

## Limited contemporary gene flow and high self-replenishment drives peripheral isolation in an endemic coral reef fish

Martin H. van der Meer<sup>1,2,3</sup>, John B. Horne<sup>1,2</sup>, Michael G. Gardner<sup>4,5</sup>, Jean-Paul A. Hobbs<sup>6,7</sup>, Morgan Pratchett<sup>2,3</sup> & Lynne van Herwerden<sup>1,2,8</sup>

<sup>1</sup>Molecular Ecology and Evolution Laboratory, Australian Tropical Sciences and Innovation Precinct, James Cook University, Townsville 4811, Australia

<sup>2</sup>School of Marine and Tropical Biology, James Cook University, Townsville 4811, Australia

<sup>3</sup>ARC Centre of Excellence for Coral Reef Studies, James Cook University, Townsville 4811, Australia

<sup>4</sup>School of Biological Sciences, Flinders University, GPO Box 2100, Adelaide 5001, South Australia, Australia

<sup>5</sup>Evolutionary Biology Unit, South Australian Museum, Adelaide 5000, South Australia, Australia

<sup>6</sup>The Oceans Institute and School of Plant Biology, University of Western Australia, 35 Stirling Highway, Crawley 6009, Australia

<sup>7</sup>Australian Institute of Marine Science, Perth 6009, Western Australia, Australia

<sup>8</sup>Centre for Sustainable Tropical Fisheries and Aquaculture, James Cook University, Townsville 4811, Australia

### Keywords

*Chaetodon*, coral reefs, extinction risk, Lord Howe Island, marine dispersal, Norfolk Island.

### Correspondence

Martin H. van der Meer, Molecular Ecology and Evolution Laboratory, Australian Tropical Sciences and Innovation Precinct, James Cook University, Townsville, Australia, 4811. Tel: +61-(07)-4871-5423; Fax: +61-(07)-4781-5511; E-mail: martinhvandermeer@gmail.com

### Funding Information

We are grateful for the valuable support and assistance provided by Sallyann Gudge and Ian Kerr from Lord Howe Island Marine Park. We thank the Lord Howe Island Board, Envirofund Australia (Natural Heritage Trust) and the Lord Howe Island Marine Park for financial and logistical support. We thank the Australian Department of the Environment and Water Resources for funding and the Capricorn Star for excellent logistical support. At Norfolk Island we thank Dave Biggs (Charter Marine), James Edward (Bounty Divers), Doug Creek, Michael Smith, Jack Marges, Karlene Christian, and Judith and Peter Davidson (Reserves and Forestry) for their assistance. This research was funded by the ARC Centre of Excellence for Coral Reef Studies (CE0561435).

Received: 30 October 2012; Revised: 26 March 2013; Accepted: 29 March 2013

*Ecology and Evolution* 2013; 3(6): 1653–1666

doi: 10.1002/ece3.584

### Abstract

Extensive ongoing degradation of coral reef habitats worldwide has led to declines in abundance of coral reef fishes and local extinction of some species. Those most vulnerable are ecological specialists and endemic species. Determining connectivity between locations is vital to understanding recovery and long-term persistence of these species following local extinction. This study explored population connectivity in the ecologically-specialized endemic three-striped butterflyfish (*Chaetodon tricinctus*) using mt and msatDNA (nuclear microsatellites) to distinguish evolutionary versus contemporary gene flow, estimate self-replenishment and measure genetic diversity among locations at the remote Australian offshore coral reefs of Middleton Reef (MR), Elizabeth Reef (ER), Lord Howe Island (LHI), and Norfolk Island (NI). Mt and msatDNA suggested genetic differentiation of the most peripheral location (NI) from the remaining three locations (MR, ER, LHI). Despite high levels of mtDNA gene flow, there is limited msatDNA gene flow with evidence of high levels of self-replenishment ( $\geq 76\%$ ) at all four locations. Taken together, this suggests prolonged population recovery times following population declines. The peripheral population (NI) is most vulnerable to local extinction due to its relative isolation, extreme levels of self-replenishment (95%), and low contemporary abundance.

## Introduction

Coral reef fishes have evolved in a close relationship with coral reef habitats to produce the most diverse vertebrate communities on earth (Bellwood 1996; Wood 1999; Bellwood and Wainwright 2002; Bellwood *et al.* 2010). However, coral reef habitats are coming under increasing pressure, facing a multitude of impacts including destructive and excessive fishing, sedimentation, pollution, disease, coral bleaching, ocean warming, and acidification (Hoegh-Guldberg 1999; Hughes *et al.* 2003; Bellwood *et al.* 2004). These disturbances have combined to cause sustained and ongoing declines in the abundance of corals on reefs worldwide (e.g., Gardner *et al.* 2003; Bellwood *et al.* 2004) with approximately 20% of the world's coral reefs recently destroyed and a further 50% in decline (Wilkinson 2002); whilst coral cover on the Great Barrier Reef has halved in the last 27 years (De'ath *et al.* 2012). Given their strong reliance on live coral habitats, the abundance and diversity of reef fishes invariably declines with severe and/or prolonged declines in coral cover (Jones *et al.* 2004; Graham *et al.* 2006; Wilson *et al.* 2006; Pratchett *et al.* 2008). Extensive coral loss has resulted in the local extinction of some coral reef fishes, particularly those species that rely on live coral (Kokita and Nakazono 2001; Graham *et al.* 2006; Pratchett *et al.* 2008). Local extinction of coral dependent fishes are likely to increase if major disturbances that cause acute and extensive coral loss, such as coral bleaching, increase in incidence, as predicted (Hoegh-Guldberg 1999; Sheppard 2003).

In terrestrial habitats, endemic species (particularly on isolated islands) typically have higher rates of extinction and lower genetic diversity (Frankham 1997; Whittaker and Fernández-Palacios 2007). Coral reef fish communities on isolated islands tend to have a high proportion of endemics (Jones *et al.* 2002), and account for some of the most recent fish extinctions (Dulvy *et al.* 2003). Endemic species may be particularly vulnerable to widespread disturbances with their inherent small geographical range and small population size (Gaston 1998). This risk of extinction is further increased if endemic species have specific dietary (Pratchett *et al.* 2006; Graham 2007) or specialist habitat (Munday 2004; Wilson *et al.* 2006, 2008) requirements. The ability for coral dependent fishes to recover from local extinction will be dependent on the regeneration of their coral resources and larval replenishment from distant locations as assessed by gene flow. Thus, there is an urgent need to understand gene flow between, and genetic diversity at, locations inhabited by endemic reef fishes for ongoing monitoring and conservation, and to determine their recolonization ability and resilience.

To thoroughly understand gene flow it is important that both evolutionary and contemporary levels of gene flow are determined (i.e., at various time and spatial scales; Palstra *et al.* 2007) as some reef fish studies have shown discrepancies, up to an order of magnitude difference, in gene flow over these different time/spatial scales (i.e., high evolutionary but limited contemporary gene flow: Evans *et al.* 2010; Harrison *et al.* 2012; van der Meer *et al.* 2012a). Although determining levels of gene flow is important, of equal importance is conserving genetic diversity. Conserving genetic diversity is an International Union for Conservation of Nature (IUCN) priority (McNeely *et al.* 1990) for at least two reasons: (i) it provides the raw material for natural selection to act on over evolutionary (Johannesson and Andre 2006) and contemporary time scales (Bell and Okamura 2005); and (ii) low genetic diversity increases the risk of inbreeding depression (Reed and Frankham 2003).

Large data sets of highly polymorphic msatDNA loci (nuclear microsatellites) produced by next generation sequencing (e.g., Gardner *et al.* 2011) and advancements in statistical techniques (e.g., Pritchard *et al.* 2000; Beerli and Felsenstein 2001; Wilson and Rannala 2003; Excoffier *et al.* 2005; Jombart *et al.* 2010) have increased the sophistication of population genetic studies. However, to date, few such studies have been able to sample all existing locations across a species limited range. Unsampled "ghost" locations can affect key demographic estimates (i.e., population size, genetic diversity, migration rate; Beerli 2004). Here we investigate patterns of gene flow and measure population genetic diversities in an ecological specialist reef fish, the endemic three-striped butterflyfish (*Chaetodon tricinctus*), by complete sampling across its four geographically isolated locations (all found within Australian waters): Middleton Reef – MR, Elizabeth Reef – ER, Lord Howe Island – LHI, and Norfolk Island – NI.

The three-striped butterflyfish is an endemic to the LHI region (Randall 1976). This region is a hotspot for endemic coral reef fishes (Marine Parks Authority 2010) ranking fifth in the Indo-Pacific for percent endemism (7.2%, Randall 1998). Marine Protected Areas (MPAs) have been established to conserve reef fishes at three of these locations (MR, ER, LHI), but no protection exists at NI. This is an ideal study system as reef fishes occur on only four discrete islands/reefs that are separated by deep ocean water. Thus, connectivity of reef fish populations across the four locations is restricted to oceanic dispersal of pelagic larvae over known distances (e.g., 45–600 km).

Previous research on another endemic species in this system, the McCulloch's anemonefish (*Amphiprion mccullochi*), revealed limited contemporary gene flow between ER, MR, and LHI (van der Meer *et al.* 2012a,b).

