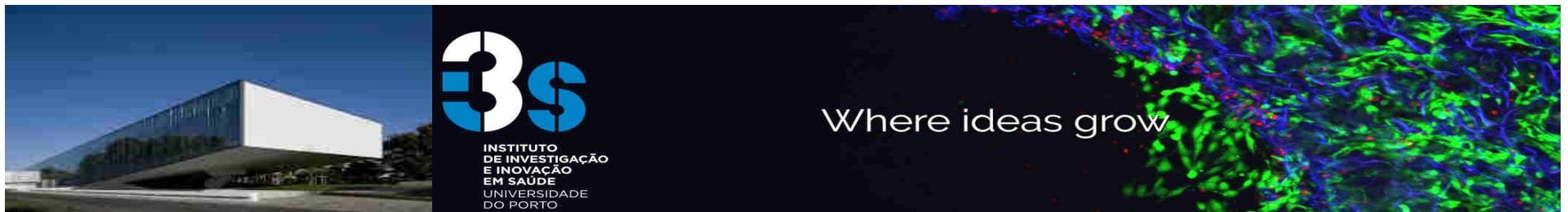




Molecular Pathology of Gastric Cancer

Novel insights and possible future applications

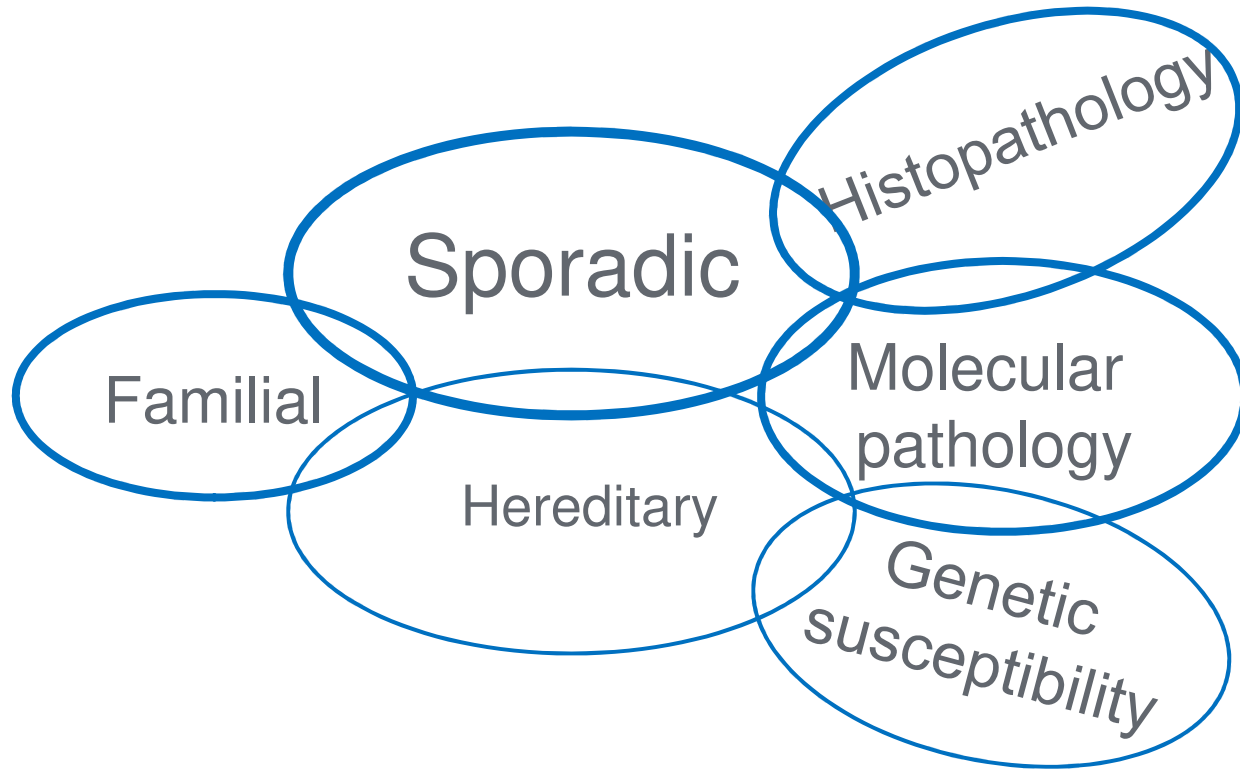
Fátima Carneiro
i3S/Ipatimup & Medical Faculty/ CHS João
Porto, Portugal



Outline

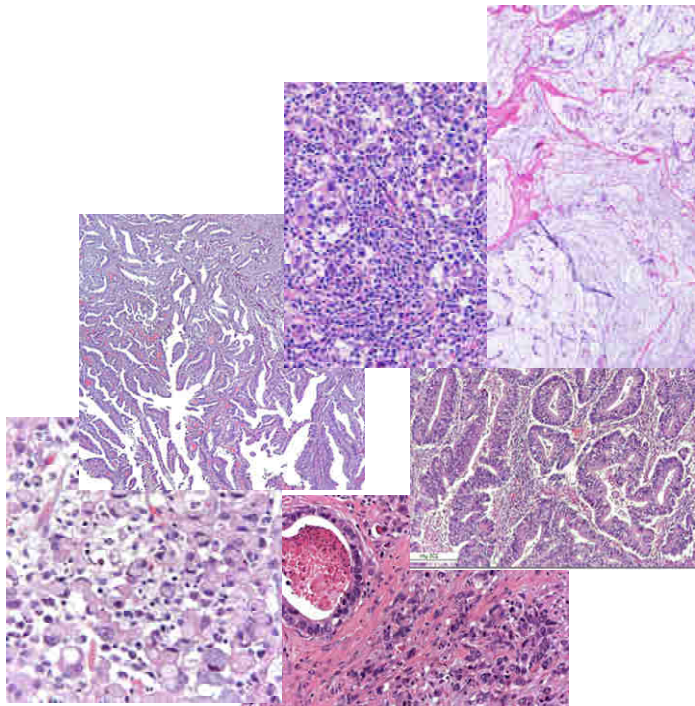
1. Major histological types of gastric cancer and the variants with clinical relevance (WHO 2018)
2. Major molecular classifications of gastric cancer
3. Molecular targets for therapy

Gastric Cancer

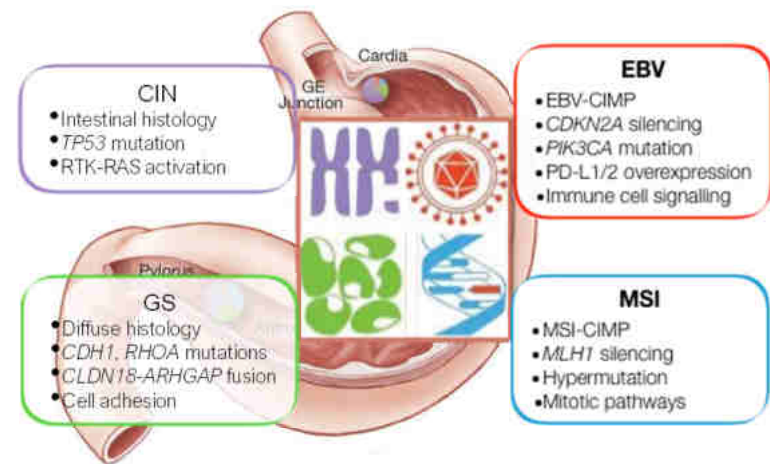


Gastric cancer

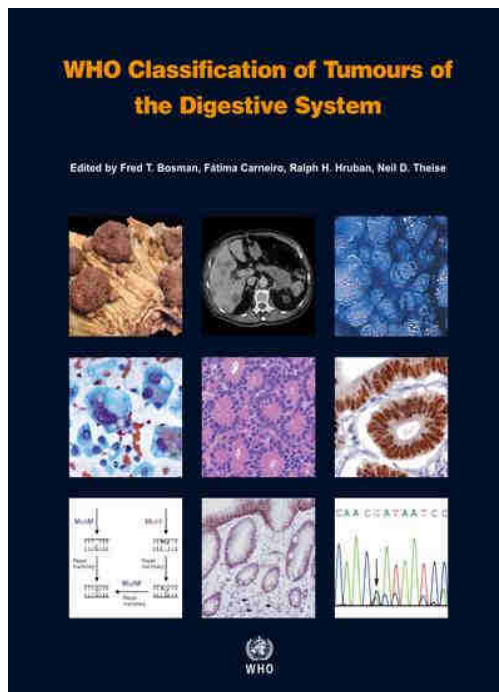
Morphological heterogeneity



Molecular heterogeneity



WHO Classification of Tumours of the Digestive System, 4th edition, 2010

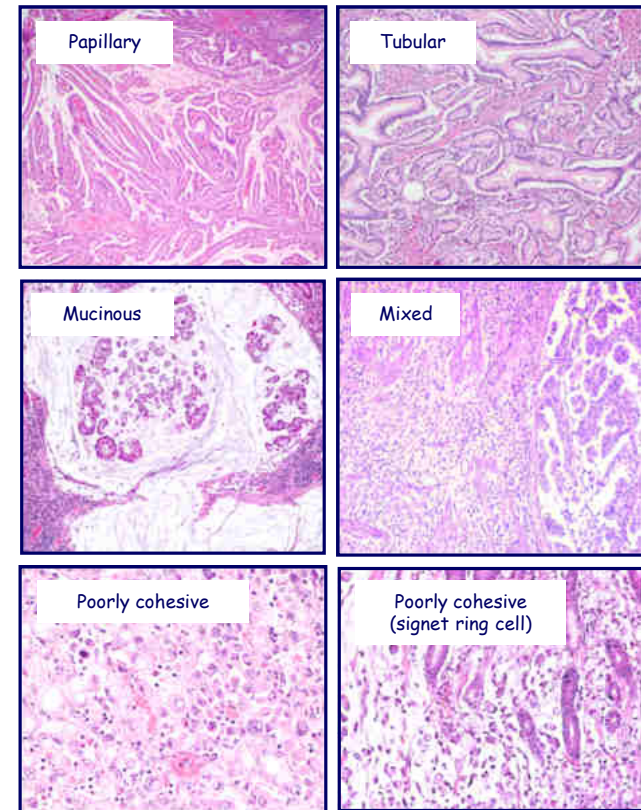


WHO Classification of Gastric Carcinoma, 4th edition, 2010

Gregory Y. Lauwers
Fátima Carneiro
David Y. Graham
Maria-Paula Curado
Silvia Franceschi
Elizabeth Montgomery
Masae Tatematsu
Takenori Hattori

