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# PHYTOCHEMICAL AND ANTIOXIDANT SCREENING OF ANACYLUS PYRETHRUM, APIUM GRAVEOLENS, BOERHAAVIA DIFFUSA, CINNAMOMUM CASSIA BLUME, CUSCUMIS MELO LINN, CUSCUMIS SATIVUS LINN, DAUCUS SATIVUS, FOENICULUM VULGARE, TRACHYSPERMUM AMMII AND THEIR EFFECT ON VARIOUS HUMAN AILMENTS

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## ARTICLE DETAILS

## ABSTRACT

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Herbal medicinal plants are commonly intended for the cure and prevention of countless diseases for the reason that of low price, more effectiveness and no side effects. The medicinal plants consist of several types of phytochemical constituents as alkaloids, saponin, flavonoids, phenols, tannins and carbohydrates that are used for various human ailments. Such types of constituents not only have biochemical compounds but also have antioxidant, antimicrobial, antifungal activity and intended for the treatment of diabetes, kidney, liver and stomach ailments. The use of herbal natural plants is more economic and reliable for these types of diseases. These medicinal plants have been second-hand for different human disorders with no side effects of reoccurrence.

### KEYWORDS

Phytochemical, Herbal medicinal, human ailments, antimicrobial, antioxidant.

## 1. INTRODUCTION

The medicinal herbal plants are second-hand in the urban countries population for the excellent human health continuation while the less developed countries population (3.3 billion) used the plants on routine basis that are observed by UNESCO. Thousands of years ago, the herbal plants consist of chief constituents which are used as natural remedies for the whole populations [1]. In the world, the plant species are about to 258, 650, out of 10% plants are used for the treatment of diseases in world populations. Majority of Pakistani people depends upon the medicinal plants for the treatment of major and minor diseases [2]. Alkaloids, phenols, tannins and flavonoids are the principal constituents having different characteristics which are used for different human health problems [3]. Mostly 75-80 % of world population depends upon the herbal medicine for primary health care, predominantly in developing countries. According to World Health Organization (WHO) the used of herbal product are increasing two (2) to three (3) times all over the world [4].

Reactive oxygen species (ROS) are generally known as oxygen free radical that has a role as a secondary messenger in intracellular signaling pathways [5]. Reactive oxygen species (ROS) are very reactive molecules because of presence of unpaired electron in their outmost shell. Reactive oxygen species (ROS) are generated in the living organism as a byproducts of regular cellular metabolic reactions. Physiologically reactive oxygen species (ROS) are produced in each cell at low concentration but if they are created at high concentration they can interact with essential cellular targets, including proteins, lipids and DNA, compromising cell viability and functions [6]. In this review paper, the *Anacyclus pyrethrum*, *Apium graveolens*, *Boerhaavia diffusa*, *Cinnamomum cassia blume*, *Cuscutis melo linn*, *Cuscutis sativus linn*, *Daucus sativus*, *Foeniculum vulgare*, *Trachyspermum Ammii* are to be discussed as phytochemical and ROS

screening and used for different human health.

## 2. REVIEW OF LITERATURE

### 2.1 *Anacyclus Pyrethrum*

*Anacyclus pyrethrum* belongs to family Asteraceae. *Anacyclus pyrethrum L.* (Asteraceae) commonly known as Akarkara widely used in Ayurvedic system of medicine [7]. Analysis of plant roots indicates the presence of ten (10) unsaturated amides, a sterol and three (3) fatty acids. Plant roots had the most important compounds such as phenylethylamine, polyacetylenic amides, sesamin, pellitorin, anacyclin and inulin [8]. It is used for the treatment of paralysis, epilepsy, toothache, rheumatism, sciatic and neuralgia [9]. The roots of medicinal plant had a various pharmacological effects including immunostimulating, memory-enhancing, insecticidal, local anesthetic, antidepressant and anticonvulsant [10].

Antioxidants are able to block the free radical mediated disease such as hepatic disorders, diabetes, neurodegenerative disorders, hemorrhagic shock and carcinogenesis [11]. The antibacterial potential of methanolic and aqueous extract of *Anacyclus Pyrethrum* was determined against six (6) bacterial strains such as *Bacillus subtilis*, *Aeruginosa*, *Escherichia coli*, *Klebsiella pneumonia*, *Staphylococcus aureus*, *Pseudomonas* and *Micrococcus luteus*. It had a broad spectrum of antibacterial activity which was determined with the help of disk diffusion and minimum inhibitory concentration (MIC) methods [12]. The antioxidant effects of *A. pyrethrum* were due to presence of phenolic and flavonoid compounds in extracts [13].

The aqueous *Anacyclus pyrethrum* root extract was used to determine the antidiabetic potential in diabetes induced rats. Aqueous extract was orally administered to rats at the dose of 150mg/kg and 300mg/kg. After the

treatment, blood glucose level returned back to near standard values in the diabetic rats [8]. The root decomposition showed the presence of a resin compounds called pyrethrin or pellitorin and alkyl amide have positive effects on reproduction and have directly or indirectly positive effects on testosterone to increase reproductive activity [14]. Extracts (*Anacyclus pyrethrum*) of roots with water had considerable effects on spermatogenesis and sperm count which used as an aphrodisiac and sexual stimulant but also have anabolic effects [15]. *Anacyclus pyrethrum* roots extract produce antidepressant effect due to interaction between adrenergic and dopamine receptor with increasing level of dopamine and noradrenaline in the brains of mice [16].

Aqueous root extracts of *Anacyclus pyrethrum* L. was used to determine the effects on gonadotropins (LH and FSH) and testosterone serum level in adult male rats. The results showed that the aqueous root extracts had significantly increased in serum level of gonadotropin hormone and testosterone serum [14]. The ethanolic extract of *Anacyclus pyrethrum* is

useful in the treatment of memory dysfunction. When ethanolic extract of *Anacyclus pyrethrum* was administered orally, it improves the memory. It increases the brain cholinesterase level due to increase central cholinergic neurotransmission and had a memory enhancing activity in scopolamine induced amnesia [17]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Anacyclus pyrethrum* has been reported

## 2.2 *Apium graveolens*

The family of *Apium graveolens* is Umbelliferae [18]. *Apium Graveolens* (Umbelliferae) are commonly known as Celery [19]. *Apium graveolens* (Celery) contains a major bioactive compound called furanocoumarins which is class of phenolic compounds.

**Table 1:** (A) Qualitative analysis of plants

Medicinal Plants	Anthocyanins	Alkaloids	Amino acids	Carbohydrates	Fixed oils	Flavonoids	Glycosides	Phenols	Proteins	Saponin	Steroids	Tannins	Terpenoids
<i>Anacyclus pyrethrum</i> [9,13]	-	+ +		+		+ +				+ -	+ +	+ +	+ +
<i>Apium graveolens</i> [18]		+	+	+		+	+	+		+	+	+	+
<i>Boerhaavia diffusa</i> [20]		+				+	-			+	-	+	-
<i>Cinnamomum cassia</i> [21,22]		+ +		+		+ +	+		+			+	+ +
<i>Cuscutis melo linn</i> [23,24]		+ -	+ +	+		+ +	- -	+	+				
<i>Cuscutis sativus linn</i> [25,26]		+				-	+						+ + +

+Ve: Indicate positive test.

-Ve: Indicate negative test.