However, anemonefish have the shortest pelagic larval duration (PLD) of reef fishes (11–17 days: Victor 1986; Thresher *et al.* 1989; Wellington and Victor 1989; Victor and Wellington 2000) and their self-recruitment to natal areas has been well documented (Jones *et al.* 2005; Planes *et al.* 2009). While McCulloch's anemonefish provide a test of population connectivity in reef fishes at the lower limit of dispersal potential within the LHI region, determining the connectivity of reef fishes in general requires examining species from a common group with PLD's more typical of reef fish (20–50 days). Butterflyfishes (Chaetodontidae) are one of the 10 common families of fishes that are characteristic of coral reefs (Bellwood and Wainwright 2002). The PLD of *C. tricinctus* (mean = 35 days; M. van der Meer & J.-P. A. Hobbs, unpublished data), is typical of butterflyfishes (26–53; e.g., Brothers *et al.* 1983; Brothers and Thresher 1985) and many other reef fishes. *C. tricinctus* is also one of the 41 butterflyfish species that feed directly on scleractinian corals (Cole *et al.* 2008; Rotjan and Lewis 2008). Thus, *C. tricinctus* provides a test of population connectivity in a common group of reef fishes, that is closely associated with coral reefs, and with a dispersal potential typical of most reef fishes.

*Chaetodon tricinctus* faces a higher risk of extinction as a consequence of its small geographic range, compared to its closest relatives *C. bennetti*, *C. plebeius*, and *C. trifascialis* (Bellwood *et al.* 2010), which are distributed widely throughout the Indo-Pacific (Allen *et al.* 1998). Moreover, *C. tricinctus* feeds exclusively on live corals (Kuitert 1996) and is mostly found in close association with corals of the genus *Acropora* (Hobbs *et al.* 2009). The abundance of *C. tricinctus* is positively linked to the abundance of *Acropora* spp., indicating that a loss of this coral could cause decreases in abundance and potential local extinction of *C. tricinctus* (Hobbs *et al.* 2009). The global abundance of *C. tricinctus* is likely to be much smaller than its widespread congeners, and if it cannot alter its diet following coral loss, then these factors will compound upon its small geographic range and greatly increasing its vulnerability to local and possibly global extinction. Dramatic declines in abundance of several other butterflyfishes have occurred following extensive coral loss (Syms 1998; Pratchett *et al.* 2006), but some of the most vulnerable species have been spared from extinction due to their large geographic range (Lawton *et al.* 2011). Given that *C. tricinctus* exists at a few isolated locations and may be particularly vulnerable to local extinction, there is an obvious need to determine patterns of population connectivity and replenishment for this species.

The aims of this study were threefold: (i) to determine patterns and levels of gene flow between locations using mtDNA (mitochondrial DNA) and msatDNA; (ii) to esti-

mate levels of self-replenishment (as a proxy for realized self-recruitment) and recent migration; and (iii) to measure population genetic diversities at all locations as an indicator of potential resilience of populations to environmental change and extinction.

## Materials and Methods

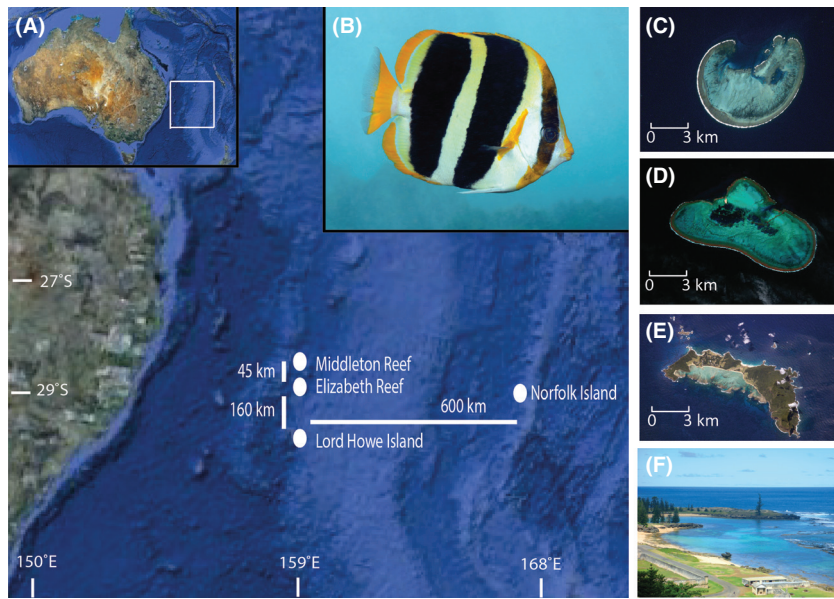
We applied a range of frequency and Bayesian based molecular tools to establish mtDNA and msatDNA levels of phylogenetic and population genetic structure. This resulted in a comprehensive understanding of gene flow in this study system and together these tools provided a comprehensive view of dispersal (Leis *et al.* 2011). However, due to the large number of analyses, we present only methods related to this study below, whilst general Materials and Methods (i.e., genetic and laboratory techniques and, in-depth mt and msatDNA analyses) are presented in van der Meer *et al.* (2012a,b,c). Fin clip sample sizes ranged from 21 to 31: MR ( $n = 30$ ), ER ( $n = 31$ ), LHI ( $n = 26$ ), and NI ( $n = 21$ ). We used a large number of polymorphic microsatellite loci ( $n = 20$ ) and sampled all known locations, to compensate for the small sample sizes used in this study (see Selkoe and Toonen 2006). Furthermore, we recognize that our estimates for “self-replenishment” inferred indirectly from genetic markers are merely a proxy for self-recruitment, which is typically assessed using more direct methods (e.g., natural or artificial otolith tags), such as those used by Swearer *et al.* (1999), Jones *et al.* (2005), and Almany *et al.* (2007). Nevertheless, direct approaches are not feasible for our study species, without negatively impacting populations, due to the large sample sizes typically required for such parentage-based studies. Therefore, we believe that our indirect estimates of self-replenishment represent the best possible substitute for realized self-recruitment obtainable for this species.

## Ethics statement

The main aim of this study was to determine gene flow between and genetic diversity at isolated locations using the endemic three-striped butterflyfish (*C. tricinctus*) as a model organism. Fin clip samples were obtained from fishes of adult size (>100 mm total length) either by spearfishing or by anesthetizing fish with clove oil, which were fin clipped in situ and released alive (Permit Numbers: LHIMP08/R01, 003-RRRWN-110,211-02, P11/0035-1.0, LHIMP/R/2010/012; Animal ethics: A1605).

## Study system

Throughout this study the three locations MR, ER, and LHI are collectively referred to as the “western region”



**Figure 1.** Location maps and focal species. (A) Google Earth image of eastern Australia showing Middleton Reef (MR), Elizabeth Reef (ER), Lord Howe Island (LHI), and Norfolk Island (NI) in the South West Pacific Ocean. (B) *Chaetodon tricinctus* swimming in the open (Photo courtesy of Justin Gilligan). Aerial photographs of MR (C); ER (D), LHI (E), and NI (F; the bay measures 1 km in length)

because they occur on the same geographic feature (Lord Howe Island Rise – remnants of volcanoes estimated to be 6.7 Ma, McDougall *et al.* 1981), are relatively close to each other (Fig. 1) and all locations support high abundances of *C. tricinctus* (Choat *et al.* 2006; Hobbs *et al.* 2010). In contrast, NI is referred to as the “peripheral location” for *C. tricinctus*, because it is the only location situated on a separate geographic feature (Norfolk Island Rise – remnant of a volcano estimated to be 2.3–3.05 Ma, Jones and McDougall 1973), it is isolated by more than 600 km from the western region (Fig. 1) and has relatively low abundance (a total population size estimated to be less than 30 individuals – authors unpublished data).

## Gene flow between locations – mtDNA

### mtDNA phylogenetic analysis

mtDNA Cytochrome b (cyt b) sequence data were obtained from GenBank for the following three most closely related species which acted as out-groups: *C. trifascialis* (FJ167707.1), *C. plebius* (AF108602.1), and *C. bennetti* (FJ167686.1) based on the findings of Bellwood *et al.* (2010). jModeltest (Posada 2008) identified an TrN + G model under Akaike Information Criterion with gamma = 0.759. The three most commonly used phylogenetic analyses: Maximum Likelihood (ML), Maximum Parsimony (MP), and Bayesian Inference (Mr Bayes – MB and BEAST) were performed on the aligned mtDNA sequence data as described in van der Meer *et al.* (2012a,b). This was done to identify any underlying evolutionary partitions in the data, based on the use of rigorous analytical tools. A Minimum Spanning Tree (MST) was generated

based on output obtained from ARLEQUIN 3.5 (Excoffier *et al.* 2005) to explicitly identify shared haplotypes between *C. tricinctus* from the four locations.

### Quantifying the level of mtDNA gene flow

mtDNA migration rates and effective population sizes of *C. tricinctus* were estimated between or within each of the four locations using MIGRATE-n 2.4.3 (<http://popgen.sc.fsu.edu/Migrate-n.html>; Beerli and Felsenstein 2001; Beerli 2004). We tested a combination of various migration priors ( $F_{st}$  and own: isolation-by-distance), custom-migration models (Stepping-stone, Island-n, and variable Theta only) and a geographic matrix – all with a constant mutation rate. A Log Maximum-Likelihood analysis (Ln ML) selected a migration prior ( $F_{st}$ ), custom-migration model (migration model with variable Theta), constant mutation rate with an F84 mutation model, migration rate parameters (Theta and M to a maximum of 1 and 15,000, respectively), and a Bayesian analysis, using a heating search strategy of one long chain that sampled every 20th of 60 k sampled trees and applied a 20 k iteration burn-in. All parameters converged and fell within the 90% CI yielding values for  $\theta$  and M (mutation-scaled migration rate) per location.