ICD-O Code

Adenocarcinoma	8140/3
Papillary adenocarcinoma	8260/3
Tubular adenocarcinoma	8211/3
Mucinous adenocarcinoma	8480/3
Poorly cohesive carcinoma (Signet-ring cell carcinoma and variants)	8490/3
Mixed carcinoma	8255/3



WHO-5th Edition –Editorial board



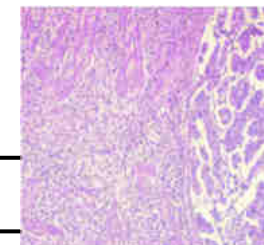
Histological classifications of gastric cancer



WHO, 5th edition

Laurén (1965)	Nakamura (1968)	JGCA (2017)	WHO (2018)
Intestinal	Differentiated	Papillary: pap Tubular 1, well-differentiated: tub1 Tubular 2, moderately-differentiated: tub2	Papillary Tubular, well-differentiated Tubular, moderately-differentiated
Indeterminate	Undifferentiated	Poorly 1 (solid type): por 1	Tubular, poorly-differentiated (solid)
Diffuse	Undifferentiated	Signet ring cell carcinoma (SRC): sig Poorly 2 (non-solid type): por2	Poorly cohesive, SRC type Poorly cohesive, NOS
Intestinal/diffuse/indeterminate	Differentiated/ Undifferentiated	Mucinous	Mucinous
Mixed		Description according to the proportion (e.g. por2>sig>tub2)	Mixed
Not defined	Not defined	Special type: Adenosquamous carcinoma Squamous cell carcinoma Undifferentiated carcinoma Carcinoma with lymphoid stroma Hepatoid adenocarcinoma Adenocarcinoma with enteroblastic differentiation Adenocarcinoma of fundic gland type	Histological variants: Adenosquamous carcinoma Squamous cell carcinoma Undifferentiated carcinoma Carcinoma with lymphoid stroma Hepatoid carcinoma Adenocarcinoma with enteroblastic differentiation Adenocarcinoma of fundic gland type Micropapillary adenocarcinoma
*JGCA, Japanese Gastric Cancer Association {978-4-307-20375-3}. **Table prepared in collaboration with Prof. Ryoji Kushima, Japan			

Histological classifications of gastric cancer



Laurén (1965)	Nakamura (1968)	JGCA (2017)	WHO (2018)
Intestinal	Differentiated	Papillary: pap Tubular 1, well-differentiated: tub1 Tubular 2, moderately-differentiated: tub2	Papillary Tubular, well-differentiated Tubular, moderately-differentiated
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*JGCA, Japanese Gastric Cancer Association {978-4-307-20375-3}. **Table prepared in collaboration with Prof. Ryoji Kushima, Japan			

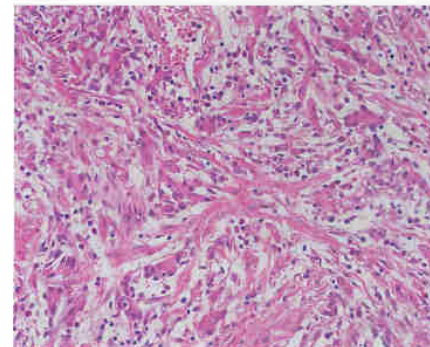
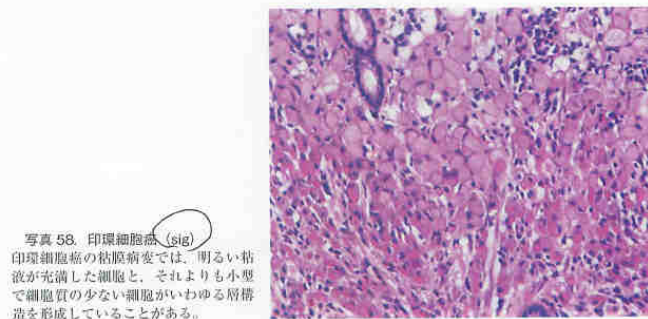
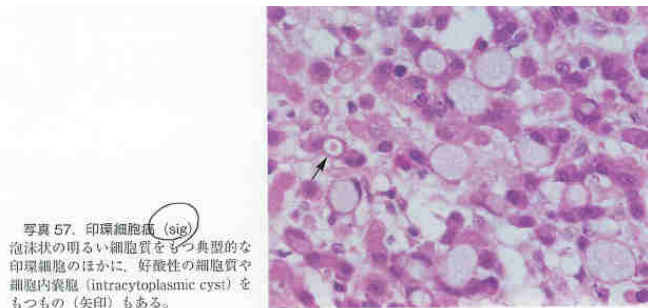
Histological classifications of gastric cancer



WHO, 5th edition

Laurén (1965)	Nakamura (1968)	JGCA (2017)	WHO (2018)
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*JGCA, Japanese Gastric Cancer Association {978-4-307-20375-3}. **Table prepared in collaboration with Prof. Ryoji Kushima, Japan			

JGCA, Japanese Gastric Cancer Association (2017)



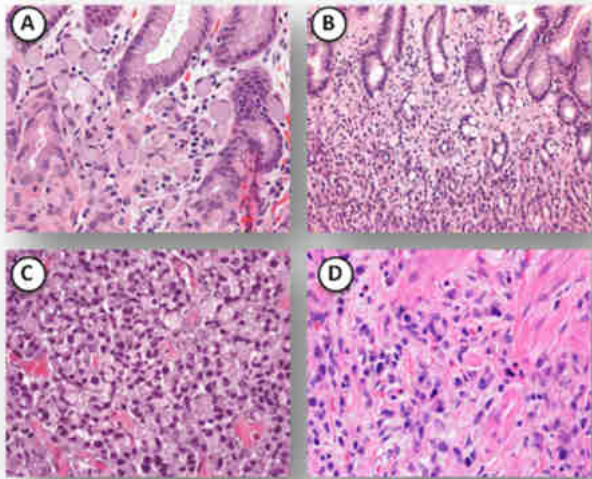
↓ WHO classification (2018) ↓

Poorly cohesive carcinoma, SRC

Poorly cohesive carcinoma, NOS

Poorly cohesive carcinoma: mutational signatures

Heterogeneity of poorly cohesive carcinoma



PCC-NOS :
TP53, BRAF, PI3CA,
SMAD4, and RHOA

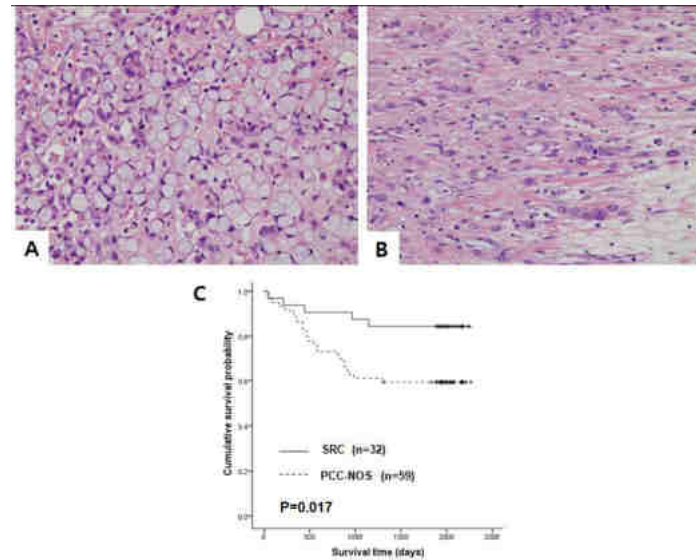
Histopathology



Histopathology 2018, 72, 556–568. DOI: 10.1111/his.13383

Gastric poorly cohesive carcinoma: a correlative study of mutational signatures and prognostic significance based on histopathological subtypes

Chae H Kwon,^{1,2} Young K Kim,^{1,2} Sojeong Lee,^{1,2} Ahrong Kim,^{1,2} Hye J Park,^{1,2} Yuri Choi,^{1,2} Yeo I Won,^{1,2} Do Y Park^{1,2} & Gregor Y Lauwers³



Consensus on the pathological definition and classification of poorly cohesive gastric carcinoma

Gastric Cancer
<https://doi.org/10.1007/s10120-018-0868-0>

SPECIAL ARTICLE



Consensus on the pathological definition and classification of poorly cohesive gastric carcinoma

C. Mariette¹ · F. Carneiro² · H. I. Grabsch^{3,4} · R. S. van der Post⁵ · W. Allum⁶ · Giovanni de Manzoni⁷ on behalf of
European Chapter of International Gastric Cancer Association

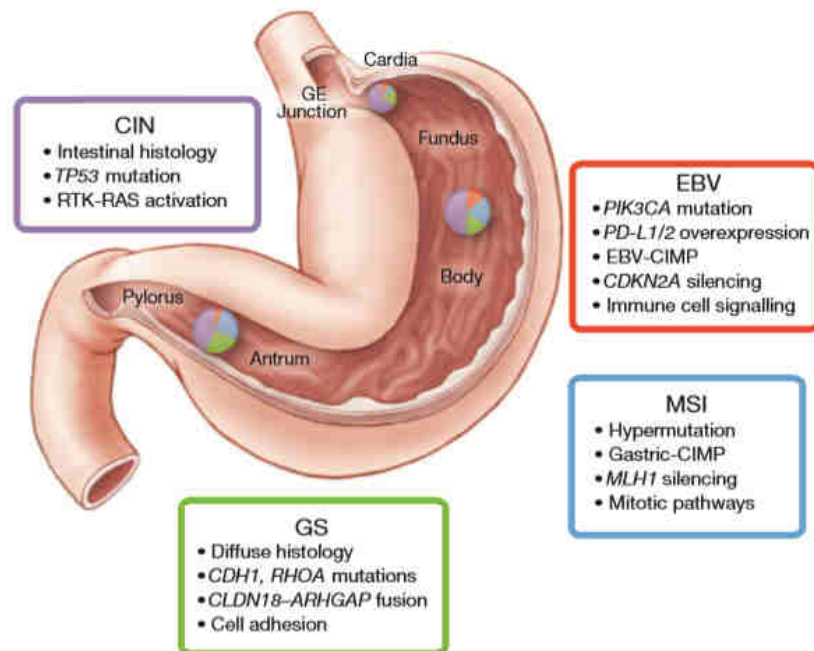
Poorly-Cohesive Carcinoma

- SRC (>90%)
- SRC/NOS (10% - 90%)
- NOS (<10%)