**Table 1:** (B) Qualitative analysis of plants

Medicinal Plants	Anthocyanins	Alkaloids	Amino acids	Carbohydrates	Fixed Oils	Flavonoids	Glycosides	Phenols	Proteins	Saponin	Steroids	Tannins	Terpenoids
<i>Daucus carota</i> [27]		+		+	-		+	+	+	-	-	+	
<i>Foeniculum vulgare</i> [28]		-				+	+			+		-	-
<i>Trachyspermum Ammi</i> [29]		+	-			+	+	+	-	-	+	+	-

**+Ve:** Indicate positive test,

**-Ve:** Indicate negative test.

Ferric reducing antioxidant power (FRAP) method was used to assess the antioxidant capacity of *Anacyclus pyrethrum*. Due to the presence of antioxidants in plant extracts, Fe<sup>3+</sup>/ferricyanide reduce to the ferrous form which could be monitored at 700nm. Maximum ferric reductive

capacity at 0.15mg/mL with methanolic extract was 0.569mg/mL as compared to water and chloroform extracts 0.301mg/mL and 0.182mg/mL respectively. At 0.15 mg/mL ascorbic acid had the highest reducing power as compared to all the extracts which was 1.84 mg/mL [13].

**Table 2:** (A) Quantitative phytochemical screening of plants

1-Quantitative Analysis of <i>Anacyclus pyrethrum</i>				
Name of constituent	Type of extract	Assay Values	References	
Phenols (a)	Methanol	310.78 ± 5.2	[13]	
	Water	183.82 ± 3.1		
	Chloroform	91.8 ± 1.7		
Flavonoids (b)	Methanol	92.50 ± 4.2		
	Water	72.50 ± 2.1		
	Chloroform	24.20 ± 1.2		
(a) µg gallic acid equivalent per mg of extract (mg GA/g) (b) mg quercetin equivalent per g of extract (mg QE/g)				
2-Quantitative Analysis of <i>Apium graveolens</i>				
Phenols	Ethyl Acetate	22.70 ± 1.56 mg/g	[30]	
	Methanol	51.09 ± 1.44 mg/g		
	Butanol	19.43 ± 0.88 mg/g		
	Water	46.40 ± 0.31 mg/g		
Flavonoids	Ethyl Acetate	4.08 ± 0.31 mg/g		
	Methanol	2.12 ± 0.08 mg/g		
	Butanol	4.80 ± 0.03 mg/g		
	Water	0.77 ± 0.01 mg/g		
3-Quantitative Analysis of <i>Boerhaavia diffusa</i>				
Carbohydrate		10.56±0.12		[31]
Protein		2.26±0.02		
Vitamin C		44.80±5.78		
Vitamin B1		ND		
Vitamin B2		22.00±4.25		
Vitamin B3		97.00 ±8.01		
Vitamin B6		ND		

**Table 2:** (B) Quantitative phytochemical screening of plants

4-Quantitative Analysis of <i>Cinnamomum cassia</i>				
Name of constituent	Type of extract	Assay Values	References	
(a) Phenols (b) Flavonoids		5.59±0.06 3.12±0.07	[32]	
(a) µg gallic acid equivalent per mg of extract (mg GA/g) (b) mg quercetin equivalent per g of extract (mg QE/g)				
5-Quantitative Analysis of <i>Cucumis sativus</i>				
Phenols	Fresh extract Dried extract	19.25 (µg/mg) 6.23 (µg/mg)	[33]	
Flavonoids	Fresh extract Dried extract	16.24 (µg/mg) 1.82 (µg/mg)		
6-Quantitative Analysis of <i>Daucus carota Linn</i>				
Total phenolic contents	leaves extract seeds extract	13.83 mg/g 7.08 mg/g	[34]	
7-Quantitative Analysis of <i>Foeniculum vulgare Mill</i>				
Total phenolic contents	Hexane fraction Chloroform fraction Methanol fraction	Nil 10.51 mg/g 48.37 mg/g	[35]	
Total flavonoid contents	Hexane fraction Chloroform fraction Methanol fraction	Nil 5.26 mg/g 21.44 mg/g		
8-Quantitative Analysis of <i>Trachyspermum ammi</i>				
Carbohydrates Protein Fat Fibre Moisture Minerals matter	38.6 µg/mg 15.4 µg/mg 18.1 µg/mg 11.9 µg/mg 8.9 µg/mg 7.1 µg/mg		[36]	

**Table 3:** Antioxidant analysis of the plants

Plants	DPPH(2,2-diphenyl-1-picrylhydrazyl)	FRAP (Ferricreducing)	NO (Nitrogen oxide radical)	OH (Hydroxyl radical)	O <sub>2</sub> <sup>-</sup> (Superoxide anion radical)	H <sub>2</sub> O <sub>2</sub> (Hydrogen peroxide)	References
<i>Anacyclus pyrethrum</i>	55.83±1.92 µg/ml	NR	32.61±1.68 µg/ml	60.14±0.43 µg/ml	NR	38.54±0.94 µg/ml	[7]
<i>Apium graveolens</i>	1.41±0.270 µg/ml	12.48±1.06 µg/ml	NR	NR	NR	NR	[37]

<i>Boerhaavia diffusa</i>	82.12 µg/ml	19.48±1.0 2 µg/ml	NR	NR	99.8 µg/ml	NR	[38]
<i>Cinnamomum cassia</i>	42.03±0.06 (µg/L)	NR	NR	NR	NR	NR	[39]
<i>Cuscumis melo</i>	21.8±0.2 µg/ml	NR	17.7±0.2 µg/ml	18.7±0.1 µg/ml	16.8±0.2 µg/ml	17.8±0.1 µg/ml	[40]
<i>Cuscumis sativus</i>	13.06 µg/ml	NR	NR	NR	NR	NR	[41]
<i>Daucus carota</i>	136±3.21 µg/ml	NR	NR	NR	NR	NR	[34]
<i>Foeniculum vulgare</i>	32.32 ± 0.77 (µg/ml)	NR	NR	NR	NR	NR	[42]
<i>Trachyspermm Ammi</i>	73.41 µg/ml	NR	67.33 µg/ml	62.48 µg/ml	63.22 µg/ml	NR	[29]

NR: Not Reported in Literature

The main furanocoumarins in celery are three phototoxic furanocoumarins, xanthotoxin, psoralen and bergapten. It is a strong diuretic and also have urinary antiseptic properties due to the volatile oil called apiol [43]. The quantitative phytochemical constitutions are D-Limonene (57.7%), Myrcene (18.74%), Terpeneol (8.6%), β-Selinene (8.1%), β-pinen (2.4%), β-caryophyllene (0.5%), Carnone (0.3%), Trans-limonene oxide (0.3%), α-Terpinolene (0.3%), α-selinene (0.2%), Trans-3-butylideneephthalide (0.1%), α-Muuroloene (0.1%), Cis-limonene oxide (0.1%), Linalool (0.1%), α-pinen (0.1%) and Trans-ocimene (0.1%) [44]. The medicinal plant have ability to do work as antibacterial, antifungal, anti-inflammatory, antioxidant, anthelmintic, antispasmodic, carminative, diuretic, laxative and sedative [44]. The recommended dose of celery seeds is about to 1.2 gm to 4 gm daily and fresh plant juice is about to 23gm (15ml) three (3) times in a day [45]. It is also used as alternative therapy for the patients suffering with arthritis, rheumatism and gout due to its anti-inflammatory effects [19].