## Gene flow between locations – msatDNA

### Patterns of gene flow (msatDNA)

Three molecular analytical tools were used to establish spatial population partitioning in msatDNA: (i) Discriminant Analysis of Principal Components (DAPC; Jombart



*et al.* 2010) uses allelic states to discriminate between the four locations, yielding scatterplots of discriminant functions based on the spatial distributions of microsatellite genotypes. DAPC also provided posterior probabilities of population assignments for each individual; (ii) a likelihood-based assignment method was used in GeneClass2 (Paetkau *et al.* 1995, 2004; Piry *et al.* 2004) to determine significant interlocation gene flow and (iii) STRUCTURE V2.3 (Pritchard *et al.* 2000; Hubisz *et al.* 2009) placed individuals into clusters that minimize Hardy–Weinberg Equilibrium (HWE) and can be used to identify contemporary gene flow between the four locations. To determine the “best value” for  $K$ , we followed the method suggested by Pritchard *et al.* (2000), which involved comparing mean log-likelihoods penalized by one-half of their variances (see Hubisz *et al.* 2009). The final run consisted of an Admixture model with 2 M iterations and a 100 k iteration burn-in.

### Quantifying the level of msatDNA gene flow

Contemporary migration rates and effective population sizes of *C. trichinotus* were tested and estimated between each of the four locations) using MIGRATE-n 2.4.3 as above. We set datatype to Microsatellite (a simple electrophoretic ladder model), migration prior ( $F_{st}$ ), custom-migration model (migration model with variable Theta), constant mutation rate with a stepwise mutation, migration rate parameters (Theta and M to a maximum of 10 and 20, respectively), and a Bayesian analysis, using a heating search strategy of one long chain that sampled every 20th of 60 k sampled trees and applied a 20 k iteration burn-in. All parameters converged and fell within the 90% CI yielding values for  $\theta$  and M (mutation-scaled migration rate) per location.

### Inferred levels of self-replenishment and recent migration

This study did not sample new butterflyfish recruits in order to determine self-recruitment as in Jones *et al.* (2005). However, we used BAYESASS v3 (Wilson and Rannala 2003), a program specifically designed for population genetic studies that estimates recent migration rates (past 2–3 generations) between populations (or locations). Conversely, this program also has the ability to estimate any individuals not migrating (i.e., self-replenishing). We used BAYESASS v3 to estimate both self-replenishment (as a proxy for realized self-recruitment) and recent migration between locations; with a Markov chain Monte Carlo (MCMC) chain, consisting of a total of 11 M steps with a 2 M step burn-in; prior values for migration rate, allele frequency, and inbreeding coefficient were specified as 0.5,

0.6, and 0.6, respectively. These priors were selected because they gave acceptance rates within the 20–40% range showing convergence of the MCMC (Faubet *et al.* 2007). Ten independent runs separately assessed convergence of the MCMC (i.e., priors fell within the 20–40% range suggesting convergence) in order to evaluate the consistency of results obtained from these inferences.

### Population genetic diversities

Molecular diversity indices for mtDNA (haplotype diversity,  $h$ ; nucleotide diversity,  $\pi$ ) and for msatDNA (genetic diversity,  $gd$ ) were estimated in ARLEQUIN 3.5 (Excoffier *et al.* 2005). Haplotype and nucleotide diversities of the data were interpreted as either low with specified cut-off values of  $h$  and  $\pi$  (%) <0.5 or high if values of  $h$  and  $\pi$  (%) were >0.5 (Grant and Bowen 1998).

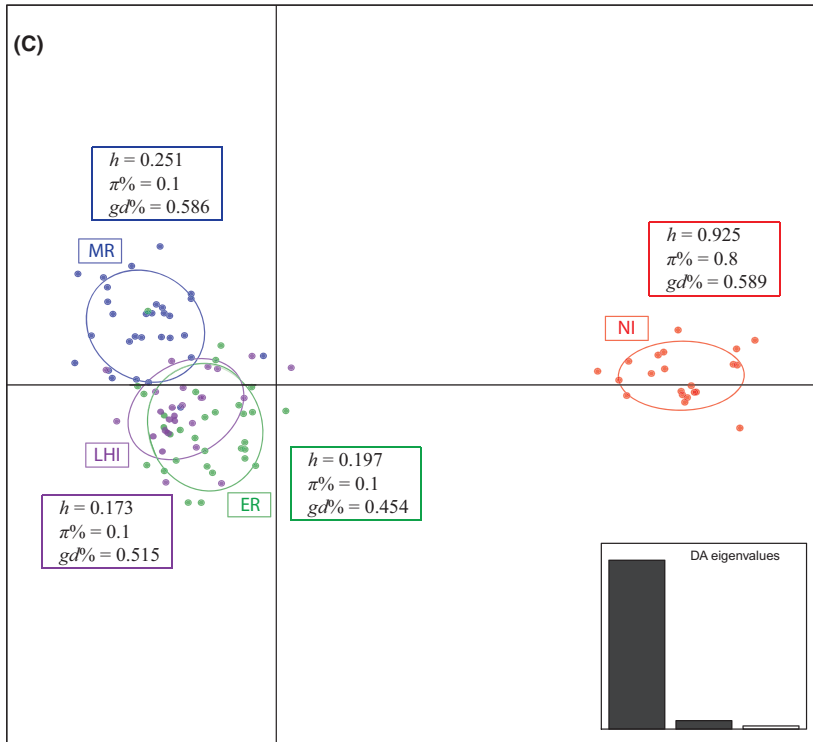
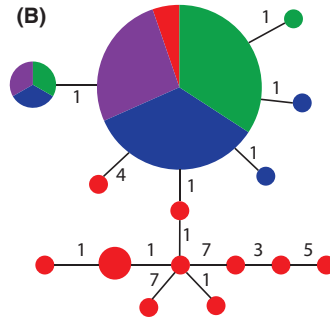
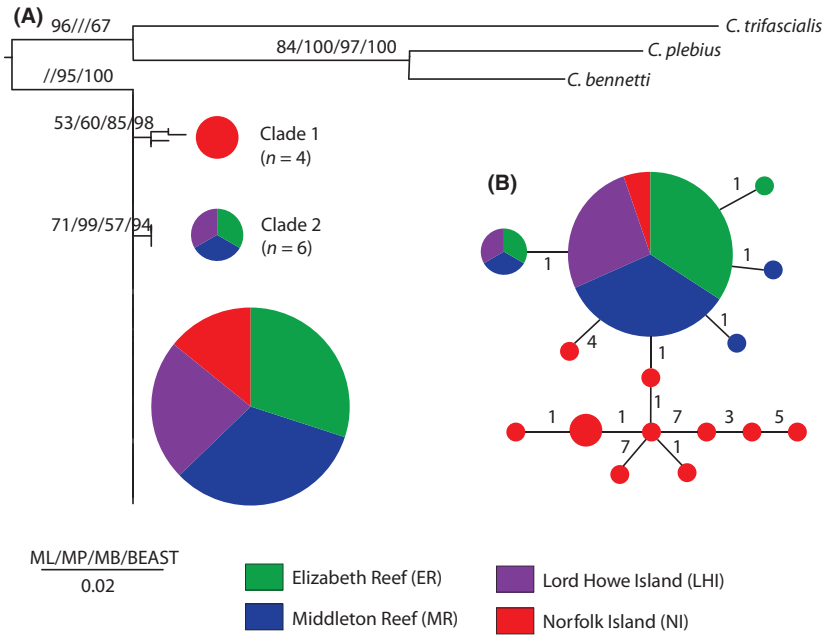
## Results

### Synopsis

Two hundred and eighty-three base pairs of mtDNA (cyt b) were resolved for 97 *C. trichinotus* individuals; with a total of 15 polymorphic sites, of which three were parsimony informative. One small clade, Clade 1 ( $n = 4$ ) contained exclusively individuals from the peripheral location. The other, Clade 2 ( $n = 6$ ), comprised of an equal frequency of all three western locations (i.e., MR, ER, LHI; Fig. 2A). A MST identified 15 haplotypes in total, one of which was observed at high frequencies representing 82% ( $n = 80$ ) of all individuals, and 12 of which were unique to single fish only in the sample examined here, nine of which were from the peripheral location (Fig. 2B). Mt and msatDNA Analysis of Molecular Variance (AMOVA) and pairwise  $F_{st}$  results indicate that there is population genetic differentiation between the western locations and the peripheral location, but there is no population genetic differentiation within the western region (i.e., when MR, ER, LHI are grouped). Haplotype and genotype diversities were low (<0.5) within the western region, but high (>0.5) at the peripheral location (Figs. 3 and S1). Genotypic diversity ( $gd$ ), in contrast, was high at three of the four locations, ER being the exception. Detailed genetic diversity, AMOVA, summary statistics, pairwise population comparisons, and locus by locus AMOVA can be found in Supporting Information (S1, S2, S3, S4, and S5, respectively).