Histological classifications of gastric cancer

Laurén (1965)	Nakamura (1968)	JGCA (2017)	WHO (2018)
Intestinal	Differentiated	Papillary: pap Tubular 1, well-differentiated: tub1 Tubular 2, moderately-differentiated: tub2	Papillary Tubular, well-differentiated Tubular, moderately-differentiated
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Diffuse	Undifferentiated	Signet ring cell carcinoma (SRC): sig Poorly 2 (non-solid type): por2	Poorly cohesive, SRC phenotype Poorly cohesive, other cell types
Intestinal/diffuse/indeterminate	Differentiated/ Undifferentiated	Mucinous	Mucinous
Mixed		Description according to the proportion (e.g. por2>sig>tub2)	Mixed
Not defined	Not defined	Special type: Adenosquamous carcinoma Squamous cell carcinoma Undifferentiated carcinoma Carcinoma with lymphoid stroma Hepatoid adenocarcinoma Adenocarcinoma with enteroblastic differentiation Adenocarcinoma of fundic gland type	Histological variants: Adenosquamous carcinoma Squamous cell carcinoma Undifferentiated carcinoma Carcinoma with lymphoid stroma Hepatoid carcinoma Adenocarcinoma with enteroblastic differentiation Adenocarcinoma of fundic gland type Micropapillary adenocarcinoma
*JGCA, Japanese Gastric Cancer Association {978-4-307-20375-3}. **Table prepared in collaboration with Prof. Ryoji Kushima, Japan			

Molecular classification of gastric cancer (TCGA)



ARTICLE

OPEN
doi:10.1038/nature13480

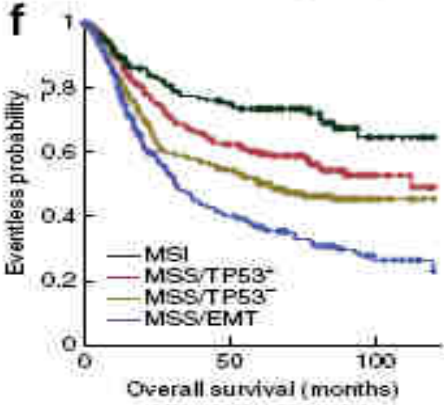
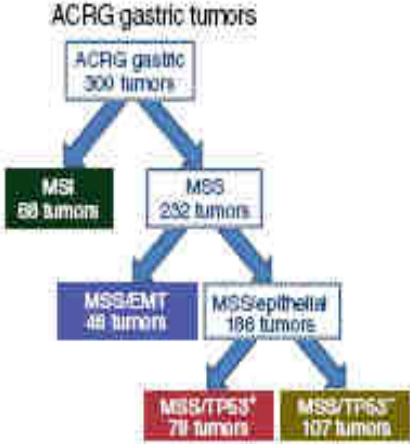
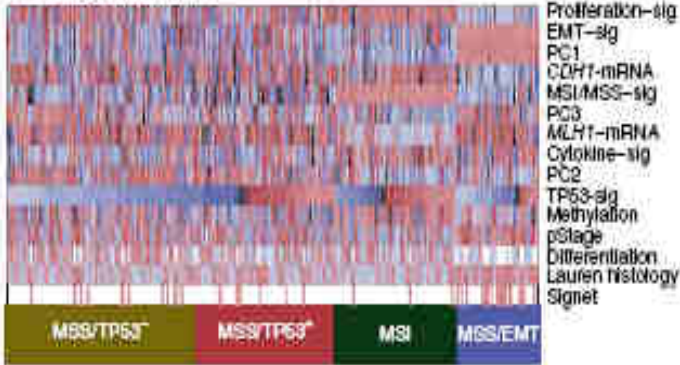
Comprehensive molecular characterization of gastric adenocarcinoma

The Cancer Genome Atlas Research Network*

The Cancer Genome Atlas (TCGA) project;
Nature 2014

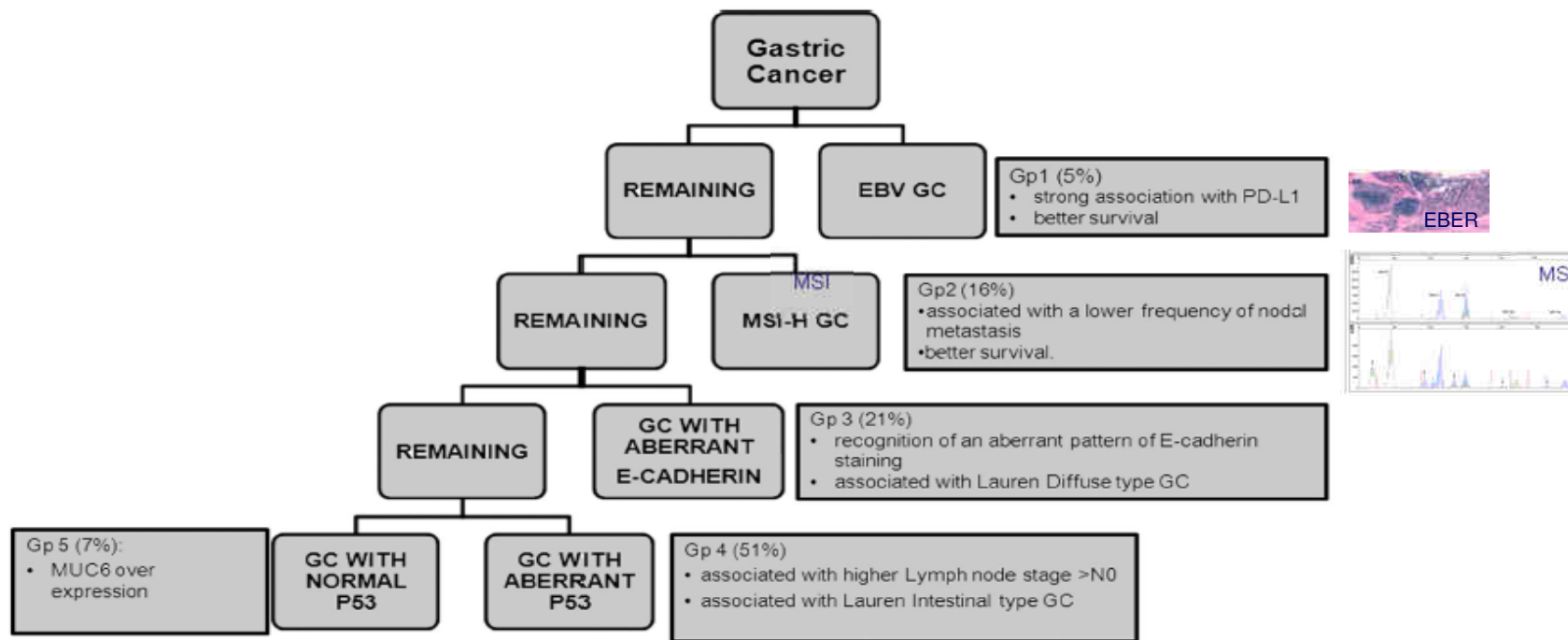
Molecular classification of gastric cancer (ACRG)

ACRG gastric tumors



Asian Cancer Research Group.
Cristescu R *et al*: Nature Medicine 21; 449, 2015

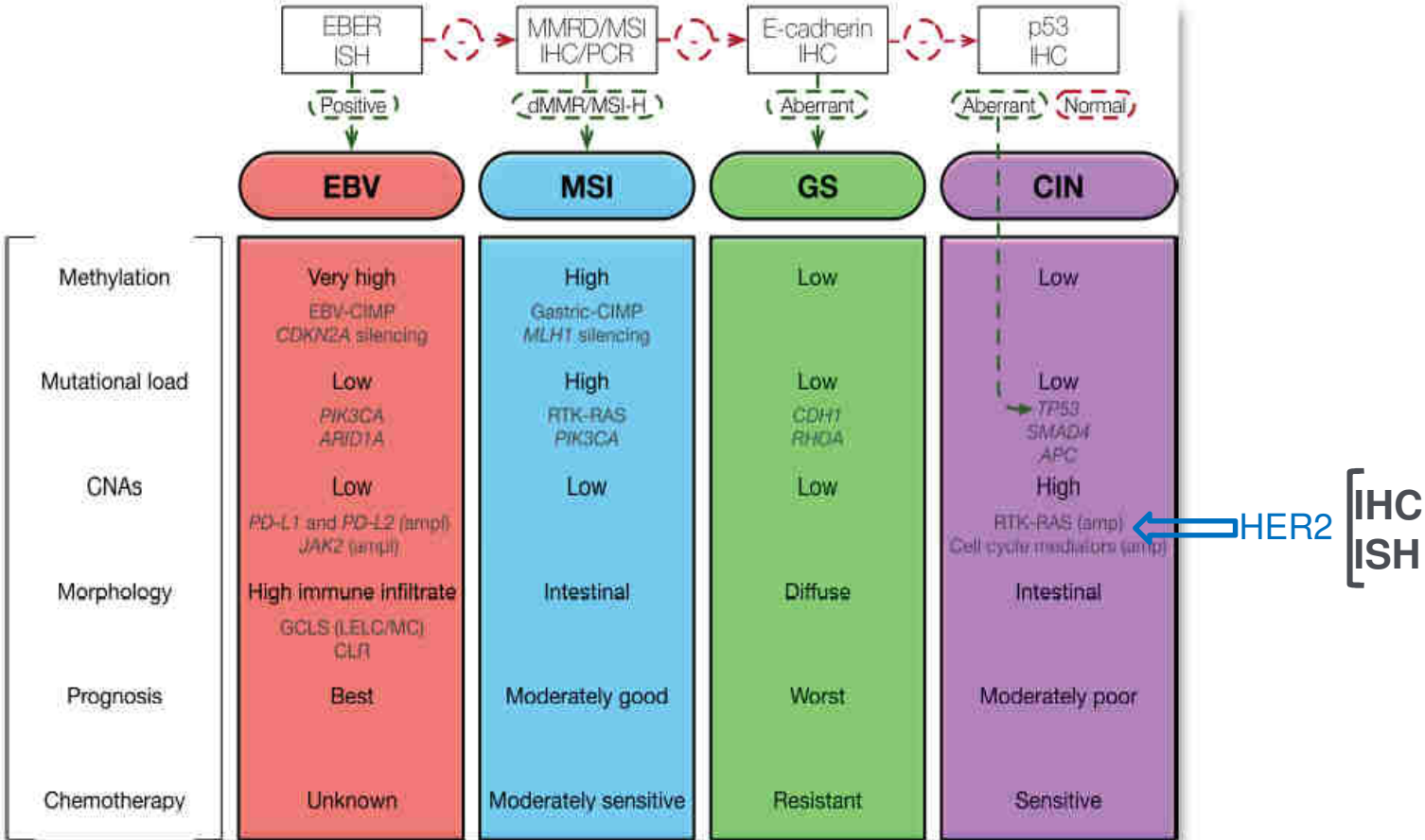
A protein and mRNA expression-based classification of gastric cancer