The flavonoids had the ability to collect a free radicals and antioxidant potential. The flavonoids and phenolic compounds have pharmacological features such as anti-oxidants, anti-mutagenic, anti-bacterial, anti-diabetic, anti-inflammatory, anti-thrombosis, hyperlipidemia and anti-cancer. *Apium Graveolens* (Celery) is one of the plants that are rich in flavonoid such as apigenin and apiin [30]. The name of celery was given to the dry food of *Apium graveolens*. The celery is commercially available as vegetable, seed, flaks and seed oleoresin [19]. Celery is known as mild

diuretic, used as a urinary antiseptic, antispasmodic, anthelmintic and sedative [44].

The celery has a very effective role in fertility rate due to its flavonoids and antioxidant properties. The hydro-alcoholic extract of celery leaves can be used for improving the fertility parameters. After taking the extract of the celery leaves it causes the improvement of the sperm parameters such as mobility and its number in male sex. The female sex hormone called progesterone also boosted due to celery [44]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Apium graveolens* has been reported.

### 2.3 *Boerhaavia Diffusa* Linn

The family of *Boerhaavia diffusa* is Nyctaginaceae [46]. The plant *Boerhaavia diffusa* (Nyctaginaceae) frequently known as punarnava [47]. This medicinal plant distributed in tropical region such as South America,

India and Africa. The phytochemical analysis of punarnava exposed that it contains steroids, lipids, lignins, flavonoids, alkaloids, triterpenoids, phlobaphenes and ursolic acid [20]. It contain main constituents such as boerhaavia acid, boeravinone, palmitic acid, sitosterol, esters of sitosterol, punarnavine, boerhaavia acid many other compounds [48].

*Boerhaavia diffusa* considered as a source of vitamins, carbohydrate, protein, tannin, saponin, flavonoid and terpinoid [46]. *Boerhaavia diffusa* are mostly used for the management of depression, diabetes, anemia, liver disorders, tumours, cancers and all kind of internal inflammations. The entire plant also used as antistress, antidiabetic, adoptogenic, antioxidant and immunostimulator. The plant has various beneficial effects in the treatment of tumors and abdominal cancers [49]. The antioxidant activity was observed due to presence of minerals, organic acids, flavonoids and phenolic compounds in the root of plant [46]. Roots of this medicinal plant were found to be exposed antioxidant potential which supports to use as traditional medicine [47].

The majority of serum parameters were altered such as alkaline phosphatase (ALP), glutamic oxaloacetic transaminase (GOT), acid phosphatase (ACP) and glutamic-pyruvic transaminase (GPT). It was found that a considerable hepatoprotective activity was observed in thioacetamide induced rats. The hepatotoxicity was experimentally induced with carbon tetrachloride in mice and rats [50]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Boerhaavia diffusa* has been reported.

### 2.4 *Cinnamomum Cassia*

*Cinnamomum cassia blume* belongs to the family Lauraceae. *Cinnamomum cassia* (Lauraceae) also called as *Cinnamomum aromaticum*, Chinese cinnamon or Chinese cassia, mostly cultivated in the countries of India, China, Uganda, Vietnam, Bangladesh and Pakistan. It is used as flavoring agent in various Asian countries [51]. According to another study, the Cinnamon contains cinnamate, cinnamic acid, cinnamaldehyde and essential oils. The essential oils which present in this medicinal plant are eugenol, L-borneol, caryophyllene oxide, cinnamyl acetate, trans-cinnamaldehyde terpineol, terpinolene, thujene, b-caryophyllene and L-bornyl [52]. The major constituents are cinnamaldehyde (75-90%), coumarin (7%) and essential oil (4%). The others constituents which are present in traces amount are salicylic acid, cinnamyl alcohol, eugenol, benzoic acid, cinnamic acid, corresponding esters and aldehydes [51]. The major antioxidant constituents in this plant are tannin, flavonoid, phenol and volatile oils. This medicinal herb is used in different traditional

medicine system such as Unani, Ayurvedic, Japanese and Chinese in the treatment of dyspepsia, diabetes, brain ischemia, cancers, peptic ulcer disease and ischemic brain injury [53]. It was reported that cinnamon used as flavoring agent. It can be used to reduce the risk of colon cancer with the improvement of colon health. The cinnamon used as coagulant which prevent the bleeding and to increase the blood circulation in uterus [52].

The gas chromatography-mass spectrometry (GC-MS) method was used to investigate the essential oil of *Cinnamomum* bark. The result exposed the major chemical constituents include eugenol (9.317%),  $\alpha$ -muurolene (0.133%), o-methoxy cinnamic aldehyde (0.236%), tricyclo-nona-3,6-dien-9-on (0.173%) and naphthalene,1,2,3,4-tetrahydro-1,6-dime

(0.195%). The 2-propenal, 3-phenyl (87.013%) considered as minor constituents. These constituents are accountable for the *Cinnamomum* bark fragrance and beneficial effects [54]. The essential oils and other components had various pharmacological activities such as antimicrobial, anti-inflammatory, antitermitic, antifungal, antioxidant, antidiabetic, antimycotic and anticancer. The traditional use of Cinnamon powder is to treat dental problems, toothaches and bad breath [52]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Cinnamomum Cassia* has been reported while antibacterial activity of *Cinnamomum cassia* has been given below (Table 4)

**Table 4:** Antibacterial activity of *Cinnamomum cassia* extracts against UTI isolated bacteria. [55]

Types of Extracts	Diameter of Inhibition Zone (DIZ) in mm				
	<i>E. coli</i>	<i>K.pneumoniae</i>	<i>P.aeruginosa</i>	<i>E.faecalis</i>	<i>P.mirabilis</i>
Aqueous	4.66±0.57	2.66±0.57	3.66±0.57	5.66±0.57	7.0±1.0
Methanolic	19.66±0.57	10.0±0.81	18.0±0.60	16.0±0.6	18.6±0.57
Ethanolic	21.33±0.57	15.66±0.57	19.66±0.57	17.66±0.57	20.33±0.57

According to this observational study *Cinnamomum Cassia* have ability to reduce low density lipoprotein (LDL) and serum triglyceride while high density lipoprotein (HDL) was increased in healthy volunteers [56]. According to Food and Drug Administration (FDA) and World Health Organization (WHO), the daily average dose of *Cinnamomum cassia* is about to 1.25mg/kg for adult male [51].

## 2.5 *Cucumis melo* Linn

*Cucumis melo linn* belongs to the family Cucurbitaceae [24]. *Cucumis melo* Linn (Cucurbitaceae), commonly known as musk melon or kharboza. The phytochemical study revealed that the seeds contain chromone derivatives, phenolic glycoside, arginine, sitosterol, beta-sitosterol, alpha-galactosidases, dihydroxy triterpenes, aspartic and glutamic acids [23]. The stems of this plant had an effective role in constipation, liver disorders, purgative and flatulence [57]. Anti-inflammatory and antioxidant effects were explored with the extract of *Cucumis melo* [58]. The seeds are commonly used for the treatment of kidney and bladder stones, oliguria, jaundice, chronic fevers, bile obstruction, eczema, general debility, painful and burning micturition and ulcers in the urinary tract.