### Summary statistics

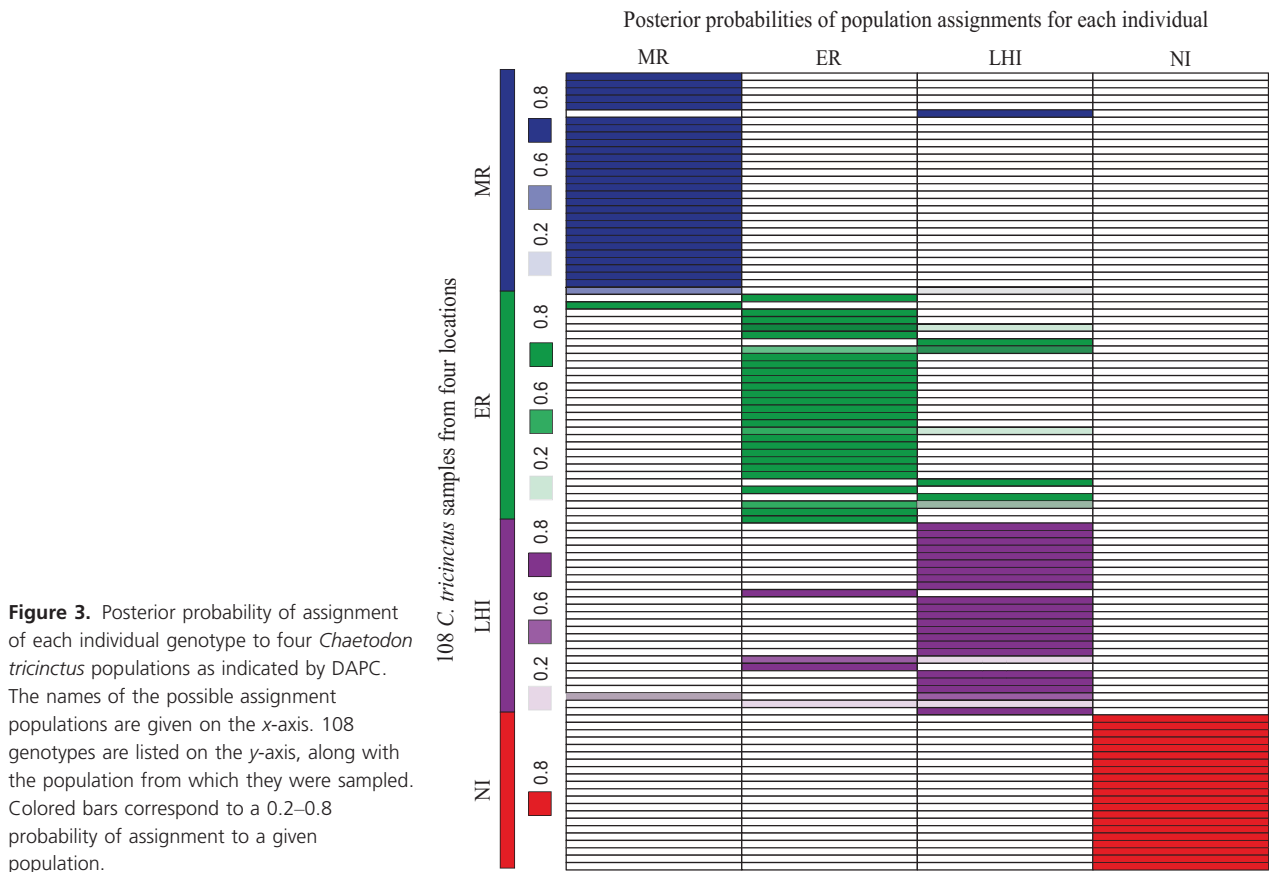
Heterozygote excess was evident from a negative inbreeding coefficient ( $F_{IS}$ ; Table S1); although this was not



**Figure 2.** mt and msatDNA *Chaetodon trilineatus* analyses. (A) A phylogram of mtDNA (cyt b) sequences from 97 *C. trilineatus* individuals from Middleton Reef, Elizabeth Reef, Lord Howe Island, and Norfolk Island. This represents the best ML tree from 10 individual analyses. Numbers on branches indicate support for each clade, based on ML, MP, MB, and BEAST analyses. (B) Haplotype minimum spanning tree (MST) with number of substitutions between haplotypes indicated on connectors. Different fills represent each of the four locations as shown on the key to the figure, and (C) Scatterplots of the discriminant analysis of principal components of the microsatellite data for four *C. trilineatus* locations using geographic sample site as priors for genetic clusters. Individual genotypes appear as dots surrounded by 95% inertia ellipses. Eigenvalues show the amount of genetic information contained in each successive principal component with X and Y axes constituting the first two principle components, respectively. Boxes indicate haplotype (*h*), nucleotide (%  $\pi$ ), and genetic diversity (*gd*) indices for *C. trilineatus*.

significant between the four locations. The peripheral location had the most private alleles, 25 across twenty loci, while the remaining three locations ranged from 6 to 15 private alleles across all loci (Table S2). Of the 20 msatDNA loci: (i) significant single locus departures from HWE were detected in 11 of 80 tests at the location level before False Discovery Rate (FDR) correction and nine

afterwards (ER: Ct4, Ct23, Ct24; MR: Ct16; NI: Ct3, Ct13, Ct17, Ct23, Ct24; Table S2), similarly, six single locus HWE departures were detected before and after FDR when all locations were considered (Table S2); (ii) null alleles were identified in MR (Ct18) and NI (Ct10, Ct16); and (iii) of the 212 locus by locus exact tests of linkage disequilibrium, 13 were significant before and 10



after FDR correction (Benjamini and Hochberg 1995). Loci that were not in HWE in more than one location (Ct23, Ct24) and had null alleles (Ct10, Ct16, Ct18) were not used in subsequent analysis (ARLEQUIN, STRUCTURE, and MIGRATE-n) and, loci in linkage disequilibrium at all sites (Ct17, Ct21, Ct23) were not used in subsequent analysis (ARLEQUIN, STRUCTURE, BAYESASS). Thus 13 loci were used in the ARLEQUIN and STRUCTURE analyses, 15 loci were used in the MIGRATE -n analysis and 17 loci in the BAYESASS analysis.

### Gene flow between locations – mtDNA

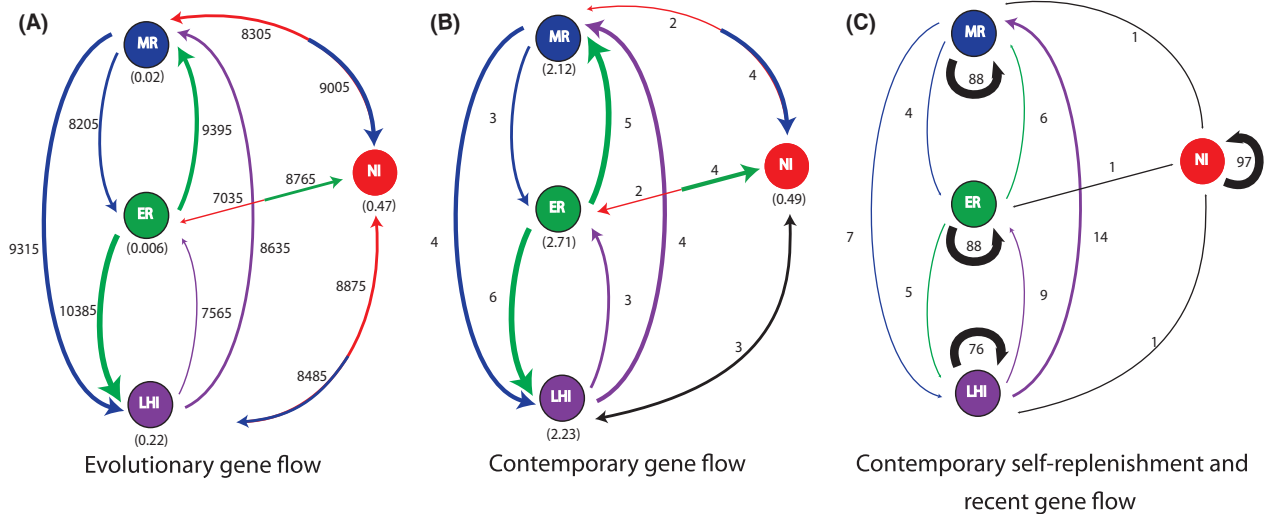
*Patterns and levels of gene flow* based on an mtDNA AMOVA indicated significant genetic variation (65.77%) within locations,  $\Phi_{st} = 0.342$  ( $P < 0.001$ , Table S3). This was due to the peripheral location mtDNA pairwise  $F_{st}$  differentiation from all three western locations (pairwise  $F_{st} = 0.190$  to  $0.221$ ,  $P < 0.001$ ; Table S4). Whilst there was no genetic differentiation among the three western locations (pairwise  $F_{st} = -0.032$  to  $-0.023$ ,  $P = 0.865$  to  $0.991$ ; Table S4). A single regional partition was also suggested between the western region and the peripheral

location, explaining 36.45% of the genetic variation, but this was not significant ( $\Phi_{ct} = 0.365$ ,  $P = 0.250$ ; Table S3).