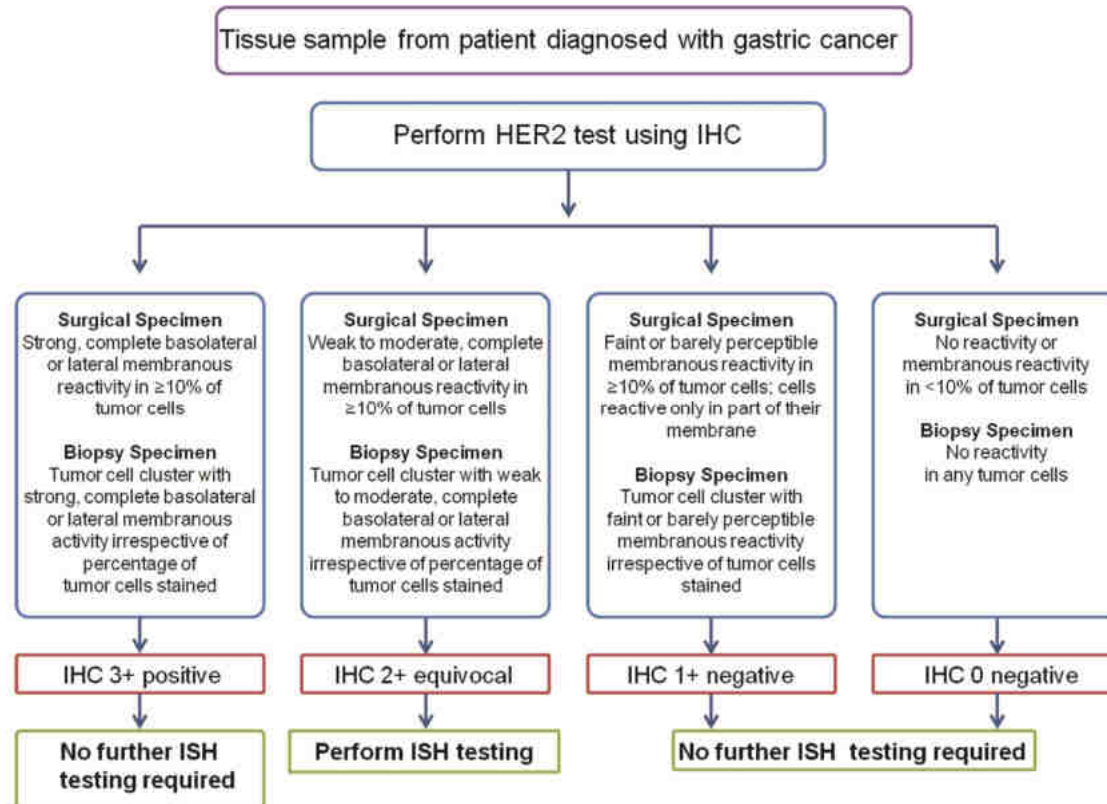
Setia M *et al.* Mod Pathol 29:772, 2016

Ahn S *et al.* Am J Surg Pathol 41:106, 2017

Molecular classification of gastric cancer

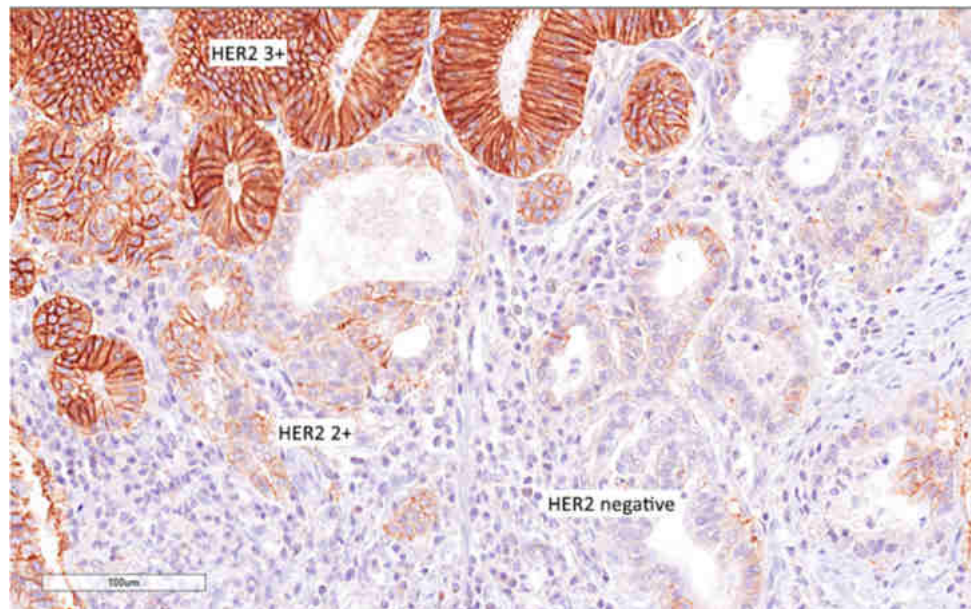


Evaluation of HER2 status

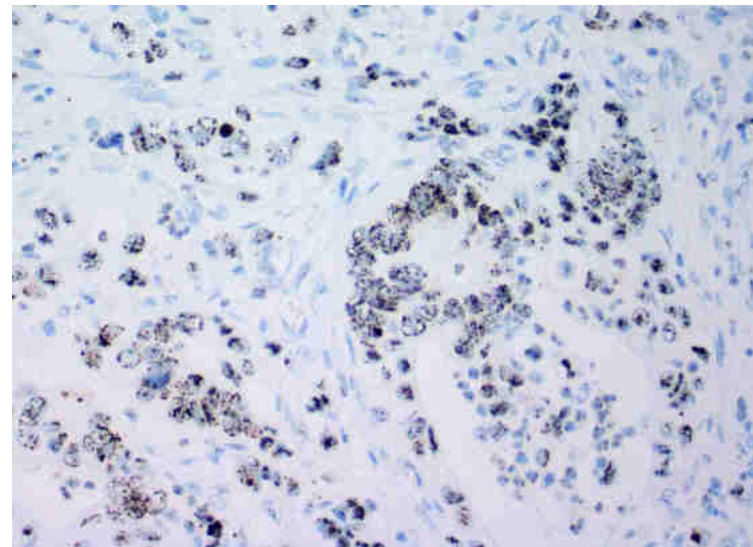


Evaluation of HER2 status

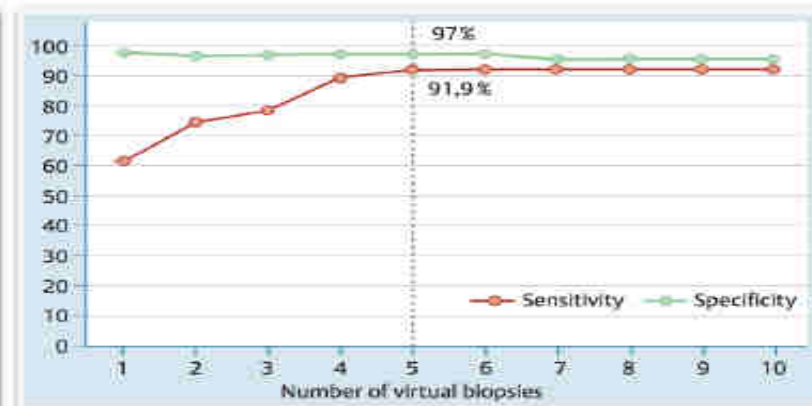
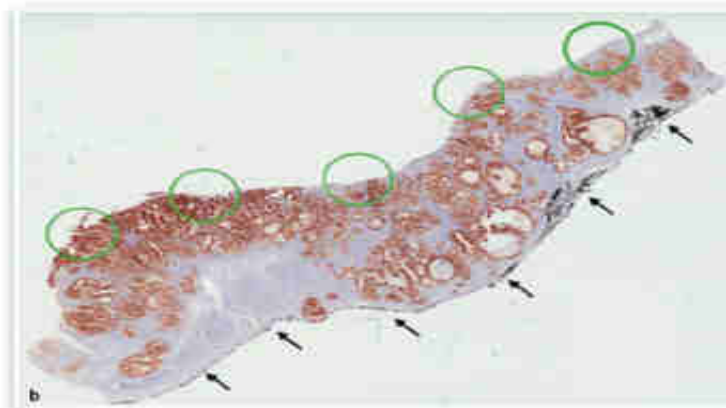
Immunohistochemistry



In situ hybridization



Minimum biopsy set for HER2 evaluation



Gastric Cancer
DOI 10.1007/s10120-015-0502-3

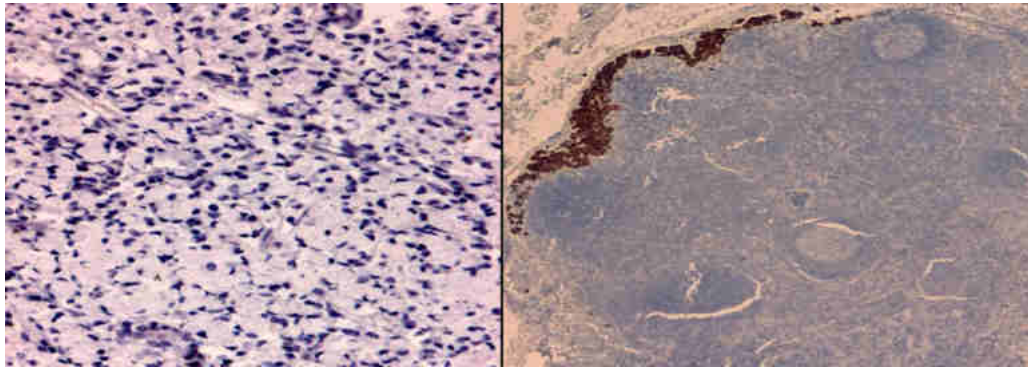
ORIGINAL ARTICLE

Five biopsy specimens from the proximal part of the tumor reliably determine HER2 protein expression status in gastric cancer

“Minimum biopsy set for HER2 evaluation in gastric and gastro-oesophageal cancer”
Endosc Int Open. 2015 Apr;3(2):E165-70
doi: 10.1055/s-0034-1391359

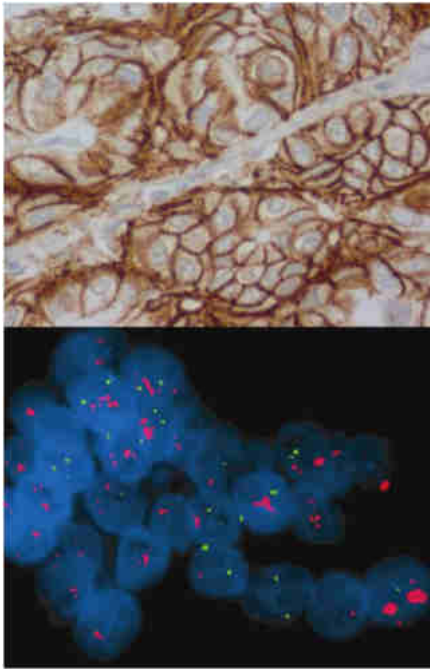
Tominaga N et al. Gastric Cancer 2016. doi: 10.1007/s10120-015-0502-3.