The seeds of the plant are mostly used as diuretic, lithotripter, laxative and refrigerant [23].

The production of peroxynitrite was inhibited due to *Cucumis melo* extract, which increase the antioxidant potential. The production of superoxide anion was inhibited, which was dose dependent [58]. The methanolic extract at dose of 500mg/kg reflects the maximum antihyperlipidemic activity as compared to aqueous extract. This result showed that methanolic extract had potent antihyperlipidemic activity in high cholesterol diet induced hyperlipidemia, as compared to atorvastatin [59]. The diuretic effect was significantly increased in treated experimental animals. These results indicate the extracts containing the combination of *Cucumis melo* and *Macrotyloma uniflorum* can be used to manage the renal ailments such as hypertension and edema [60]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Cucumis melo* has been reported while antioxidant activity of *Cucumis melo* has been given below (Table 5).

**Table 5:** Antioxidant composition of the *Cucumis melo* extracts [58].

Antioxidant	Amounts in the extract
<b>Endogenous Enzymes</b>	
Superoxide dismutase	95 ± 8 IU/mg
Catalase	10 ± 2 IU/mg
Glutathion peroxidase	1 ± 0.5 IU/mg
<b>Exogenous Enzymes</b>	
Co-enzyme Q	10 54.0 ± 4 mg/100 g
Lipoic acid	19.3 ± 1 mg/100 g

Glutathion (GSH)	215 ± 12 µg/100 g
Glutathione disulfide (GSSG)	3075 ± 55 µg/100 g
Carotenoids	350 ± 34 µg/100 g
Selenium	2.5 ± 0.2 µg/100 g
Vitamin E	240 ± 22 µg/100 g
Vitamin A	10000 ± 154 µg/100 g
Vitamin C	5000 ± 523 µg/100 g

(Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Cucumis sativus L.* has been reported.

## 2.6 *Cucumis Sativus Linn*

*Cucumis sativus L.* belong to family Cucurbitaceae. It is commonly named as Cucumber (English) and Khira (Hindi). It is widely distributed in northern India and warm and temperate countries of the world. The phytochemical analysis revealed the presence of 2, 6-nonadienal, 2-nonenal, methyl-2-methylbutanoate, cucurbitacin C, pectin, codisterol, dehydroporiferasterol, galactinol, dihydroxyhexadecanoic acid and cucurbitacins [22]. It can be used for the management of menstrual illnesses, dyspepsia, asthma, piles, leprosy and bronchitis. The traditional used of *Cucumis sativus* are laxative, astringent, anthelmintic, anti-diarrheal and antipyretic. It is also used to manage the hepatitis, bronchitis, dyspepsia and asthma [61]. The used of medicinal plant includes anti-diarrheal, antimicrobial, antihyperlipidemic, antidiabetic, antipyretic, astringent, anthelmintic, laxative and purgative. The pharmacologically plant used as anti-allergic, antihypertensive, antidiabetic, antioxidant, anti-dermatitis and anti-fungal [22]. The protective activity against gastric ulcers was improved due to the presence of flavonoid, alkaloids, steroids and polyphenols. These phytoconstituents have ability to reduce the gastric acid volume, total acidity, free acidity, anti-inflammatory and antioxidant [62].

The ethanolic extract of cucumber had a significant improvement in hypoglycemic, hypolipidemic and glycogenesis in treated animals. The cucumber extract decrease the low density lipoprotein (LDL) reduce upto 13% and glucose level in blood upto 67% after single intraperitoneal injection. The total cholesterol and triglyceride level was reduced to 29% and 72% respectively. As a result, a significant improvement in glycogenesis was observed by using the *Cucumis sativus* extracts in diabetic rats [63]. The anti-inflammatory activity was investigated in rat, treating with the extract of methanolic of *Cucumis sativus*. The extract at the dose of 150mg/kg and 250mg/kg was orally administered. The methanolic extract reduced inflammation 72.06% (250mg/kg) and 57.35% (150mg/kg) in carrageenan-induced paw edema test. These effects were compared with 79.41% (10mg/kg) indomethacin [41].

The aqueous extract of *Cucumis sativus* fruit pulp was used to assess the carminative and antacid properties. It was determined with carbondioxide evolution method. The NaHCO<sub>3</sub> considered as standard drug for the comparison of results. The aqueous extract of *Cucumis sativus* fruit pulp possessed significant carminative and antacid properties [15]. The *Cucumis sativus* extracts was used to investigate the treatment effects on the acne vulgaris, due to its antimicrobial and antioxidant potential to cure acne. The fresh extract of *Cucumis sativus* was mixed with linseed (*Linum usitatissimum*) oils and tea tree (*Melaleuca alternifolia*) oils in the formulations of polyherbal cream for the treatment of acne vulgaris [33].

It was investigated that ethanolic extracts of the plant possessed cytotoxic activity. The ethanolic extracts showed significant reducing power due to its antioxidant potential [25]. The antifungal activity was determined against *Pityrosporum ovale*, *Trichophyton spp.*, *Microsporum spp.*, *Aspergillus niger*, *Blastomyces dermatitidis* and *Candida albicans* which showed significant results. This medicinal plants had widely used in

traditional medicine as anticancer agent, antifungal and pesticide due to presence of biologically active compounds [25]. Phytochemical qualitative

## 2.7 *Daucus Carota*

*Daucus carota Linn* belong to family Apiaceae generally known as carrots. The major active constituents are phenolic, phytosterol, triterpene and polyacetylene [64]. *Daucus carota* had a remarkable quantity of vitamins, nicotinic acid, flavonoids, α, β and γ-carotene lycopene, cryptoxanthin and leutenin. They also have carotenoids such as abscisic acid, trisporic acid and β-apocarotenoids [65]. The ethanolic extract of *Daucus carota* was used to investigate the wound healing property. The cream was prepared with 1%, 2% and 4% (w/w) of ethanolic extract by using soft paraffin as a base. The antioxidant and antimicrobial potential due to various compounds phytochemical constituents such as flavonoids and phenolic derivatives which are present in ethanolic root extract [66]. This medicinal plant can be used for the management of Alzheimer patients [67]. The juice extract of the roots *Daucus carota* had antiulcer and gastroprotective potential [65].

The *Daucus carota* seeds extract with methanol was used to investigate hypolipidemic activity in normal rats. These rats were treated with extract at the doses (200mg/kg and 400mg/kg) for seven days. The lovastatin at the dose of 7.2mg/kg used as standard drugs. After the treatment, lipid profiles were assessed in blood serum on eight day. The significant reduction of triglyceride, HDL, VLDL and total cholesterol was observed in treated groups as compared control group [68]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Daucus carota* has been reported.

## 2.8 *Foeniculum vulgare mill*

*Foeniculum vulgare Mill* (Umbelliferae) commonly known as fennel in traditional medicine. The major chemical constituent in the oil of *F. vulgare* are trans-anethole, fenchone, p-cymene, and methyl chavicol [69]. The vitamins and minerals are present in its composition such as vitamin A and C, phosphorous, iron, calcium, potassium, thiamine, niacin and riboflavin. The main constituents of Fennel volatile oil are fenchone (1-20%), anethole (40-70%) and estragole (2-9%). The *F. vulgare* also contains d-apenine, camphene, pectin and methyl chavicol. *Foeniculum vulgare* consists of fat (10%), minerals (13.4%), moisture (6.3%), proteins (9.5%), fiber (18.5%) and (42.3%) carbohydrates [28]. The dried fennel when used as nutrient, it contains carbohydrate, lipids (mono and polyunsaturated), protein, minerals and vitamins [70].