*Quantifying mtDNA gene flow* using Migrate-n indicated high levels of mtDNA gene flow between all locations, with M values ranging from 7035 to 10,385 (Fig. 4A).

### Gene flow between locations – msatDNA

*Patterns and levels of gene flow* based on an msatDNA AMOVA indicated significant structure in five (of 20) locus by locus analyses corrected for null allele frequency ( $\Phi_{st} = 0.001$  to  $0.368$ ,  $P < 0.05$ ; Table S5), in six (of 20) locus by locus analyses corrected for standardized location differentiation ( $\Phi_{st} = 0.004$  to  $0.852$ ,  $P < 0.05$ ; Table S5) and in the global AMOVA as a weighted average over all microsatellite loci ( $\Phi_{st} = 0.046$ ,  $P < 0.001$ ; Table S3), with 95.39% of the genetic variation existing within locations. Raw msatDNA pairwise  $F_{st}$  comparisons showed very low to moderately significant genetic partitioning between the western locations and the peripheral location ( $F_{st} = 0.056$  to  $0.101$ ,  $P < 0.001$ ). In contrast, an Excluding Null Alleles (ENA) corrected msatDNA pairwise  $F_{st}$  value showed no significant genetic differentiation between any of the four locations as estimates of genetic differentiation



**Figure 4.** Migration rates among *Chaetodon trilineatus* locations. The thickness of the line is directionally proportional to the number of migrants (M) and the line colors indicate the predominant direction of gene flow. Population size ( $\theta$ , within parentheses) is also shown for each location. (A) Migrate-n evolutionary gene flow (mtDNA), (B) Migrate-n contemporary gene flow (msatDNA), and (C) BAYESASS analysis of self-replenishment and recent migration rates (msatDNA) shown as a percentage.

between locations fell within 95% confidence intervals ( $F_{st} = 0.005$  to  $0.084$ ,  $P > 0.05$ ; Table S4).

DAPC, GeneClass2, and STRUCTURE confirmed the presence of at least three distinct genetic populations corresponding to geographic location. DAPC partitioned *C. trilineatus* into the western region and the peripheral location (Fig. 2C). Using the four locations as *a priori* population criteria, DAPC assigned 58–100% of all individuals to the location from which they were sampled (assignment per population, ER = 74%, MR = 90%, LHI = 58%, NI = 100%; Fig. 3). Consistent with these assignments, with the allele frequencies and genotypic assignments, the 95% Genotypic Inertia Ellipses (GIE) for ER and LHI overlap, whilst the 95% GIE for MR does not overlap with either ER or LHI and the 95% GIE for NI occupy a distant area of multivariate space, along the  $x$ -axis, from all three western locations. Geographical structure in msatDNA data was confirmed by GeneClass2 analyses, where only 11 individuals were grouped with a location from which they were not sampled (MR = 1, ER = 7, LHI = 3); thereby identifying four genetically differentiated populations. Similarly, four geographically partitioned populations were identified by STRUCTURE analyses, as the likelihood of the marginal posterior probability distribution was highest when  $K = 4$ .

#### Quantifying the level of msatDNA gene flow

Migrate-n indicated a few orders of magnitude lower levels of contemporary gene flow between locations when

compared to mtDNA gene flow, with M values ranging from 2 to 6 (Fig. 4B).

#### Inferred levels of self-replenishment and migrant exchange

Despite weak genetic differentiation ( $F_{st}$ ) between locations within the western region, both DAPC and STRUCTURE partitioned the data into at least three distinct clusters. Used together, these programs are likely to be better than  $F_{st}$  values (Faubet *et al.* 2007) at determining the appropriateness of a dataset for BAYESASS because they extract more information from the genetic data than frequency-based fixation indices. However, it is important to note: (i) BAYESASS estimates of migration rates are accurate when migration rates are low, genetic differentiation between locations is not too low ( $F_{st} \geq 0.05$ ) and loci are in linkage equilibrium. Moreover, if the above-mentioned conditions are not met, then accurate estimates of migration rates are obtained only when migration rates are very low ( $m = 0.01$ ) and genetic differentiation between locations is high ( $F_{st} \geq 0.10$ ; Faubet *et al.* 2007) and (ii) when estimates of migration rates fall below 10%, populations can probably be considered demographically independent from each other with no gene flow between locations (Waples and Gaggiotti 2006). Demographic independence is suggested for all location pairs except: LHI to MR ( $m = 14\%$ ) and possibly LHI to ER ( $m = 9\%$ ; Fig. 4C). Conversely, high levels of self-replenishment (76–96%) were inferred at all four locations (Fig. 4C). This further indicates that in the



short term, each population is predominantly sustained by self-replenishment rather than replenishment from distant populations.

### Population genetic diversities

*Chaetodon tricinctus* showed low haplotype diversity ( $h$ ) and nucleotide diversity ( $\% \pi$ ) in all three western locations ( $h = 0.173$  to  $0.251$ ,  $\% \pi = 0.1$ ), whilst the peripheral location had four- to eightfold higher  $h$  and  $\% \pi$  ( $h = 0.925$ ,  $\% \pi = 0.8$ ; Fig. 2C), respectively. Three of the four locations (MR, LHI, NI) had high genetic diversity ( $gd = 0.515$ – $0.589$ ), ER being the exception ( $gd = 0.454$ ; Fig. 2C). Total haplotype, nucleotide, and genotypic diversities were low ( $h = 0.384$ ,  $\% \pi = 0.2$ ,  $gd = 0.490$ ; Table S1).

### Discussion

Understanding both time and spatial scales of gene flow and the levels of genetic diversity is vital to determine best practice management, maximize biodiversity conservation, and evaluate the capacity of coral reef fishes to recover should they become locally extinct. In this study, *C. tricinctus* was found to have (i) sufficient mtDNA gene flow connecting all locations within the western region, but low gene flow and consequent isolation of the peripheral population from the western locations; (ii) low msatDNA gene flow between all locations resulting in populations that are genetically differentiated; (iii) demographic dependence between LHI and MR (and possibly LHI and ER), yet high levels of inferred self-replenishment at all four locations; (iv) variable genetic diversities: low mtDNA genetic diversity at all three locations within the western region, but not at the peripheral location; and (v) high msatDNA genetic diversity at all four locations.

### Gene flow between locations – mtDNA

Monophyly was suggested for *C. tricinctus* with the exception of two clades, one of which consisted exclusively of the peripheral location. The lack of geographic population structure within the western region may result if a small number of recruits per generation maintain spatial genetic homogeneity (Shulman 1998; Planes 2002). Similar genetic homogeneity has been found in studies on the endemic Hawaiian butterflyfishes *Chaetodon multicinctus*, *C. miliaris*, and *C. fremblii* (Craig *et al.* 2010) and in numerous other coral reef fish species including the parrotfishes *S. frenatus* and *C. sordidus* (Dudgeon *et al.* 2000; Bay *et al.* 2004). In contrast, the genetic differentiation between the western region and the peripheral location likely results from limited gene flow due to

geographic isolation (600 km of deep ocean separating each region) and complicated ocean currents (which are seasonally stronger or weaker flowing west to east along the East Australian Current and seasonally migrating north or south along the Tasman Front; Suthers *et al.* 2011). Such strong genetic breaks at peripheral locations has been demonstrated in other reef fishes, including two widespread coral reef snappers *Lutjanus kasmira* and *L. fulvus* (Gaither *et al.* 2010) and two widespread parrotfishes *Scarus psittacus* (Winters *et al.* 2010) and *Chlorurus sordidus* (Bay *et al.* 2004).

Despite high mtDNA gene flow between all locations, conventional statistics (AMOVA and pairwise  $F_{st}$ ) indicate that the three locations within the western region and the peripheral location are genetically differentiated. Although high mtDNA gene flow may provide some assistance to distant populations through recolonization following local extinctions and increasing genetic diversity (Hanski 1999; Jones *et al.* 2009), benefits to the maintenance of distant populations may be minimal, especially if combined with little or no contemporary gene flow on demographic time scales.

### Gene flow between locations – msatDNA

*Chaetodon tricinctus* showed contemporary genetic differentiation between all locations (with the possible exception of ER and LHI). The strong discrepancy between mtDNA and contemporary levels of gene flow in *C. tricinctus* is increasingly being documented in other coral reef fishes such as snappers *Lutjanus carponotatus* (Evans *et al.* 2010; Harrison *et al.* 2012) and *Lutjanus synagris* (Gold *et al.* 2011), coral trout *Plectropomus maculatus* (Evans *et al.* 2010; Harrison *et al.* 2012), and in the endemic LHI anemonefish *A. mccullochi* (van der Meer *et al.* 2012a). This “lack of congruence” between time scales may result from genetic homogeneity over evolutionary time scales (Shulman 1998; Planes 2002) compared to substantial amounts of self-recruitment over contemporary time scales (Swearer *et al.* 1999; Jones *et al.* 2005; Almany *et al.* 2007; Planes *et al.* 2009).