Differential expression of HER2 in gastric carcinoma and lymph node metastases



Putative impact on the therapeutic management and prognosis of the patients

HER-2 in gastric carcinoma: prognostic and/or predictive factor



Prognostic factor  YES (56%)
NO (44%)

HER-2 amplification
in intestinal-type gastric carcinoma



Blood born metastases
Poor prognosis

David L *et al*; Mod Pathol 5:384, 1992
Barros-Silva J *et al*; Br J Cancer 100: 487,2009

Predictive factor

ToGA Trial

HER-2 overexpression in 22% of advanced gastric cancers; improved survival in patients treated with with trastuzumab

ASCO 2009 (LBA 4509)

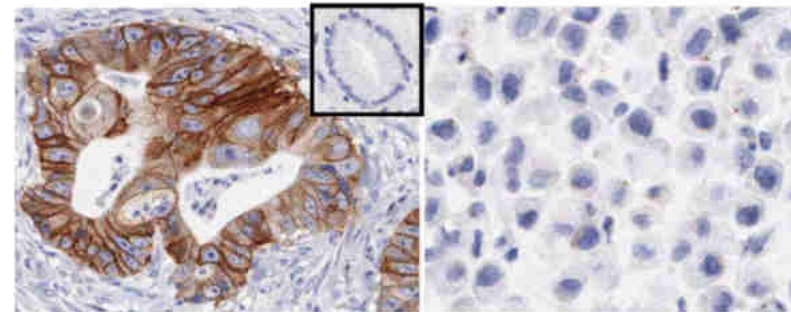
HER2 status in gastric cancer (prognostic and/or predictive factor?)

- HER2 expression is not related to gastric cancer patient prognosis and only a very small subgroup of intestinal type GC may potentially respond to HER2 targeting therapy.

Cellular Oncology 32 (2010) 57–65
DOI 10.3233/CLO-2009-0497
IOS Press

HER2 expression in gastric cancer:
Rare, heterogeneous and of no prognostic value – conclusions from 924 cases of two independent series

Heike Grabsch^{a,*}, Shivan Sivakumar^a, Saffy Gray^b, Helmut E. Gabbert^c and Wolfram Müller^d



HER2 status in gastric cancer (prognostic and/or predictive factor?)

JOURNAL OF CLINICAL ONCOLOGY

ORIGINAL REPORT

Level of *HER2* Gene Amplification Predicts Response and Overall Survival in HER2-Positive Advanced Gastric Cancer Treated With Trastuzumab

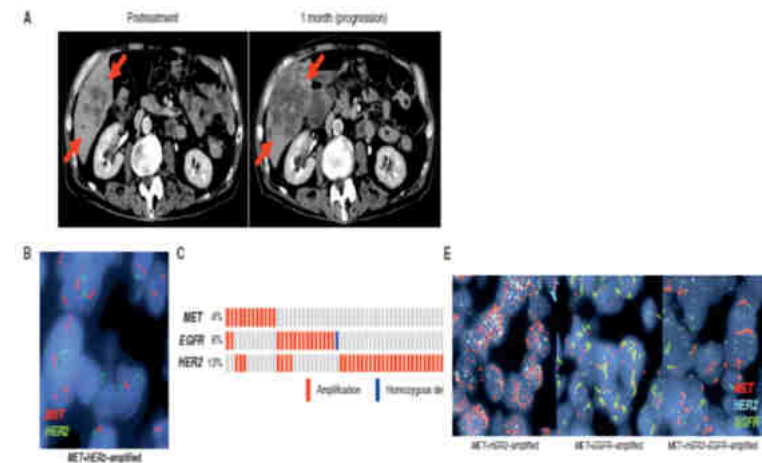
It is not only the quality but also the quantity

Resistance to HER2 targeted therapy in gastric cancer

- Patients initially respond to HER2 targeted therapy but eventually become resistant to treatment.
- Individual tumours with similar clinical stage have different clinical outcomes.

Putative causes

- Heterogeneity of HER2 expression
- Presence of ERBB2/EGFR co-amplification in the same tumour cells or even in the same tumour cells.
- HER2 copy number in ctDNA



Potential molecular targets in gastric cancer

Anti-EGFR

negative phase-3: EXPAND, REAL3

Lordick et al. Lancet Oncol 2013

Waddell et al. Lancet Oncol 2013

Anti-MET

negative phase-3: MetMab, RiloMet

Shah et al. ASCO 2015

Cunningham et al. ASCO 2015

anti-FGFR

preliminary phase-2: Shine

Bang et al. ASCO 2015

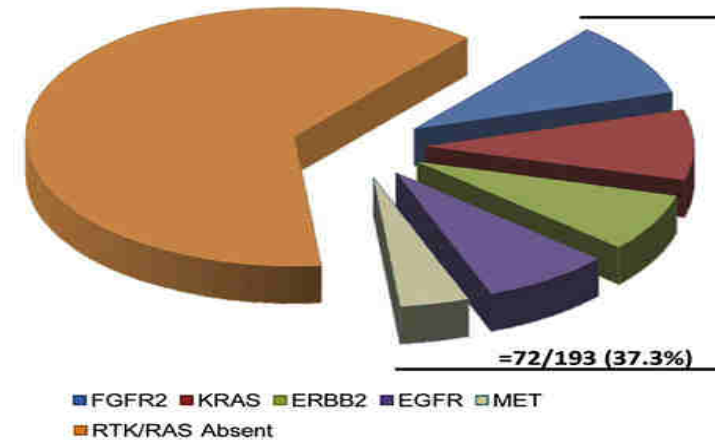
KRAS

non druggable (?)

HER2

positive phase-3: ToGA

Bang et al. Lancet 2010



Deng N, et al. *Gut* 2012;61:673-84

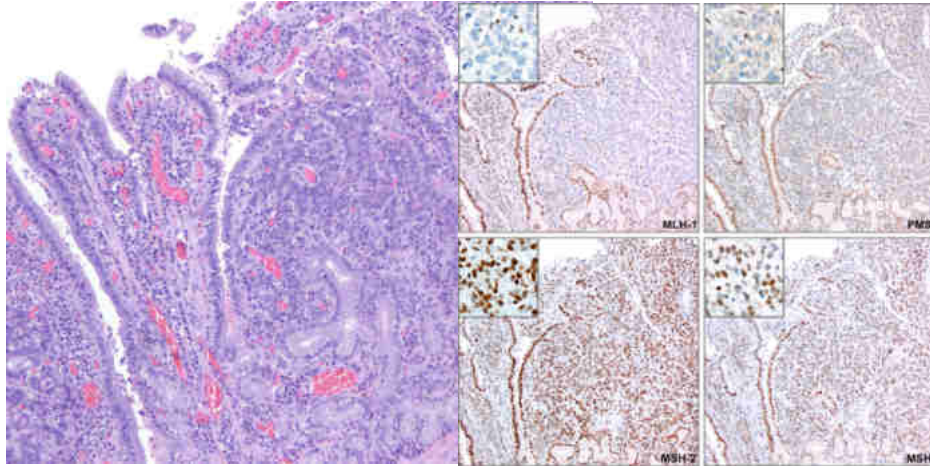
Actionable gene-based classification by NGS toward precision medicine

Table 1 Frequent gene alterations in 207 Japanese gastric cancers

Number	Mutation gene	Frequency	SCNA gene	Alteration	Frequency
1	<i>TP53</i>	53.1%	<i>ERBB2</i>	AMP	12.1%
2	<i>ARID1A</i>	15.9%	<i>CCNE1</i>	AMP	6.8%
3	<i>CDH1</i>	14.0%	<i>KRAS</i>	AMP	5.8%
4	<i>BRCA2</i>	10.6%	<i>ZNF217</i>	AMP	5.8%
5	<i>ARID1B</i>	10.1%	<i>CDKN2A</i>	DEL	5.3%
6	<i>ATM</i>	9.7%	<i>CDKN2B</i>	DEL	5.3%
7	<i>PIK3CA</i>	8.7%	<i>GATA4</i>	AMP	4.3%
8	<i>APC</i>	8.2%	<i>MYC</i>	AMP	2.4%
9	<i>ACVR2A</i>	7.2%	<i>CCND3</i>	AMP	1.9%
10	<i>CHD2</i>	6.3%	<i>CD274</i>	AMP	1.9%
11	<i>KMT2D</i>	6.3%	<i>CDK6</i>	AMP	1.9%
12	<i>RNF43</i>	5.8%	<i>EGFR</i>	AMP	1.9%
13	<i>EPHA2</i>	5.8%	<i>FGFR2</i>	AMP	1.9%
14	<i>TGFB2</i>	5.3%	<i>JAK2</i>	AMP	1.9%
15	<i>FLCN</i>	4.3%	<i>GNAS</i>	AMP	1.9%
16	<i>PALB2</i>	4.3%	<i>CCND1</i>	AMP	1.4%
17	<i>PTPR</i>	4.3%	<i>MET</i>	AMP	1.4%
18	<i>RAD50</i>	4.3%	<i>HSP90AB1</i>	AMP	1.4%
19	<i>BRCA1</i>	3.9%	<i>SMAD4</i>	DEL	1.4%
20	<i>STK11</i>	3.9%	<i>TEK</i>	DEL	1.4%

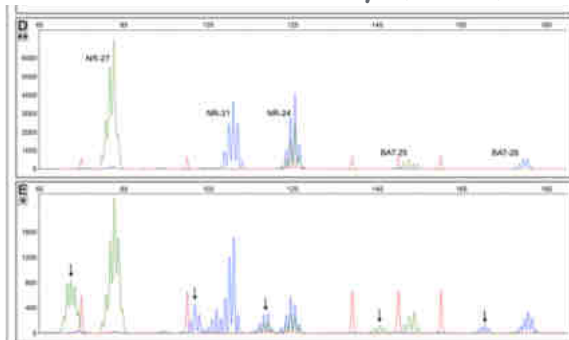
MSI in gastric carcinoma

Mismatch Repair Deficiency (MMRd)



(Immunohistochemistry)

Microsatellite Instability (MSI)