*Foeniculum vulgare* had antiseptic, anti-inflammatory and palliative effects, had ability to treat infertility in female albino mice due to folliculogenesis effects. The alcoholic extract of at doses of 100mg/kg and 200mg/kg were used [71]. It can be used for the management of digestive, endocrine, reproductive and respiratory complaints [69]. The pharmacological uses includes antimicrobial, anti-inflammatory, antipyretic, antitumor, antispasmodic, antidiabetic, hepatoprotective, hypolipidemic and memory enhancing effects [70]. The *Foeniculum vulgare* seeds were used to manage the polycystic ovary syndrome (PCOS). The renoprotective potential of *Foeniculum vulgare* aqueous extract was studied in female rats. The aqueous extract at the dose of 150 mg/kg was intragastrically administered. The estradiolvalerate at the dose of 4mg in 0.2ml of same oils was considered as a standard drug. The aqueous extract of fennel exhibited the positive effect on renal functions [72].

The analgesic activity of ethanolic extract at the dose of 200mgm/kg, 100mgm/kg and 50mgm/kg were determined in rats and mice. The anti-inflammatory activity of ethanolic *Foeniculum vulgare* extract was investigated. The conclusions of this study reflect the significant activities

of analgesic and anti-inflammatory such as rheumatism, fever and dental pain of ethanolic extract while their effect on antioxidant enzymes of *F. vulgare* has been given below [Table 6] [73].

**Table 6:** Effects of *F. vulgare* extracts on antioxidant enzymes, malondialdehyde and lipid levels in the plasma of rats [74].

Antioxidant Enzymes and Lipid Levels	Groups	
	Group Control	Group Treated with <i>F. vulgare</i> Extract
Superoxide dismutase (U/mg protein)	10.8± 0.167	19.867± 0.011
Catalase (nmol H <sub>2</sub> O <sub>2</sub> degraded/min/mg protein)	10.14± 3.43	33.41± 4.05*
Malondialdehyde (nmol/ml)	1.2± 0.09	0.696± 0.147*
Triglyceride (mg/100 ml)	2.278± 0.054	2.267± 0.126
Total cholesterol (mg/100 ml)	233.766± 25.25	221.591± 1.61
LDL cholesterol (mg/100 ml)	231.7± 25.22	219.397± 1.56
HDL cholesterol (mg/100 ml)	1.611± 0.029	1.741± 0.027*

(\*P≤0.05 vs. control.)

The study of aqueous extract of *Foeniculum vulgare* was used to investigate its anti-diabetic effects. This extract is useful in lowering the blood sugar in streptozotocin induce diabetic rats. The methanolic extract of fruit *F. vulgare* decreased the blood glucose and triglyceride. This medicinal plant can be used in the manufacturing of antidiabetic drugs. The *Foeniculum vulgare* had significant protecting effects on gastrointestinal complaints. The infant with colic disorders was treated with fennel oil emulsions. It showed 65% improvement in infant than the control group. It was also investigated *F. vulgare* had a defensive effect on gastric ulcer. These functions of the plant are present due to its antioxidant capacity [75]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *F. vulgare* has been reported.

## 2.9 *Trachyspermum Ammi*

*Trachyspermum ammi* (*Ptychotis ajowan*) belongs to the family Apiaceae [76]. *Trachyspermum ammi* L. (Apiaceae) commonly known as ajwan. The gas chromatography (GC) and gas chromatography mass spectrometry (GC-MS) investigates the main components of the essential oil which includes p-cymene (26.8%), thymol (43.7%) and  $\gamma$ -terpinene (24.9%) [77]. It is used in the management of gastrointestinal diseases such as diarrhea, dyspepsia and cramp disorders. This medicinal plant had various pharmacological properties such as antifungal, antihypertensive, antitussive and cytotoxic. The medicinal uses are in the treatment of stomach disorders, carminative, antiseptic and antimicrobial [78]. *Trachyspermum ammi* extracts reflect the significant antihyperlipidaemia effect in albino rabbits [79].

Some researchers essential oil of *Trachyspermum ammi* seed used to investigate the antimicrobial activity [80]. Antimicrobial activity was observed with four types of bacteria such as *Salmonella typhi*, *Escherichia*

*coli*, *Lactobacillus* and *Bacillus licheniformis* by agar disc diffusion method. The essential oil indicates superior antimicrobial activity in contrast to different solvent extracts. Antioxidants activity of methanolic and aqueous seed extracts of *Foeniculum Vulgare* and *Trachyspermum ammi* were reported due to presence of high contents level of total phenolic in extracts [11]. The ethanolic and aqueous extracts of seeds of *Trachyspermum ammi* were used to investigate the anthelmintic activities. The effects of extracts were observed in earthworm *P. posthuma* (Annelida) at the dose of 10mg/ml, 20mg/ml and 40mg/ml. The albendazole considered as reference drug. The death of worms and time of paralysis parameters were used to determine anthelmintic effect. The dose dependent anthelmintic activities of seeds of *Trachyspermum ammi* L were observed during this study [81].

*Trachyspermum ammi* seeds has nephroprotective effects. The nephroprotective activity of plant seed aqueous extracts due to the presence of antioxidative polyphenolic compounds. It was concluded that polyphenolic constituent, to inhibit the nephrotoxicity due to oxidative stress and to restore the kidney markers [82]. The traditional used of seed ajwan is to treat of urinary stone. The seed of ajwan were decocted in milk, and then orally administered to the patient suffering from urinary stone. The treatment should be continued for nine days [83]. The anticholinergic activity was observed due to presence of thymol as main constituent of *Trachyspermum ammi* [76]. The traditional used of *Trachyspermum ammi* as stimulant in digestive system. It was recommended to increase gastric acid, bile acids and digestive enzymes secretion level. The effects on the enzymatic system of pancreatic lipase and amylase improve the digestive stimulant activity [83]. Phytochemical qualitative (Table 1 A, B), quantitative (Table 2 A, B) and antioxidant screening (Table 3) of *Trachyspermum ammi* has been reported.