The estimation of contemporary gene flow is important for conservation because models predict that a few recruits per generation over evolutionary time scales will not sustain populations (Cowen *et al.* 2000, 2002). In light of this, MPAs are designed to be large enough for locations to sustain themselves and yet spaced close enough so that larvae produced within an MPA can potentially be exported to unprotected areas (see Halpern and Warner 2003; Shanks *et al.* 2003; Jones *et al.* 2005; Harrison *et al.* 2012). In the case of *C. tricinctus*, it is unlikely that the current MPAs in the western region will deliver any substantial recruitment to the peripheral

location due to the geographic isolation and complicated ocean currents around the LHI and Norfolk Island Rise regions, and the high levels of larval retention possibly facilitated by natal homing (Botsford 2005; Hilborn *et al.* 2006). The high abundance of *C. tricinatus* at the western locations (Choat *et al.* 2006; Hobbs *et al.* 2009) reduces the likelihood of local extinction, while higher levels of contemporary gene flow, when compared to the peripheral location, are likely to facilitate recovery following population declines (or local extinction). Given the extremely small population size of *C. tricinatus* at NI (estimated to be less than 30 individuals), a slow recovery time is expected following population declines, due to intermittent pulse replenishment.

Less than 10% gene flow between populations suggests demographic independence (Waples and Gaggiotti 2006) and high levels of self-recruitment, which is vital for populations to persist (Hastings and Botsford 2006). However, trying to classify populations as “open” or “closed” may not be appropriate (Mora and Sale 2002; Largier 2003). Rather, populations that have 80% of the successful recruits generated internally, will take substantially longer to recover following local extinction than ones with only 20% self-recruitment (Miller and Shanks 2004) and should be considered *largely closed* or *largely open*, respectively. All locations appear demographically independent and may be considered largely closed. However, both MR and to a lesser extent ER, receives some gene flow from LHI, suggesting that the population at LHI is important for management and continued protection because it exports individuals to MR and ER. Levels of inferred self-replenishment found in *C. tricinatus* ( $\geq 76\%$ ) are highly similar to the estimated levels of self-recruitment in other congeneric butterflyfish in Papua New Guinea (PNG; Almany *et al.* 2007) and other island populations of coral reef fishes (Swearer *et al.* 1999; Jones *et al.* 2005; Planes *et al.* 2009). The consistency of results between the indirect methods of the present study and the direct methods of former studies suggest that self-replenishment can be used to approximate self-recruitment in coral reef fish populations, given a sufficient number of unlinked loci, high detectable levels of self-replenishment and no unsampled ghost locations. Moreover, estimates of self-replenishment in *C. tricinatus* tended to be slightly higher than estimates of self-recruitment in the above studies. It is unlikely that this difference is due entirely to methodological considerations, given that indirect genetic methods are thought to overestimate gene flow (Hellberg *et al.* 2002). Rather, high levels of self-recruitment in *C. tricinatus* might be inherent of its small geographic range and thus high self-recruitment is needed to sustain isolated populations in this system (Hobbs *et al.* 2011). Alternatively, studies sampling new

recruits are also estimating self-recruitment during the postsettlement mortality period. Consequently, the genetic makeup of the recruit cohort that survives through to adulthood is changed creating a disparity between estimates of self-replenishment and self-recruitment.

### Population genetic diversities

*Chaetodon tricinatus* showed high mtDNA genetic diversity at the peripheral location and low diversity at the western locations. While msatDNA genetic diversity was high at three of the four locations (MR, LHI, NI), ER being the exception. Similar genetic diversities have been found in other coral reef fish using cytochrome b including the endemic Hawaiian butterflyfishes *Chaetodon fremblii*, *C. miliaris*, and *C. multicoloratus* (Craig *et al.* 2010) and in the more widespread butterflyfishes *C. lunulatus*, *C. trifascialis*, and *C. trifasciatus* (Lawton *et al.* 2011; Montanari *et al.* 2011). Species with high genetic diversity may have some resilience to extinction as a decrease in genetic diversity is generally associated with decreased fitness (Hoelzel *et al.* 2002). Thus, the high overall genetic diversity at the peripheral location, resulting from pulse recruitment periodically bringing new genetic material into the population resident here and the occurrence of rare haplotypes (see below), is encouraging, as it may buffer a small, demographically isolated population against some impacts. However, the reverse patterns occur at the western locations, where the risk of extinction associated with low genetic diversity is counteracted by high population abundances. Of interest are the rare haplotypes seen only at the peripheral location that may represent either historical polymorphisms (a relic or refugium population) or mutation accumulation. Given the high abundance of *Acropora* at this location (authors unpublished data) but extremely low abundance of *C. tricinatus* individuals, it is likely that self-recruitment is limiting population numbers. If unique genetic diversity is a feature of NI populations of endemics within the region, then protecting these populations and the habitats they rely on is vitally important.

### Conclusion

Given the low contemporary gene flow between western and peripheral locations (and high self-replenishment) in *C. tricinatus*, the MPAs at the western region are of limited benefit to the unprotected peripheral location (NI). Therefore, the peripheral location requires some protective management strategies to conserve its genetically unique population of *C. tricinatus*. However, within the western region, LHI is an important source of gene flow to both MR and ER and as such, warrants continued MPA protection and monitoring. Similar patterns of gene

flow between locations has also been found for the endemic McCulloch's anemonefish, *A. mccullochi* (van der Meer *et al.* 2012a) and may be indicative of generalized patterns of gene flow of all endemics in the region. Given the importance of the LHI region as an endemic hotspot, determining patterns of gene flow across a number of endemic species with varying biological and ecological characteristics will be crucial for developing effective conservation management strategies.

## Acknowledgments

We are grateful for the valuable support and assistance provided by Sallyann Gudge and Ian Kerr from Lord Howe Island Marine Park. We thank the Lord Howe Island Board, Envirofund Australia (Natural Heritage Trust) and the Lord Howe Island Marine Park for financial and logistical support. We thank the Australian Department of the Environment and Water Resources for funding and the Capricorn Star for excellent logistical support. At Norfolk Island we thank Dave Biggs (Charter Marine), James Edward (Bounty Divers), Doug Creek, Michael Smith, Jack Marges, Karlene Christian, and Judith and Peter Davidson (Reserves and Forestry) for their assistance. This research was funded by the ARC Centre of Excellence for Coral Reef Studies (CEO561435).

## Conflict of Interest

None declared.

## References

- Allen, G. R., R. Steene, and M. Allen. 1998. A guide to angelfishes and butterflyfishes. Odyssey Publishing-Tropical Reef Research, Perth.
- Almany, G. R., M. L. Berumen, S. R. Thorrold, S. Planes, and G. P. Jones. 2007. Local replenishment of coral reef fish populations in a marine reserve. *Science* 316:742–744.
- Bay, L. K., J. H. Choat, L. van Herwerden, and D. R. Robertson. 2004. High genetic diversities and complex genetic structure in an Indo–Pacific tropical reef fish (*Chlorurus sordidus*), evidence of an unstable evolutionary past? *Mar. Biol.* 144:757–767.
- Beerli, P. 2004. Effect of unsampled populations on the estimation of population sizes and migration rates between sampled populations. *Mol. Ecol.* 13:827–836.
- Beerli, P., and J. Felsenstein. 2001. Maximum likelihood estimation of a migration matrix and effective population sizes in *n* subpopulations by using a coalescent approach. *Proc. Natl. Acad. Sci.* 98:4563–4568.
- Bell, J. J., and B. Okamura. 2005. Low genetic diversity in a marine nature reserve, re-evaluating diversity criteria in reserve design. *Proc. Biol. Sci.* 272:1067–1074.
- Bellwood, D. R. 1996. The Eocene fishes of Monte Bolca, the earliest coral reef fish assemblage. *Coral Reefs* 15:11–19.
- Bellwood, D. R., and P. W. Wainwright. 2002. The history and biogeography of fishes on coral reefs. Pp. 5–32 in P. F. Sale, ed. *Coral reef fishes, dynamics and diversity in complex ecosystems*. Academic Press, San Diego.
- Bellwood, D. R., T. P. Hughes, C. Folke, and M. Nystrom. 2004. Confronting the coral reef crisis. *Nature* 429:827–833.
- Bellwood, D. R., S. Klanten, P. F. Cowman, M. S. Pratchett, N. Konow, and L. van Herwerden. 2010. Evolutionary history of the butterflyfishes (f, Chaetodontidae) and the rise of coral feeding fishes. *J. Evol. Biol.* 23:335–349.
- Benjamini, Y., and Y. Hochberg. 1995. Controlling the false discovery rate, a practical and powerful approach to multiple testing. *J. R. Stat. Soc., Ser. B* 57:289–300.
- Botsford, L. W. 2005. Potential contributions of marine reserves to sustainable fisheries, recent modeling results. *Bull. Mar. Sci.* 76:245–260.
- Brothers, E. B., and R. E. Thresher. 1985. Pelagic duration, dispersal and the distribution of Indo-Pacific coral reef fishes. Pp. 53–69 in M. L. Reaka, ed. *The ecology of coral reefs*. Symposia Series for Undersea Research. National Oceanic and Atmospheric Administration, Washington, D.C.
- Brothers, E. B., D. M. Williams, and P. F. Sale. 1983. Length of larval life in twelve families of fishes at 'One Tree Lagoon' Great Barrier Reef, Australia. *Mar. Biol.* 76: 319–324.
- Choat, J. H., L. van Herwerden, W. D. Robbins, J-P. A. Hobbs, and A. M. Ayling. 2006. A report on the ecological surveys conducted at Middleton and Elizabeth Reefs, February 2006. Report to the Australian Government Department of Environment and Heritage, Canberra (unpublished report).
- Cole, A. J., M. S. Pratchett, and G. P. Jones. 2008. Diversity and functional importance of coral-feeding fishes on tropical coral reefs. *Fish Fish.* 9:286–307.
- Cowen, R. K., K. M. M. Lwiza, Su Sponaugle, C. B. Paris, and D. B. Olson. 2000. Connectivity of marine populations, open or closed? *Science* 287:857–859.
- Cowen, R. K., C. B. Paris, D. B. Olson, and J. L. Fortuna. 2002. The role of long distance dispersal versus local retention in replenishing marine populations. *Gulf Carrib. Res.* 14:129–138.
- Craig, M. T., J. A. Eble, and B. W. Bowen. 2010. Origins, ages and population histories, comparative phylogeography of endemic Hawaiian butterflyfishes (genus *Chaetodon*). *J. Biogeogr.* 37:2125–2136.
- De'ath, G., K. E. Fabricius, H. Sweatman, and M. Puotinen. 2012. The 27-year decline of coral cover on the Great Barrier Reef and its causes. *Proc. Natl Acad. Sci.* 199: 17995–17999.