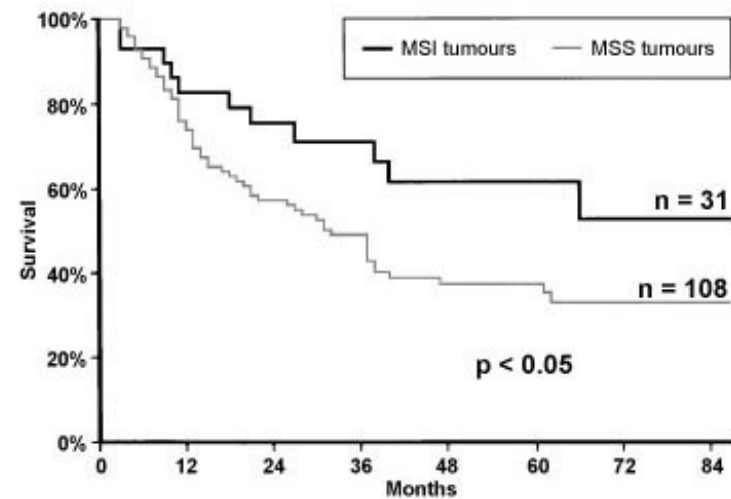
(Fluorescent Multiplex PCR)

Next Generation Sequencing (NGS)

- Instability signatures
- Instability burden (correlation with overall survival)

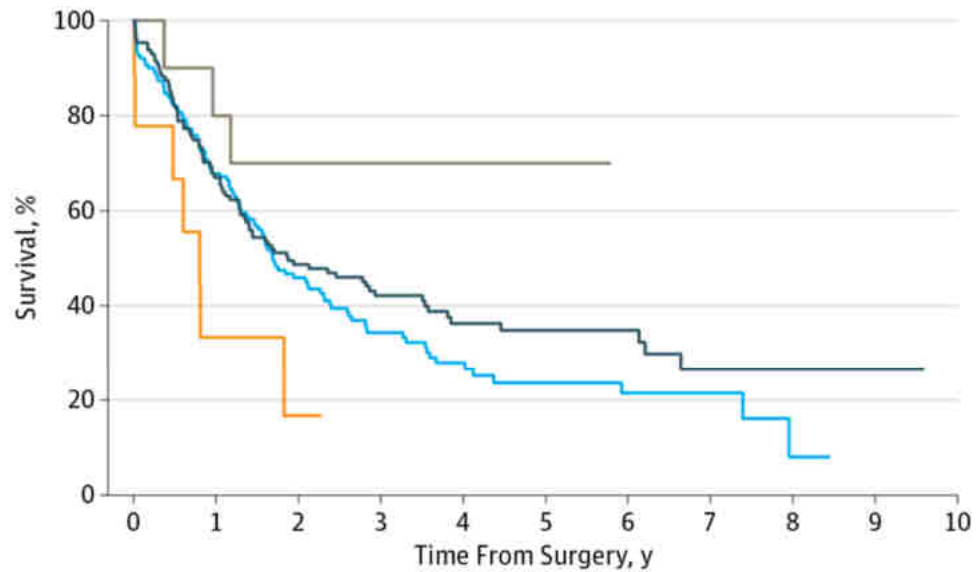
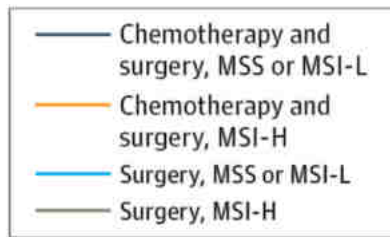
MSI in gastric carcinoma

Molecular marker of good prognosis in sporadic gastric cancer (caused by *hMLH1* promoter hypermethylation)



Survival of patients

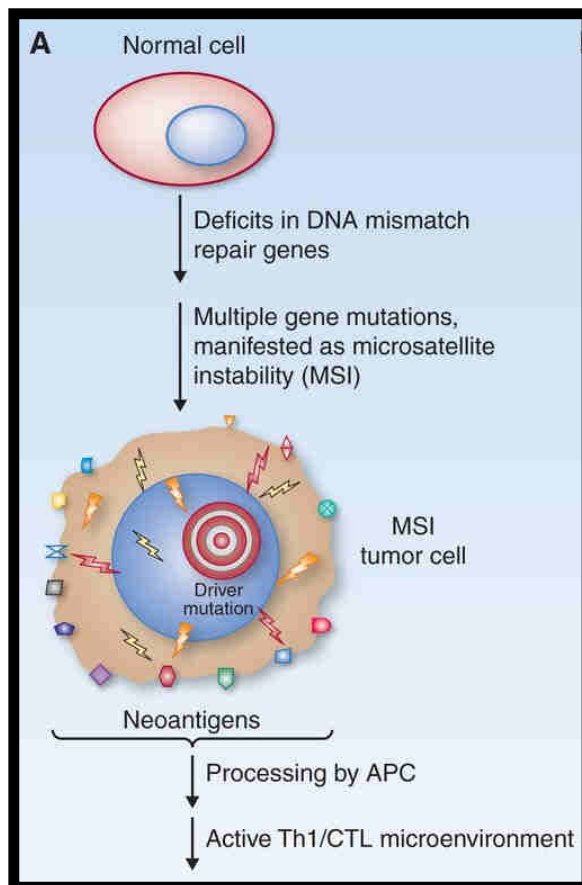
MSI and Prognosis



No. at risk

Chemotherapy and surgery, MSI-negative patients	129	85	58	42	27	22	15	6	3	1
Chemotherapy and surgery, MSI-positive patients	9	3	1							
Surgery, MSI-negative patients	151	100	58	37	21	13	9	7	1	
Surgery, MSI-positive patients	10	8	6	3	1	1				

MSI and immunotherapy



Science REPORTS

Cite as: D. T. Le *et al.*, *Science* 10.1126/science.aan6733 (2017).

Mismatch-repair deficiency predicts response of solid tumors to PD-1 blockade

Dung T. Le,^{1,2,3} Jennifer N. Durham,^{1,2,3*} Kellie N. Smith,^{1,2*} Hao Wang,^{3*} Bjarne R. Bartlett,^{3,4*} Laveet K. Aulakh,^{3,4} Steve Lu,^{3,4} Holly Kemberling,³ Cara Wilt,³ Brandon S. Luber,³ Fay Wong,^{3,4} Nilofer S. Azad,^{1,2} Agnieszka A. Rucki,^{1,2} Dan Laheru,³ Ross Donehower,³ Atif Zaheer,³ George A. Fisher,⁴ Todd S. Crocenzi,⁷ James J. Lee,⁸ Tim F. Greten,⁹ Austin G. Duffy,³ Kristen K. Ciombor,¹⁰ Aleksandra D. Eyring,¹¹ Bao H. Lam,¹² Andrew Joe,¹¹ S. Peter Kang,¹² Matthias Holdhoff,³ Ludmila Danilova,^{1,2} Leslie Cope,^{1,2} Christian Meyer,³ Shibin Zhou,^{1,2,4} Richard M. Goldberg,¹² Deborah K. Armstrong,² Katherine M. Bever,³ Amanda N. Fader,¹² Janis Taube,^{1,2} Franck Housseau,^{1,2} David Spetzler,¹² Nianqing Xiao,¹⁴ Drew M. Pardoll,^{1,2} Nickolas Papadopoulos,^{3,4} Kenneth W. Kinzler,^{3,4} James R. Eshleman,¹² Bert Vogelstein,^{1,2,4} Robert A. Anders,^{1,2,15} Luis A. Diaz Jr.,^{1,2,3,12}

The genomes of cancers deficient in mismatch repair (MMR) contain exceptionally high numbers of somatic mutations. In a proof-of-concept study, we previously showed that colorectal cancers with MMR deficiency were sensitive to immune checkpoint blockade with anti-PD-1 antibodies. We have expanded this study to now evaluate efficacy of PD-1 blockade in patients with advanced MMR-deficient cancers across 12 different tumor types. [...] These data support the hypothesis that the large proportion of mutant neoantigens in MMR-deficient cancers make them sensitive to immune checkpoint blockade, regardless of the cancers' tissue of origin.

Science 28;357(6349), 2018; 409-413. doi: 10.1126/science.aan6733

Gastric cancer and immune checkpoint blockade

Predictive biomarkers



MSI-high *status*

Science

REPORTS

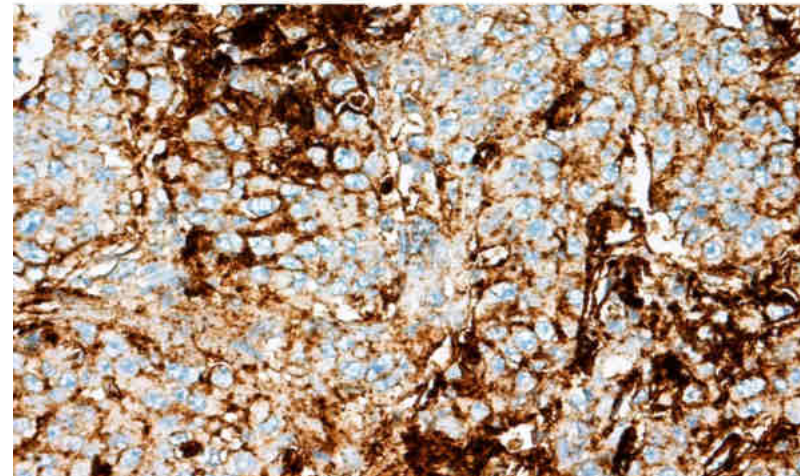
Cite as: D. T. Le *et al.*, *Science*
10.1126/science.aan6733 (2017).