### 3. CONCLUSION

Herbal plants are used as natural remedies throughout the world for the treatment of different diseases because of presence of phytochemical constituents and bioactive compounds likewise phenols, saponin, flavonoids, alkaloids etc. Now a day, scientist's attention is gradually more turning to ethno botanical practices because of less cost, more effectiveness, reliable and no serious side effects of herbal plants for development of new drugs in opposition to very old and advanced diseases

### REFERENCES

- [1] Singh, R. 2015. Medicinal Plants: A Review. Journal Plant Sciences, 3 (1-1), 50-55.
- [2] Shinwari, Z.K. 2010. Medicinal plants research in Pakistan. Journal of Medicinal Plants Research, 4 (3), 161-176.
- [3] Edeoga, H.O., Okwu, D.E., Mbaebie, B.O. 2005. Phytochemical constituents of some Nigerian medicinal plants. African Journal Biotechnology, 4 (7), 685-688.
- [4] Pal, S.K., Shukla, Y. 2003. Herbal Medicine: Current Status and the Future. Asian Pacific Journal of Cancer Prevention, 4 (1), 281-288.
- [5] Valko, M., Rhodes, C.J., Moncola, J., Izakovic, M., Mazura, M. 2006. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chemo Biological Interactions, 160, 1-40.
- [6] Birben, E., Sahiner, U.M., Sackesen, C., Erzurum, S., Kalayci, O. 2012. Oxidative stress and antioxidant defense. World Allergy Organization Journal, 5, 9-19.
- [7] Suijth, K., Darwin, C.R., Suba, V. 2011. Antioxidant activity of ethanolic root extract of anacyclus pyrethrum. International Research Journal of Pharmacy, 2 (12), 222-2260.
- [8] Usmani, A., Khushtar, M., Arif, M., Siddiqui, M.A., Sing, S.P., Mujahid, M. 2016. Pharmacognostic and phytopharmacology study of *Anacyclus pyrethrum*: An insight. Journal of Applied Pharmaceutical Science, 6 (03), 144-150.
- [9] Elazzouzi, H., Aminata, S., Fatima, E., Amar, B., Belghiti, M.A.E., Touriya, Z. 2014. Phytochemical study of *Anacyclus pyrethrum* (L.) of Middle Atlas (Morocco), and *in vitro* study of antibacterial activity of pyrethrum. International Journal of Natural and Applied Sciences, 8 (8), 131-140.
- [10] Annalakshmi, R., Uma, R., Chandran, G.S., Muneeswaran, A. 2012. A treasure of medicinal herb - *Anacyclus pyrethrum* a review. Indian Journal of Drugs and Diseases, 1 (3), 59-67.
- [11] Chatterjee, S., Goswami, N., Bhatagar, P. 2012. Estimation of phenolic components and *in vitro* antioxidant activity of Fennel (*Foeniculum vulgare*) and Ajwain (*Trachyspermum ammi*) seeds. Advances in Biological Research, 3 (2), 109-118.
- [12] Latifa, C., Meddah, B., Alnamer, R., Chibani, F., Cherrrah, Y. 2012. *In vitro* antibacterial activity of the methanolic and aqueous extract of anacyclus pyrethrum used in Moroccan traditional medicine. International Journal of Pharmacy and Pharmaceutical Sciences, 4 (3), 402-405.
- [13] Selles, C., Dib, M.E.A., Allali, H., Tabti, B. 2012. Evaluation of antimicrobial and antioxidant activities of solvent extracts of *Anacyclus pyrethrum* L., from Algeria. Mediterranean Journal of Chemistry, 2 (2), 408-415.
- [14] Shahraki, S., Rad, J.S., Rostami, F.M., Shahraki, M.R., Arab, M.R. 2014. Effects of aqueous root extracts of *Anacyclus Pyrethrum* on gonadotropins and testosterone serum in adult male rats. American Journal of Phytomedicine and Clinical Therapeutics, 2 (6), 767-772.
- [15] Sharma, S., Dwivedi, J., Paliwal, S. 2012. Evaluation of antacid and carminative properties of *Cucumis sativus* under simulated conditions. Der Pharmacia Lettre, 4 (1), 234-239.
- [16] Badhe, S.R., Badhe, R.V., Ghaisas, M.M., Chopade, V.V., Deshpande, A.D. 2012. Evaluations of antidepressant activity of *Anacyclus pyrethrum* root extract. International Journal of Green Pharmacy, 1 (1), 79-82.
- [17] Rani, S., Kaushik, V., Saini, V., Nain, P., Rani, D. 2013. Biological studies of *Anacyclus Pyrethrum*. Indo American Journal of Pharmaceutical Research, 3 (6), 4590-4596.
- [18] Shanmugapriya, R., Ushadevi, T. 2015. Phytochemical screening and GC-MS analysis of *Apium graveolens* L. seed extracts. International Journal of Pharma and Bio Sciences, 6 (2), 814 - 820.
- [19] Fazal, S.S., Single, R.K. 2012. Review on the pharmacognostical and pharmacological Characterization of *Apium Graveolens* Linn. Indo Glob. Journal of Pharmaceutical Sciences, 2 (1), 36-42.
- [20] Danboyina, D.A., Mawak, J., Egwim, E.C., Auta, Y., Wunzani, D.K., Ghaji, J.A. 2014. A study on the efficacy of extracts of *Boerhavia diffusa* L on bacterial isolates of fingertip infections (whitlow). International Journal of Basic, Applied and Innovative Research, 3 (4), 125-136.
- [21] Gupta, B.K., Tailang, M., Gavatia, N.P. 2012. A systematic study and profiling of leaves of *Cinnamomum cassia* Blum for its pharmacognostical and phytochemical investigation. Journal of Pharmaceutical Research, 5 (2), 814-816.
- [22] Ahmad, M.I., Ansari, S.H., Naquvi, K.J., Shuaib, M. 2012. Quality standards of fruit of *Cucumis sativus* Linn. Journal of Pharmacy Research, 5 (1), 22-25.
- [23] Fahamiya, N., Aslam, M., Siddiqui, A., Shiffa, M., Ahmed, A., Khan, M.S. 2012. Pharmacognostical Study and Development of Quality Control Parameters for *Cucumis melo* Linn. American Journal of PharmTech Research, 2 (4), 510-523.
- [24] Babulreddy, N., Sahoo, S.P., Ramachandran, S., Dhanaraju, M.D. 2013. Anti-hyperglycemic activity of *cucumis melo* leaf extracts in streptozotocin induced hyperglycemia in rats. International Journal of Pharmaceutical Research and Allied Sciences, 2 (4), 22-27.
- [25] Malik, J., Akhter, R. 2012. Phytochemical Screening and *In-vitro* Evaluation of Reducing Power, Cytotoxicity and Anti-Fungal Activities of Ethanolic Extracts of *Cucumis sativus*. Journal of Pharmaceutical Biology, 3 (3), 555-560.
- [26] Senthil, V., Ramasamy, P., Elaiyaraja, C., Elizabeth, A.R. 2010. Some phytochemical properties affected by the infection of leaf spot disease of *Cucumis Sativus* (Linnaeus) caused by *Penicillium notatum*. African Journal of Basic & Applied Sciences, 2 (3-4), 64-70.
- [27] Patil, M.V.K., Kandhare, A.D., Bhise, S.D. 2012. Pharmacological evaluation of ethanolic extract of *Daucus carota* Linn root formulated cream on wound healing using excision and incision wound model. Asian Pacific Journal Tropical Biomedicine, 1 (1), 646-655.
- [28] Jamwal, N.S., Kumar, S., Rana, A.C. 2013. Phytochemical and pharmacological review on *Foeniculum vulgare*. International Journal of Pharmaceutical Sciences and Research, 4 (3), 327-341.
- [29] Bajpai, V.K., Agrawal, P. 2015. Studies on phytochemicals, antioxidant, free radical scavenging and lipid peroxidation inhibitory effects of *Trachyspermum ammi* seeds. Indian Journal of Pharmaceutical Education and Research, 49 (1), 58-65.
- [30] Kooti, W., Ghasemiboroon, M., Samani, M.A., Ahangarpour, A., Zamani, M., Amirzaegar, A., Hardani, A. 2014. The effect of halcoholic extract of Celery leaves on the delivery rate (fertilization and stillbirths), the number, weight and sex ratio of rat off spring. Advances in Environmental

Biology, 8 (10), 824-830.