- Dudgeon, C. L., N. Gust, and D. Blair. 2000. No apparent genetic basis to demographic differences in scarid fishes across continental shelf of the Great Barrier Reef. *Mar. Biol.* 137:1059–1066.
- Dulvy, N. K., Y. Sadovy, and J. D. Reynolds. 2003. Extinction vulnerability in marine populations. *Fish Fish.* 4:25–64.
- Evans, R. D., L. van Herwerden, G. R. Russ, and A. J. Frisch. 2010. Strong genetic but not spatial subdivision of two reef fish species targeted by fishers on the Great Barrier Reef. *Fish. Res.* 102:16–25.
- Excoffier, L., G. Laval, and S. Schneider. 2005. Arlequin (version 3.0), an integrated software package for population genetics data analysis. *Evol. Bioinform. Online* 1:47–50.
- Faubet, P., R. Waples, and O. Gaggiotti. 2007. Evaluating the performance of a multi-locus Bayesian method for the estimation of migration rates. *Mol. Ecol.* 16:1149–1166.
- Frankham, R. 1997. Do island populations have less genetic variation than mainland populations? *Heredity* 78:311–327.
- Gaither, M. R., R. J. Toonen, D. R. Robertson, S. Planes, and B. W. Bowen. 2010. Genetic evaluation of marine biogeographical barriers, perspectives from two widespread Indo-Pacific snappers (*Lutjanus kasmira* and *Lutjanus fulvus*). *J. Biogeogr.* 37:133–147.
- Gardner, T. A., I. M. Côté, J. A. Gill, A. Grant, and A. R. Watkinson. 2003. Longterm region-wide declines in Caribbean corals. *Science* 301:958–960.
- Gardner, M. G., A. J. Fitch, T. Bertozzi, and A. J. Lowe. 2011. Rise of the machines—recommendations for ecologists when using next generation sequencing for microsatellite development. *Mol. Ecol. Resour.* 11:1093–1101.
- Gaston, K. J. 1998. Rarity as double jeopardy. *Nature* 394:229–230.
- Gold, J. R., E. Saillant, N. J. Cummings, and M. A. Renshaw. 2011. Genetic divergence and effective size among lane snapper in US waters of the Western Atlantic ocean. *North Am. J. Fish. Manag.* 31:209–223.
- Graham, N. A. J. 2007. Ecological versatility and the decline of coral feeding fishes following climate driven mortality. *Mar. Biol.* 153:127–199.
- Graham, N. A. J., S. K. Wilson, S. Jennings, N. V. C. Polunin, J. P. Bijoux, and J. Robinson. 2006. Dynamic fragility of oceanic coral reef ecosystems. *Proc. Natl Acad. Sci.* 103:8425–8429.
- Grant, W. S., and B. M. Bowen. 1998. Shallow population histories in deep evolutionary lineages of marine fishes, insights from the sardines and anchovies and lessons for conservation. *J. Hered.* 89:415–426.
- Halpern, B. S., and R. R. Warner. 2003. Review Paper Matching marine reserve design to reserve objectives. *Proc. Biol. Sci.* 270:1871–1878.
- Hanski, I. A. 1999. *Metapopulation ecology*. Oxford University Press, Oxford.
- Harrison, H. B., D. H. Williamson, R. D. Evans, G. R. Almany, S. R. Thorrold, G. R. Russ, et al. 2012. Larval export from marine reserves and the recruitment benefit for fish and fisheries. *Curr. Biol.* 22:1023–1028.
- Hastings, A., and L. W. Botsford. 2006. Persistence of spatial populations depends on returning home. *Proc. Natl. Acad. Sci.* 103:6067–6072.
- Hellberg, M. E., R. S. Burton, J. E. Neigel, and S. R. Palumbi. 2002. Genetic assessment of connectivity among marine populations. *B. Mar. Sci.* 70:273–290.
- Hilborn, R., F. Micheli, and G. A. De Leo. 2006. Integrating marine protected areas with catch regulation. *Can. J. Fish. Aquat. Sci.* 63:642–649.
- Hobbs, J.-P. A., J. Neilson, and J. J. Gilligan. 2009. Distribution, abundance, habitat association and extinction risk of marine fishes endemic to the Lord Howe Island region. Report to Lord Howe Island Marine Park (unpublished report).
- Hobbs, J.-P. A., G. P. Jones, and P. L. Munday. 2011. Extinction risk in endemic marine fishes. *Conserv. Biol.* 25:1053–1055.
- Hoegh-Guldberg, O. 1999. Coral bleaching, climate change and the future of the world's Coral Reefs. *Mar. Freshw. Res.* 50:839–866.
- Hoelzel, A. R., R. C. Fleischer, C. Campagna, B. J. Le Boeuf, and G. Alvord. 2002. Impact of a population bottleneck on symmetry and genetic diversity in the northern elephant seal. *J. Evol. Biol.* 15:567–575.
- Hubisz, M. J., D. Falush, M. Stephens, and J. K. Pritchard. 2009. Inferring weak population structure with the assistance of sample group information. *Mol. Ecol. Resour.* 9:1322–1332.
- Hughes, T. P., A. H. Baird, D. R. Bellwood, M. Card, S. R. Connolly, C. Folke, et al. 2003. Climate change, human impacts, and the resilience of coral reefs. *Science* 301:929–933.
- Johannesson, K., and C. Andre. 2006. Life on the margin, genetic isolation and diversity loss in a peripheral marine ecosystem, the Baltic Sea. *Mol. Ecol.* 15:2013–2029.
- Jombart, T., S. Devillard, and F. Balloux. 2010. Discriminant analysis of principal components, a new method for the analysis of genetically structured populations. *BMC Genet.* 11:94.
- Jones, J. G., and I. McDougall. 1973. Geological history of Norfolk and Philip Islands, southwest Pacific Ocean. *J. Geol. Soc. Aust.* 20:239–254.
- Jones, G. P., M. J. Caley, and P. L. Munday. 2002. Rarity in coral reef fish communities. Pp. 81–101 in P. F. Sale, ed. *Coral reef fishes, dynamics and diversity in complex ecosystems*. Academic Press, San Diego.
- Jones, G. P., M. I. McCormick, M. Srinivasan, and J. V. Eagle. 2004. Coral decline threatens fish biodiversity in marine reserves. *Proc. Natl. Acad. Sci.* 101:8251–8253.
- Jones, G. P., S. Planes, and S. R. Thorrold. 2005. Coral reef fish larvae settle close to home. *Curr. Biol.* 15:1314–1318.
- Jones, G. P., G. R. Almany, G. R. Russ, P. F. Sale, R. R. Steneck, M. J. H. van Oppen, et al. 2009. Larval retention