Mismatch-repair deficiency predicts response of solid tumors to PD-1 blockade

PD-L1 expression

Original Article

Clinical Utility of the Combined Positive Score for Programmed Death Ligand-1 Expression and the Approval of Pembrolizumab for Treatment of Gastric Cancer



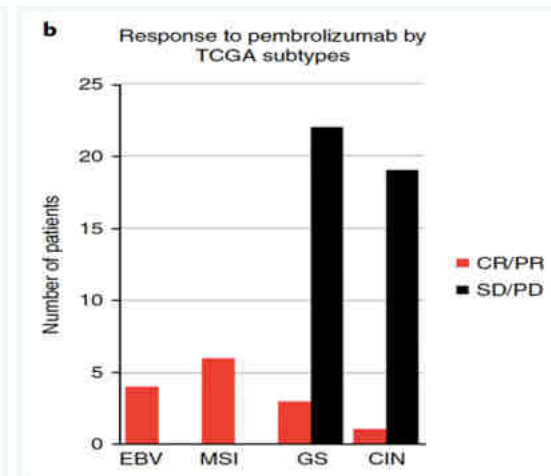
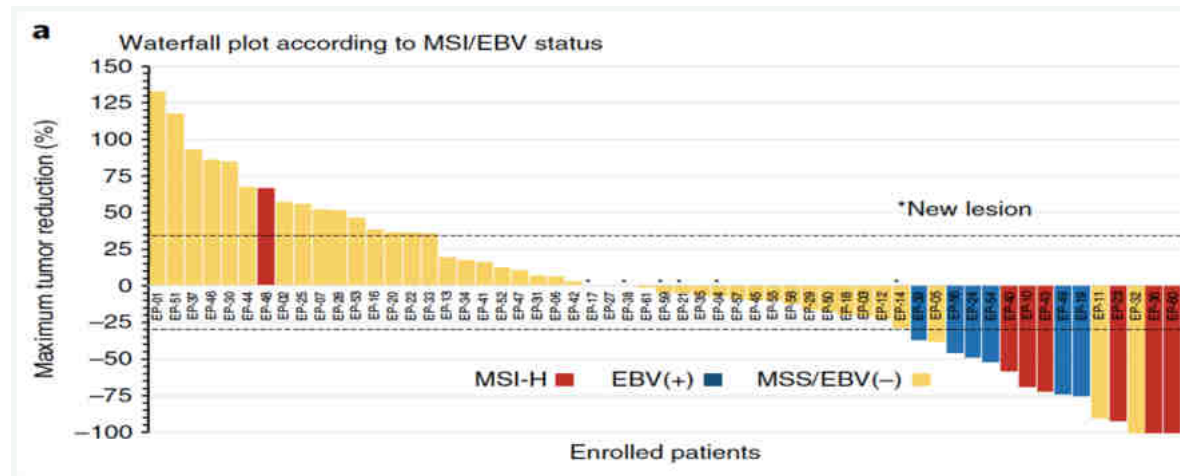
Gastric cancer and immune checkpoint blockade

Predictive biomarkers

EBV+ and MSI-high status



Comprehensive molecular characterization of clinical responses to PD-1 inhibition in metastatic gastric cancer



Le DT et al Science 2017; Kulangara K Arch Pathol Lab Med. 2018; Kim ST Nat Med 2018

EBV infection and MSI in gastric cancer




International Journal of
Molecular Sciences

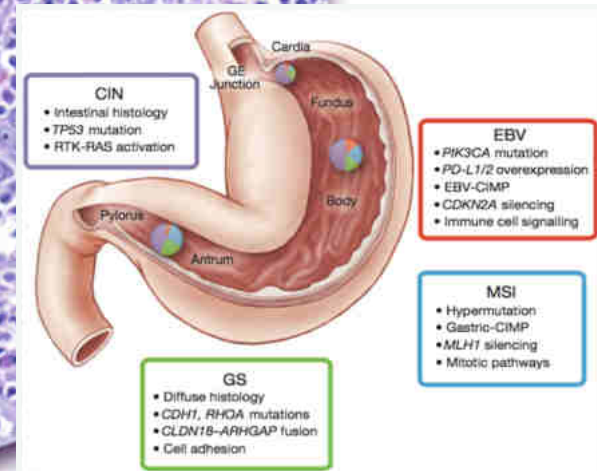
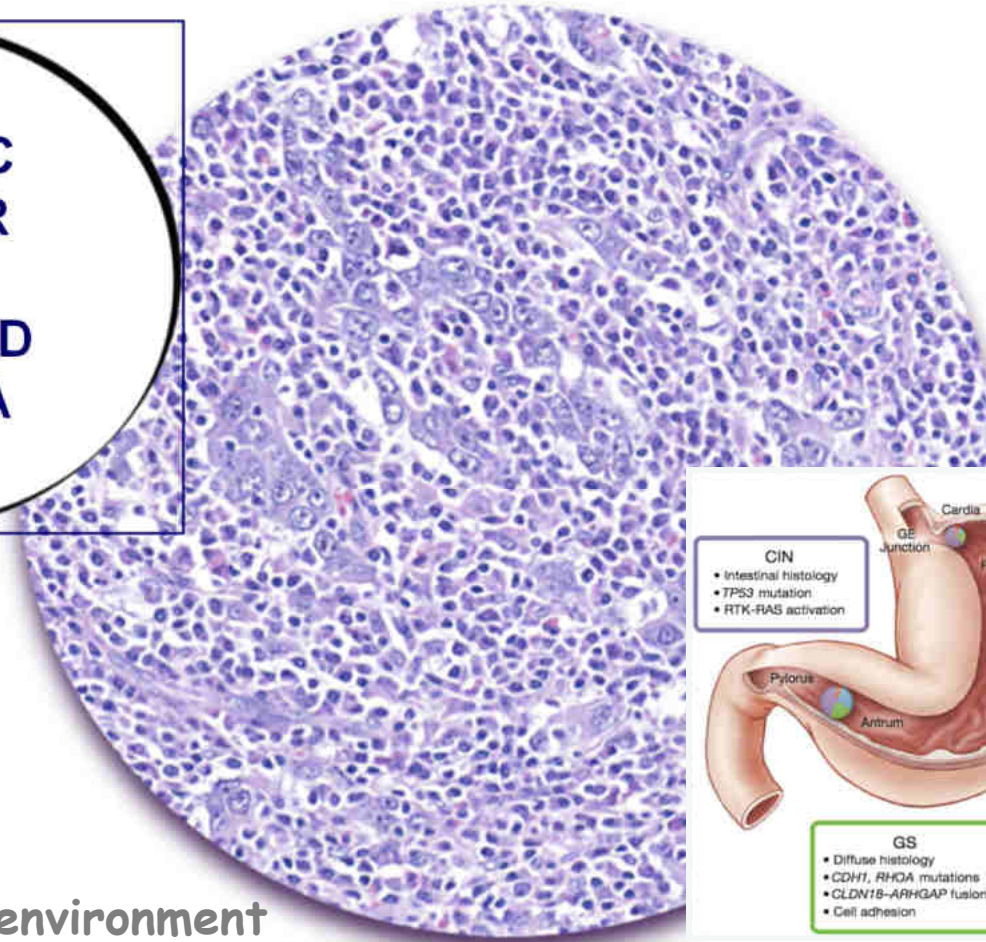


Article

The Transcriptomic Landscape of Gastric Cancer: Insights into Epstein-Barr Virus Infected and Microsatellite Unstable Tumors

Irene Gullo ^{1,2,3,4}, Joana Carvalho ^{3,4}, Diana Martins ^{3,4}, Diana Lemos ^{3,4}, Ana Rita Monteiro ^{3,4},
Marta Ferreira ^{3,4}, Kakoli Das ⁵, Patrick Tan ^{5,6,7}, Carla Oliveira ^{3,4}, Fátima Carneiro ^{1,2,3,4}  and
Patrícia Oliveira ^{3,4,*}

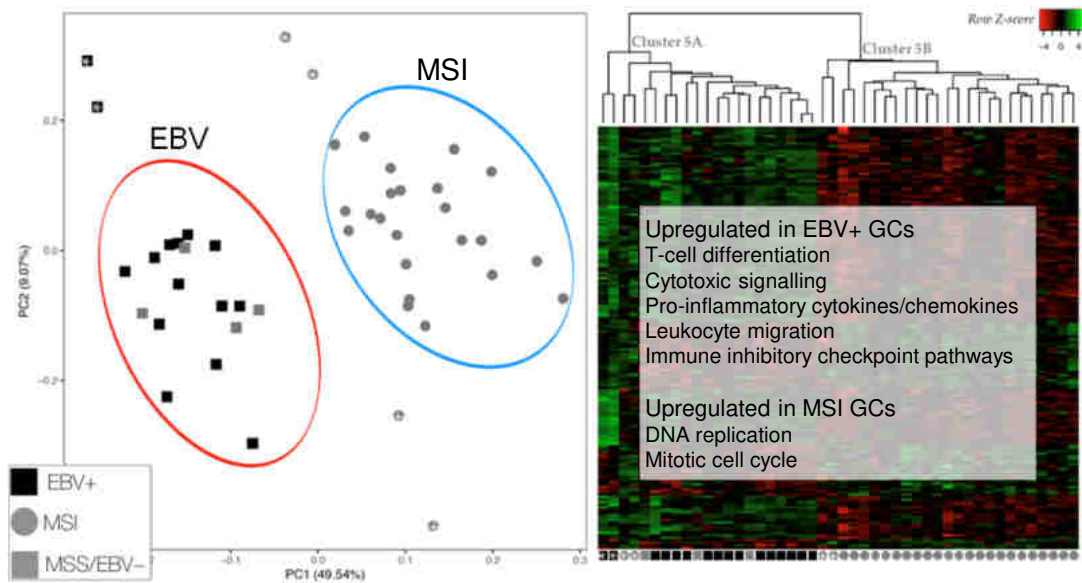
GASTRIC CANCER WITH LYMPHOID STROMA



The tumour microenvironment

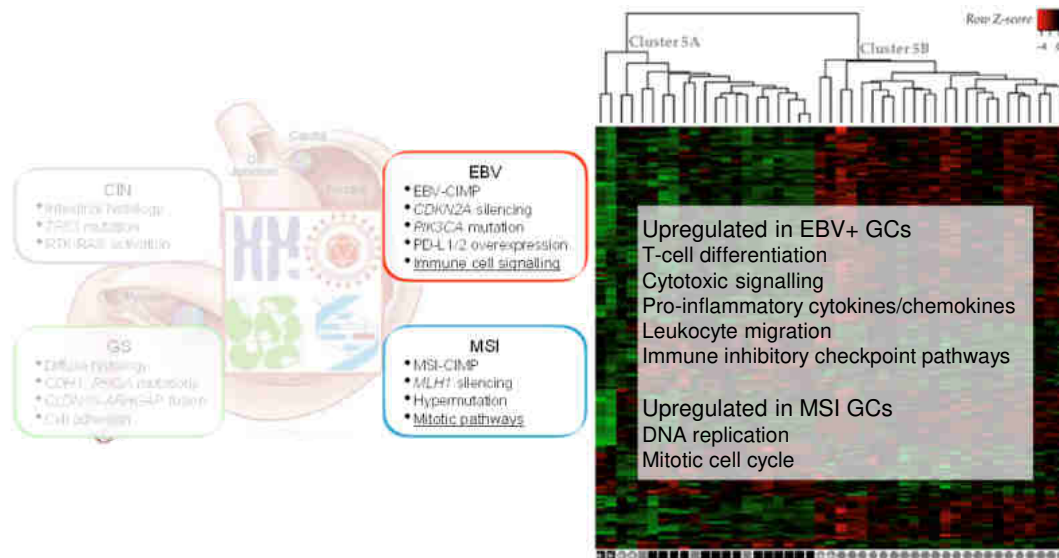
EBV+ and MSI GCs displayed distinct transcriptomic signatures

