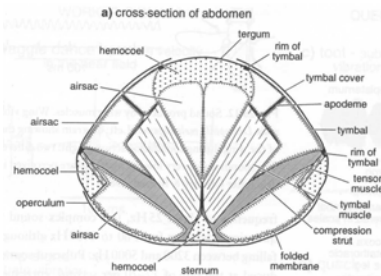
- burrow is an exponential horn
- wings of male form a diaphragm across it
- sound travels 600m

tymbal of a cicada with sclerotized ribs

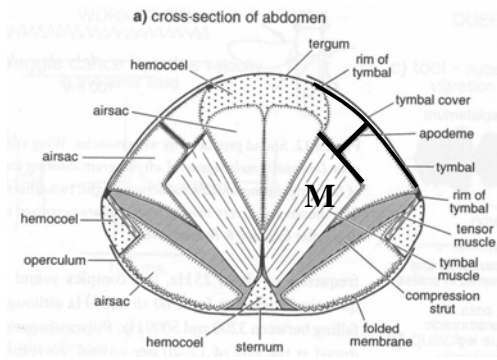


cicada

- males only
- first abdominal segment
- resilin membrane
- protected by cover



cicada



also has tympanum

- sound can radiate from both tymbal and tympanum - volume of air sacs varies intensity

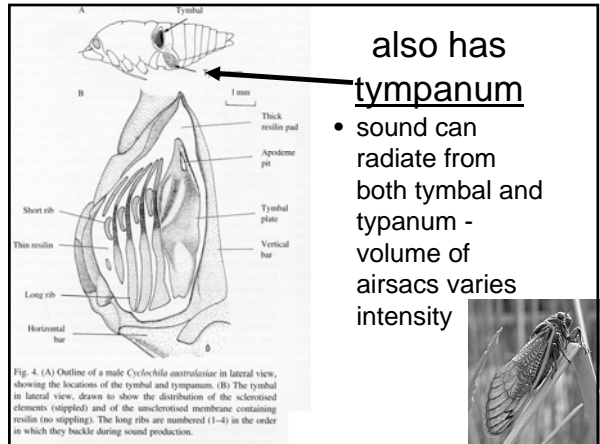


Fig. 4. (A) Outline of a male *Cicadella australis* in lateral view, showing the locations of the tymbal and tympanum. (B) The tymbal in lateral view, drawn to show the distribution of the sclerotized elements (stippled) and of the unsclerotized membranes containing resilin (no stippling). The long ribs are numbered (1-4) in the order in which they buckle during sound production.



vibration of wings made by wing muscles

- sound is at the frequency of the wing beat



Tree and leaf hoppers

- produce vibrations through their tymbal organs (they think)
- abdominal vibrations have been observed during signal production.
- Vibrations are transmitted to the substrate through the legs.