[31] Ujowundu, C.O., Igwe, C.U., Enemor, V.H.A., Nwaogu, L.A., Okafor, O.E. 2008. Nutritive and anti-nutritive properties of *Boerhavia diffusa* and *Commelina nudiflora* leaves. Pakistan Journal of Nutrition, 7 (1), 90-92.

[32] Yang, C.H., Chang, H.W., Line, J.Y., Chuang, L.Y. 2013. Evaluation of Antioxidant and Antimicrobial Activities from 28 Chinese Herbal Medicines. Journal of Pharmacognosy and Phytochemistry, 2 (1), 294-305.

[33] Budhirajaa, H., Guptaa, R.K., Nand, P. 2014. Formulation and characterization of *cucumis sativus* extract in the treatment of acne. World Journal of Pharmaceutical Sciences, 3 (12), 143-1057.

[34] Ksouri, A., Dob, T., Belkebir, A., Krinat, S., Chelghoum, C. 2015. Chemical composition and antioxidant activity of the essential oil and the methanol extract of Algerian wild carrot *Daucus carota* L. ssp. *carota*. (L.) Thell. Journal of Materials and Environmental Science, 6 (3), 784-791.

[35] Garg, C.D., Ansari, P.S.H., Khan, P.S.A., Garg, D.M. 2011. Effect of *Foeniculum vulgare* mill fruits in obesity and associated cardiovascular disorders demonstrated in high fat diet fed albino rats. Journal of Pharmaceutical and Biomedical Sciences, 8 (08), 1-5.

[36] Dwivedi, S.N., Mishra, R.P., Alava, S. 2012. Phytochemistry, pharmacological studies and traditional benefits of *Trachyspermum ammi* (Linn.) sprague. International Journal of Pharmacy and Life Sciences, 3 (5), 1705-1709.

[37] Din, Z.U., Shadi, A.A., Bakht, J., Ullah, I., Jan, S. 2015. *In vitro* antimicrobial, antioxidant activity and phytochemical screening of *Apium graveolens*. Pakistan Journal of Pharmaceutical Sciences, 5 (28), 1699-1704.

[38] Beegum, G.R.J., Beevy, S.S., Sugunan, V.S. 2016. Natural antioxidant activity of *Boerhavia diffusa* L. International Journal of Pharmacognosy and Phytochemical Research, 8 (1), 8-13.

[39] Prasad, N.K., Yang, B., Dong, X., Jiang, G., Zhang, H., Xie, H., Jiang, Y. 2009. Flavonoid contents and antioxidant activities from *Cinnamomum* species. Innovative Food Science and Emerging Technologies, 10 (1), 627-632.

[40] Krishna, T.M., Shiva, D., Keerthi, D., Ahmed, A., Reddy, V.B., Kumar, A. 2013. *In vitro* evaluation of anti-oxidant properties of *Cucumis melo* L. extracts of leaves and fruit. International Journal of Pharma and Bio Sciences, 4 (1), 705 - 712.

[41] Nasrin, F., Bulbul, I.J., Aktar, F., Rashid, M.A. 2015. Anti-inflammatory and antioxidant activities of *cucumis sativus* leaves. Bangladesh Pharmaceutical Journal, 18 (2), 169-173.

[42] Anwara, F., Alia, M., Hussaina, A.I., Shahid, M. 2009. Antioxidant and antimicrobial activities of essential oil and extracts of fennel (*Foeniculum vulgare* Mill.) seeds from Pakistan. Flavour and Fragrance Journal, 24, 170-176.

[43] Tyagil, S.P., Chirag, P., Dhruv, M., Ishita, M., Gupta, A.K., Usman, M.R.M., Nimbiwal, B., Maheshwari, D.R.K. 2013. Medical benefits of *apium graveolens* (celery herb). Journal of Drug Discovery and Therapeutics, 1 (5), 36-38.

[44] Kooti, W., Akbari, S.A., Samani, M.A., Ghadery, H., Larky, D.H. 2014. A review on medicinal plant of *Apium graveolens*. Advanced Herbal Medicine, 1 (1), 48-59.

[45] Al-Snafi, A.E. 2014. The Pharmacology of *Apium graveolens*. A Review. International Journal for Pharmaceutical Research Scholars, 3 (1), 671-677.

[46] Khalid, M., Siddiqui, H.H., Freed, S. 2012. Pharmacognostical evaluation and qualitative analysis of *Boerhaavia diffusa* roots. International Journal

of Pharma and Bio Sciences, 3 (1), 16-23.

[47] Mahesh, A.R., Kumar, H., Ranganath, M.K., Devkar, R.A. 2012. Detail Study on *Boerhaavia Diffusa* Plant for its Medicinal Importance- A Review. Research Journal of Pharmaceutical Sciences, 1 (1), 28-36.

[48] Bhowmik, D., Kumar, K.P.S., Srivastava, S., Paswan, S., Sankar, A., Dutta, D. 2012. Traditional Indian herbs *Punarnava* and its medicine importance. Journal of Pharmacognosy and Phytochemistry, 1 (1), 52-57.

[49] Deepti, M., Khan, A., Ishaq, F. 2013. Phytochemical screening and antibacterial effect of root extract of *Boerhaavia diffusa* L. (Family Nyctaginaceae). Journal of Applied and Natural Science, 5 (1), 221-225.

[50] Riaz, H., Raza, S.A., Hussain, S., Mahmood, S., Malik, F. 2014. An overview of ethnopharmacological properties of *Boerhaavia diffusa*. African Journal of Pharmacy and Pharmacology, 8 (2), 49-58.

[51] Zaidi, F.S., Aziz, M., Muhammad, J.S., Kadowaki, M. 2015. Diverse pharmacological properties of *Cinnamomum cassia*: A review. Pakistan Journal of Pharmaceutical Sciences, 28 (4), 1433-1438.

[52] Rao, P.V., Gan, S.H. 2014. Cinnamon: A Multifaceted Medicinal Plant. Evid. Based. Complement. Alternat. Med. Article ID 642942, 1-12.

[53] Ervina, M., Nawu, Y.E., Esar, S.Y. 2016. Comparison of *in vitro* antioxidant activity of infusion, extract and fractions of Indonesian Cinnamon (*Cinnamomum burmannii*) bark. International Food Research Journal, 23 (3), 1346-1350.

[54] Adinew, B. 2014. GC-MS and FT-IR analysis of constituents of essential oil from *Cinnamon* bark growing in South-west of Ethiopia. International Journal of Herbal Medicine, 1 (6), 22-31.

[55] Tabassum, H., Ali, M.N., Jameil, N.A., Khan, F.A. 2013. Evaluation of antibacterial potential of selected plant extracts on bacterial pathogens isolated from urinary tract infections. International Journal of Current Microbiology and Applied Sciences, 2 (10), 353-368.

[56] Hamad, M.I., Salman, A.M.A., Salman, W.A.M.A., Abdelkhalig, E.M. 2015. Effect of *cinnamomum cassia* on lipid profile of apparently healthy subjects. Journal Forest Products Industries, 4 (4), 144-149.

[57] Vijikumar, S., Ramanathan, K., Devi, B.P. 2011. *Cuscuta reflexa* roxb- A wonderful miracle plant in ethnomedicine. Indian Journal of Natural Sciences, 11 (9), 676-683.