- and connectivity among populations of corals and reef fishes, history, advances and challenges. *Coral Reefs* 28:307–325.
- Kokita, T., and A. Nakazono. 2001. Rapid response of an obligately corallivorous filefish *Oxymonacanthus longirostris* (Monacanthidae) to a mass coral bleaching event. *Coral Reefs* 20:155–158.
- Kuiter, R. H. 1996. *Guide to sea fishes of Australia*. New Holland, Sydney.
- Largier, J. 2003. Considerations in estimating larval dispersal distances from oceanographic data. *Ecol. Appl.* 13:71–89.
- Lawton, R. J., V. Messmer, M. S. Pratchett, and L. K. Bay. 2011. High gene flow across large geographic scales reduces extinction risk for a highly specialised coral feeding butterflyfish. *Mol. Ecol.* 20:3584–3598.
- Leis, J. M., L. van Herwerden, and H. M. Patterson. 2011. Estimating connectivity in marine fish populations, What works best? *Oceanogr. Mar. Biol. Annu. Rev.* 49:193–234.
- Marine Parks Authority. 2010. Natural values of Lord Howe Island. NSW Marine Parks Authority (unpublished report).
- McDougall, I., B. J. J. Embleton, and D. B. Stone. 1981. Origin and evolution of Lord Howe Island, Southwest Pacific. *J. Geol. Soc. Aust.* 28:155–176.
- McNeely, J. A., K. R. Miller, W. V. Reid, R. A. Mittermeier, and T. B. Werner. 1990. *Conserving the world's biological diversity*. World Conservation Union, World Resources Institute, Conservation International, World Wildlife Fund-US, and the World Bank, Washington, D.C.
- van der Meer, M. H., J.-P. A. Hobbs, G. P. Jones, and L. van Herwerden. 2012a. Genetic connectivity among and self-replenishment within island populations of a restricted range subtropical reef fish. *PLoS ONE* 7:e49660.
- van der Meer, M. H., J.-P. A. Hobbs, G. P. Jones, and L. van Herwerden. 2012b. Historic hybridisation between two Australian anemonefish species *Amphiprion* and present-day patterns of connectivity. *Ecol. Evol.* 2:1592–1604.
- van der Meer, M. H., M. G. Gardner, J.-P. A. Hobbs, M. S. Pratchett, and L. van Herwerden. 2012c. Identification of 21 microsatellite loci for conservation genetic studies of the endemic butterflyfish *Chaetodon tricinctus*. *Conserv. Genet. Resour.* 4:243–246.
- Miller, J. A., and A. L. Shanks. 2004. Evidence for limited larval dispersal in black rockfish (*Sebastes melanops*), implications for population structure and marine-reserve design. *Can. J. Fish. Aquat. Sci.* 61:1723–1735.
- Montanari, S. R., L. van Herwerden, M. S. Pratchett, and J.-P. A. Hobbs. 2011. Reef fish hybridization, lessons learnt from butterflyfishes (genus *Chaetodon*). *Ecol. Evol.* 2:310–328.
- Mora, C., and P. F. Sale. 2002. Are populations of coral reef fishes open or closed? *Trends Ecol. Evol.* 17:422–428.
- Munday, P. L. 2004. Habitat loss, resource specialisation, and extinction on coral reefs. *Glob. Change Biol.* 10:1642–1647.
- Paetkau, D., W. Calvert, I. Stirling, and C. Strobeck. 1995. Microsatellite analysis of population structure in Canadian polar bears. *Mol. Ecol.* 3:489–495.
- Paetkau, D., R. Slade, M. Burden, and A. Estoup. 2004. Genetic assignment methods for the direct, real-time estimation of migration rate, a simulation-based exploration of accuracy and power. *Mol. Ecol.* 13:55–65.
- Palstra, F. P., M. F. O'Connell, and D. E. Ruzzante. 2007. Population structure and gene flow reversals in Atlantic salmon (*Salmo salar*) over contemporary and long-term temporal scales, effects of population size and life history. *Mol. Ecol.* 16:4504–4522.
- Piry, S., A. Alapetite, J. M. Cornuet, D. Paetkau, L. Baudouin, and A. Estoup. 2004. GENECLASS2, a software for genetic assignment and first generation migrants detection. *J. Hered.* 95:536–539.
- Planes, S. 2002. Biogeography and larval dispersal inferred from population genetic analysis. Pp. 201–220 *in* P. F. Sale, ed. *Coral reef fishes, dynamics and diversity in a complex ecosystem*. Academic Press, New York.
- Planes, S., G. P. Jones, and S. R. Thorrold. 2009. Larval dispersal connects fish populations in a network of marine protected areas. *Proc. Natl. Acad. Sci.* 106:5693–5697.
- Posada, D. 2008. jModelTest, phylogenetic model averaging. *Mol. Biol. Evol.* 25:1253–1256.
- Pratchett, M. S., S. K. Wilson, and A. H. Baird. 2006. Declines in the abundance of *Chaetodon* butterflyfishes (Chaetodontidae) following extensive coral depletion. *J. Fish Biol.* 69:1269–1280.
- Pratchett, M. S., P. L. Munday, S. K. Wilson, N. A. J. Graham, J. E. Cinner, D. R. Bellwood, et al. 2008. Effects of climate induced coral bleaching on coral reef fishes-ecological and economic consequences. *Oceanogr. Mar. Biol. Annu. Rev.* 46:251–296.
- Pritchard, J. K., M. Stephens, and P. Donnelly. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155:945–959.
- Randall, J. E. 1976. The endemic shore fishes of the Hawaiian Islands, Lord Howe Island, and Easter Island. *Trav. Doc. ORSTOM* 47:49–73.
- Randall, J. E. 1998. Zoogeography of shore fishes of the Indo-Pacific region. *Zool. Stud.* 37:227–268.
- Reed, D. H., and R. Frankham. 2003. Correlation between fitness and genetic diversity. *Conserv. Biol.* 17:230–237.
- Rotjan, R. D., and S. M. Lewis. 2008. Impact of coral predators on tropical reefs. *Mar. Ecol. Prog. Ser.* 367:73–91.
- Selkoe, K. A., and R. J. Toonen. 2006. Microsatellites for ecologists, a practical guide to using and evaluating microsatellite markers. *Ecol. Lett.* 9:615–629.
- Shanks, A. L., B. A. Grantham, and M. H. Carr. 2003. Propagule dispersal distance and the size and spacing of marine reserves. *Ecol. Appl.* 13:159–169.
- Sheppard, C. R. C. 2003. Predicted recurrences of mass coral mortality in the Indian Ocean. *Nature* 425:294–297.



- Shulman, M. J. 1998. What can population genetics tell us about dispersal and biogeographic history of coral–reef fishes. *Aust. J. Ecol.* 23:216–225.
- Suthers, I. M., J. M. Young, M. R. Roughan, J. D. Everett, G. B. Brassington, M. Byrne, et al. 2011. The strengthening East Australian Current, its eddies and biological effects—an introduction and overview. *Deep-Sea Res. Part II* 58:538–546.
- Swearer, S. E., J. E. Caselle, D. W. Lea, and R. R. Warner. 1999. Larval retention and recruitment in an island population of coral-reef fish. *Nature* 402:799–802.
- Syms, C. 1998. Disturbance and the structure of coral reef fish communities on the reef slope. *J. Exp. Mar. Biol. Ecol.* 230:151–167.
- Thresher, R. E., P. L. Colin, and L. J. Bell. 1989. Planktonic duration, distribution and population structure of western and central Pacific damselfishes (Pomacentridae). *Copeia* 1989:420–434.
- Victor, B. C. 1986. Duration of the planktonic larval stage of one hundred species of Pacific and Atlantic wrasses (family Labridae). *Mar. Biol.* 90:317–326.
- Victor, B. C., and G. M. Wellington. 2000. Endemism and the pelagic larval duration of reef fishes in the eastern Pacific Ocean. *Mar. Ecol. Prog. Ser.* 205:241–248.
- Waples, R. S., and O. Gaggiotti. 2006. What is a population? An empirical evaluation of some genetic methods for identifying the number of gene pools and their degree of connectivity. *Mol. Ecol.* 15:1419–1439.
- Wellington, G. M., and B. C. Victor. 1989. Planktonic larval duration of one hundred species of Pacific and Atlantic damselfishes (Pomacentridae). *Mar. Biol.* 101:557–568.
- Whittaker, R. J., and J. M. Fernández-Palacios. 2007. *Island biogeography, ecology, evolution, and conservation*. 2nd ed. Oxford University Press, Oxford.
- Wilkinson, C. 2002. *Status of coral reefs of the world, 2002*. Australian Institute of Marine Science, Townsville, Australia.
- Wilson, G. A., and B. Rannala. 2003. Bayesian inference of recent migration rates using multilocus genotypes. *Genetics* 163:1177–1191.
- Wilson, S. K., N. A. J. Graham, M. S. Pratchett, G. P. Jones, and N. V. C. Polunin. 2006. Multiple disturbances and the global degradation of coral reefs, are reef fishes at risk or resilient? *Glob. Change Biol.* 12:2220–2234.
- Wilson, S. K., R. Fisher, M. Pratchett, N. A. J. Graham, N. K. Dulvy, R. A. Turner, et al. 2008. Exploitation and habitat degradation as agents of change within coral reef fish communities. *Glob. Change Biol.* 14:2796–2809.
- Winters, K. L., L. van Herwerden, J. H. Choat, and D. R. Robertson. 2010. Phylogeography of the Indo-Pacific parrotfish *Scarus psittacus*, isolation generates distinctive peripheral populations in two oceans. *Mar. Biol.* 157:1679–1691.
- Wood, R. 1999. *Reef Evolution*. Oxford University Press, New York.

## Supporting Information

Additional Supporting Information may be found in the online version of this article:

**Table S1.** Sample sizes for mtDNA (cytochrome b, total  $n = 97$ ).

**Table S2.** Summary statistics for twenty microsatellite loci (Ct2–24) from *Chaetodon tricinctus*.

**Table S3.** AMOVA analysis for (A) mtDNA (Cyt b) sequences from *Chaetodon tricinctus* structured into the westernmost location (MR, ER, LHI) versus the peripheral location (NI) and (B) global AMOVA weighted across all 20 microsatellite loci.

**Table S4.** Pairwise population structures ( $F_{st}$ ) generated for mtDNA (cytochrome b,  $n = 97$ ) and for twenty microsatellite loci ( $n = 108$ ) from four *Chaetodon tricinctus* locations showing raw and corrected  $F'_{st}$  for null allele frequencies.

**Table S5.** AMOVA fixation indices ( $F'_{st}$ ) for *Chaetodon tricinctus* across all locations surveyed.