[58] Vouldoukis, I., Lacan, D., Kamate, C., Coste, P., Calenda, A., Mazier, D., Conti, M., Dugasa, B. 2004. Antioxidant and anti-inflammatory properties of a *Cucumis melo* LC. extract rich in superoxide dismutase activity. Journal of Ethnopharmacology, 94 (1), 67-75.

[59] Bidkar, J.S., Ghanwat, D.D., Bhujbal, M.D., Dama, G.Y. 2012. Anti-hyperlipidemic activity of *Cucumis melo* fruit peel extracts in high cholesterol diet induced hyperlipidemia in rats. Journal Complementary Integrative Medicine, 9 (1).

[60] Ravishankar, K., Priya, P.S.V.V. 2012. Evaluation of diuretic effect of ethanolic seed extracts of *macrotyloma uniflorum* and *cucumis melo* in rats. International Journal of Pharma and Bio Sciences, 3 (3), 251- 255.

[61] Malik, J., Das, P., Das, S. 2013. Pharmacological activity of *Cucumis sativus* L. - A complete overview. Asian Journal of Pharmaceutical Research and Development, 1 (1), 2-6.

[62] Pradhan, D., Biswasroy, P., Singh, G., Suri, K.A. 2013. Anti-ulcerogenic activity of ethanolic extract of *cucumis sativus* l. against nsaid (aspirin) induced gastric ulcer in wistar albino rats. International Journal of Herbal Medicine, 1 (3), 115-119.

[63] Sharmin, R., Khan, M.R.I., Akhter, M.A., Alim, A., Islam, M.A.,

- Anisuzzaman, A.S.M., Ahmed, M. 2013. Hypoglycemic and hypolipidemic effects of cucumber, white pumpkin and ridge gourd in alloxan induced diabetic rats. *Journal of Scientific Research*, 5 (1), 161-170.
- [64] Chatatikum, M., Chiabachalard, A. 2013. Phytochemical screening and free radical scavenging activities of orange baby carrot and carrot (*Daucus carota* Linn.) root crude extracts. *Journal of Chemical and Pharmaceutical Research*, 5 (4), 97-102.
- [65] Khatib, N., Angle, G., Naynab, H., Kumare, J.R. 2010. Gastroprotective activity of the aqueous of the aqueous extract from the roots of *Daucus carota* L in rats. *International Journal of Research in Ayurveda and Pharmacy*, 1 (1), 112-119.
- [66] Vishwanath, M.P., Kandhare, A.D., Bhaise, S.D. 2012. Pharmacological evaluation of ethanolic extract of *Daucus carota* Linn root formulated cream on wound healing using excision and incision wound model. *Asian Pacific Journal of Tropical Biomedicine*, 646-655.
- [67] Manil, V., Parle, M., Ramasamy, K., Majeed, A.B.A. 2010. Anti-dementia potential of *Daucus carota* seed extract in rats. *Journal of Pharmacology and Pharmacotherapeutics*, 1 (1), 52-565.
- [68] Singh, K., Dhogade, H., Singh, N., Kashyap, P. 2010. Hypolipidemic activity of ethanolic extract of *Daucus carota* seeds in normal rats. *International Journal of Biomedical and Advance Research*, 1 (03), 73-80.
- [69] Abdossi, V., Ghahremani, A., Hadipanah, A., Ardalani, H., Aghaee, K. 2015. Quantitative and qualitative responses in chemical composition of three ecotypes of fennel (*Foeniculum vulgare* Mill) cultivated in Iran climatic conditions. *Journal of Biodiversity and Environmental Sciences*, 6 (3), 401-407.
- [70] Badgular, S.B., Patel, V.V., Bandivdekar, A.H. 2014. *Foeniculum vulgare* Mill: A Review of Its Botany, Phytochemistry, Pharmacology, Contemporary Application and Toxicology. *BioMed Research International*, Article ID 842674, 1-32.
- [71] Khazaei, M., Montaser, A., Khazaei, M.R., Khanahmadi, M. 2011. Study of *Foeniculum vulgare* effect on folliculogenesis in female mice. *International Journal of Fertility & Sterility*, 5 (3), 122-127.
- [72] Sadrefozalayi, S., Farokhi, F. 2014. Effect of the aqueous extract of *Foeniculum vulgare* (fennel) on the kidney in experimental PCOS female rats. *Avicenna Journal of Phytomedicine*, 4 (2), 110-117.
- [73] Elizabeth, A.A., Josephine, G., Muthiah, N.S., Muniappan, M. 2014. Evaluation of analgesic and anti-inflammatory effect of *Foeniculum vulgare*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 5 (2), 658-668.
- [74] Choi, E.M., Hwang, J.K. 2004. Antiinflammatory, analgesic and antioxidant activities of the fruit of *Foeniculum vulgare*. *Fitoterapia Journal*, 75 (1), 557-565.
- [75] Kooti, W., Moradil, M., Akbari, S.A., Ahyazi, N.S., Samani, M.A., Larky, D.A. 2015. **Therapeutic and pharmacological potential of *Foeniculum vulgare* Mill: a review.** *Journal of Herbmed Pharmacology*, 4 (1), 1-9.
- [76] Hejazian, S.H., Bagheri, S.M., Safari, F. 2014. Spasmolytic and anti-spasmodic action of *Trachyspermum ammi* essence on rat's ileum contraction. *American Journal of the Medical Sciences*, 6 (12), 643-647.
- [77] Nickavara, B., Adelia, A., Nickavar, A. 2014. TLC-bioautography and GC-MS analyses for detection and identification of antioxidant constituents of *Trachyspermum copticum* essential oil. *Iranian Journal of Pharmaceutical Research*, 13 (1), 127-133.
- [78] Jeet, K., Davi, N., Narender, T., Sunil, T., Lalit, S., Raneev, T. 2012. *Trachyspermum ammi* (ajwain): A comprehensive review. *International Research Journal of Pharmacy*, 3 (5), 133-138.
- [79] Javed, I., Iqbal, Z., Rahman, Z.U., Khan, F.H., Muhammad, F., Aslam, B., Ali, L. 2006. Comparative antihyperlipidaemic efficacy of *Trachyspermum ammi* extract in albino rabbits. *Pakistan Veterinary Journal*, 26 (1), 23-29.
- [80] Aggarwal, S., Goyal, S. 2012. In vitro antimicrobial studies of *Trachyspermum ammi*. *International Journal of Pharma and Bio Science*, 3 (4), 64 - 68.
- [81] Apte, A.K., Khot, V.S., Biradar, N.S., Patil, S.B. 2014. Anthelmintic activity of *Trachyspermum ammi* (L) extract. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6 (2), 236-238.
- [82] Ishaq, B., Khan, J.A., Murtaza, S., Abbas, R.Z., Khaliq, T., Khan, A., Arshad, H.A., Anwar, H. 2015. Protective potential of *Trachyspermum ammi* seeds in gentamicin-induced nephrotoxicity in rabbit model. *Bol. Latinoam Caribe Plant Med*, 14 (4), 280-286.
- [83] Zarshenas, M.M., Moein, M., Samani, S.M., Petramfar, P. 2014. An overview on ajwain (*Trachyspermum ammi*) pharmacological effects; Modern and Traditional. *Journal of Natural Remedies*, 14 (1), 98-106.