Unclustered analysis: differentially expressed (DE) genes

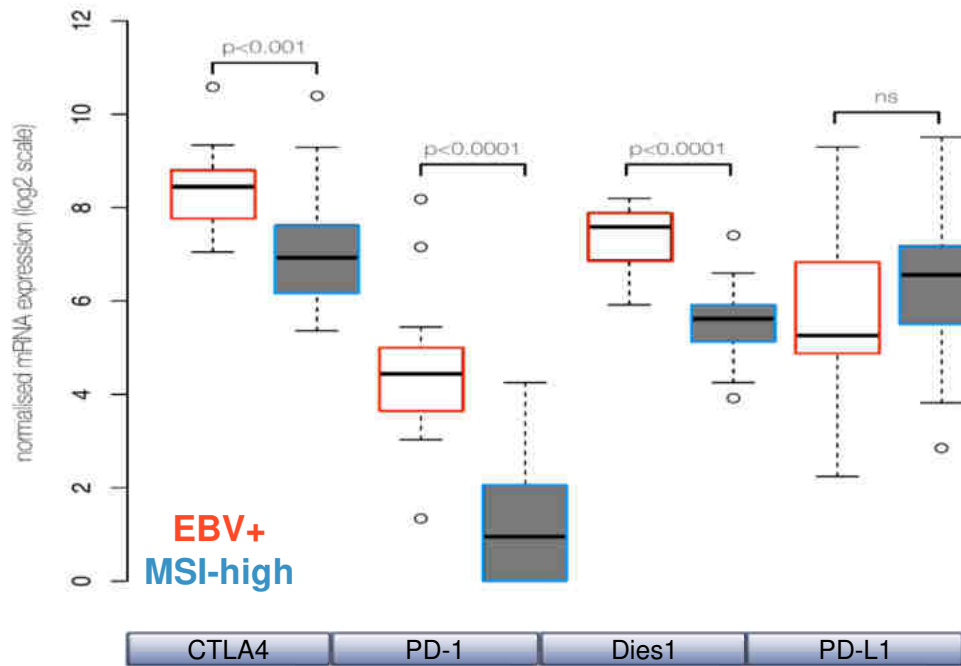


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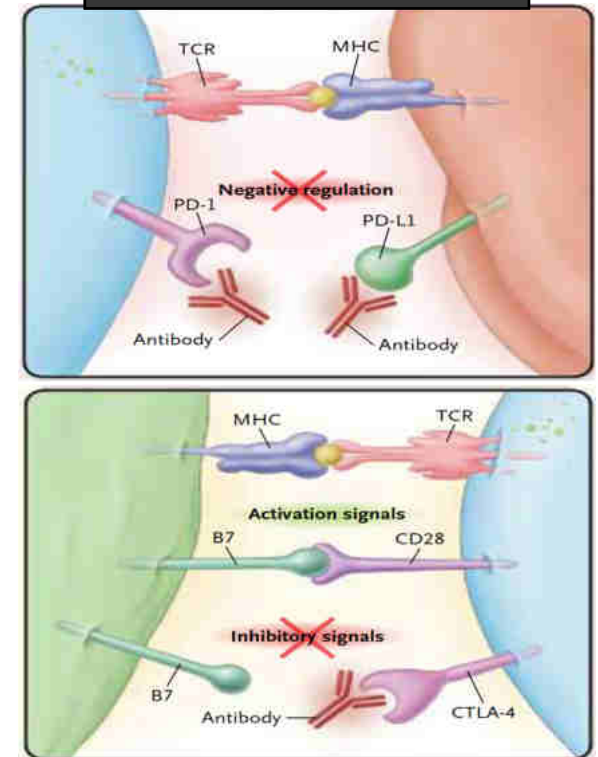
Immunotherapy targets in EBV and MSI gastric cancers



PD-L1 protein expression

- Cancer cells: No differences
- Immune cells: EBV+ showed higher expression than MSI-high cases ($p=0.0052$)

Combination immunotherapies



Ribas A et al NEJM 2012

Tumour types for which immune check point immunotherapies are FDA-approved

Tumor type	Therapeutic agent	FDA approval year
Melanoma	Ipilimumab	2011
Melanoma	Nivolumab	2014
Melanoma	Pembrolizumab	2014
Non-small cell lung cancer	Nivolumab	2015
Non-small cell lung cancer	Pembrolizumab	2015
Melanoma (BRAF wild-type)	Ipilimumab + nivolumab	2015
Melanoma (adjuvant)	Ipilimumab	2015
Renal cell carcinoma	Nivolumab	2015
Hodgkin lymphoma	Nivolumab	2016
Urothelial carcinoma	Atezolizumab	2016
Head and neck squamous cell carcinoma	Nivolumab	2016
Head and neck squamous cell carcinoma	Pembrolizumab	2016
Melanoma (any BRAF status)	Ipilimumab + nivolumab	2016
Non-small cell lung cancer	Atezolizumab	2016
Hodgkin lymphoma	Pembrolizumab	2017
Merkel cell carcinoma	Avelumab	2017
Urothelial carcinoma	Avelumab	2017
Urothelial carcinoma	Durvalumab	2017
Urothelial carcinoma	Nivolumab	2017
Urothelial carcinoma	Pembrolizumab	2017
MSI-high or MMR-deficient solid tumors of any histology	Pembrolizumab	2017
MSI-high, MMR-deficient metastatic colorectal cancer	Nivolumab	2017
Pediatric melanoma	Ipilimumab	2017
Hepatocellular carcinoma	Nivolumab	2017
Gastric and gastroesophageal carcinoma	Pembrolizumab	2017
Non-small cell lung cancer	Durvalumab	2018
Renal cell carcinoma	Ipilimumab + nivolumab	2018





James P. Allison and Tasuku Honjo

Discovery of cancer therapy by inhibition of negative immune regulation

Wei SC, Duffy CR, Allison JP. Cancer Discovery 2018. doi: 10.1158/2159-8290.CD-18-0367

Clinical relevance of molecular diagnosis

Contents lists available at [ScienceDirect](#)


 **Digestive and Liver Disease** 

journal homepage: [www.elsevier.com/locate/dl](#) [European Journal of Cancer](#) 86 (2017) 305–317


Position Paper

The multidisciplinary management of gas-
tumours

European Society of Digestive Oncology (ESDO) expert discussion and report from the 16th ESMO World Congress on Cancer, Barcelona

 Available online at [www.sciencedirect.com](#)


ScienceDirect

journal homepage: [www.ejccancer.com](#) 

Review

Clinical relevance of molecular diagnostics in gastrointestinal (GI) cancer: European Society of Digestive Oncology (ESDO) expert discussion and recommendations from the 17th European Society for Medical Oncology (ESMO)/World Congress on Gastrointestinal Cancer, Barcelona

Alexander Baraniskin ^{a,*}, Jean-Luc Van Laethem ^b, Lucjan Wyrwicz ^c, Ulrich Guller ^d, Harpreet S. Wasan ^e, Tamara Matysiak-Budnik ^f, Thomas Gruenberger ^g, Michel Ducreux ^h, Fatima Carneiro ⁱ, Eric Van Cutsem ^{j,1}, Thomas Seufferlein ^{k,1}, Wolff Schmiegel ^{a,1,*}



Clinical relevance of molecular diagnosis

Brief summary of the recommendations.

Tumor entity	Molecular genetic marker	Recommendation
Oesophageal and gastric adenocarcinoma	HER2 status	Part of standard diagnostics
	HER2 status re-testing	For patients with metastatic or recurrent cancer when initially HER2-neg. or borderline low/1–2+/FISH negative
	Liquid biopsy in HER2-testing	Not enough data, not recommended in routine use

Table 1

Brief summary of the recommendations.

Tumor entity	Molecular genetic marker	Recommendation
Oesophageal and gastric adenocarcinoma	HER2 status	Part of standard diagnostics
	HER2 status re-testing	For patients with metastatic or recurrent cancer when initially HER2-neg. or borderline low/1–2+/FISH negative
	Liquid biopsy in HER2-testing	Not enough data, not recommended in routine use
	MSI	Recommended for stage IV alone
	EBV in tissue	Not recommended in routine use
	PD-L1 expression	Not recommended in routine use
	FGFR expression/gene fusions	Not recommended in routine use
Esophageal SCC	Molecular diagnostic	No biomarkers are recommended in routine use

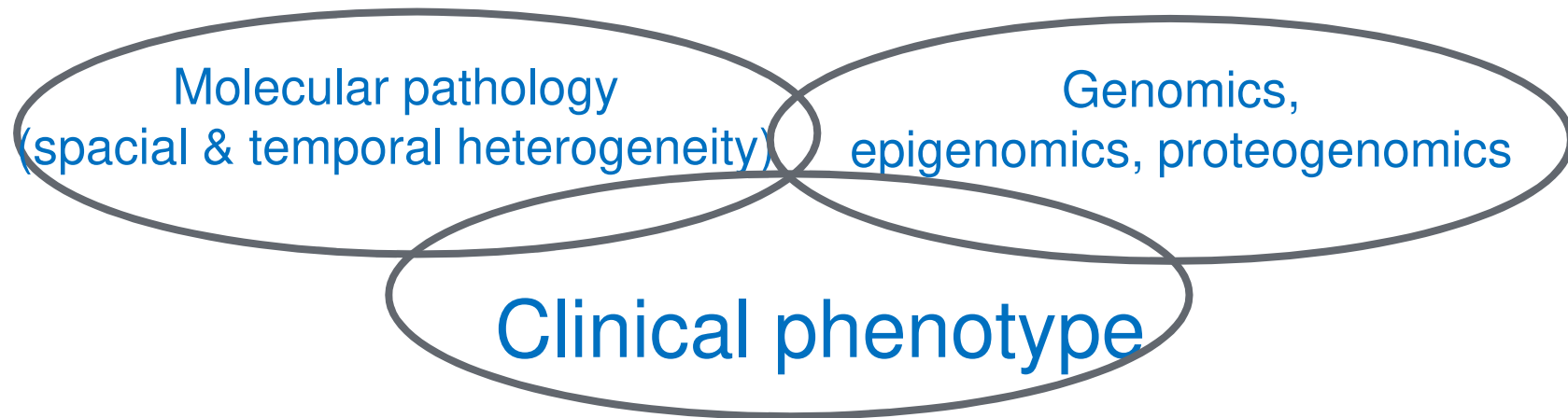
routine use

Take home lessons

- 1) **Established predictive biomarkers**, such as HER2 (anti-HER2 therapy benefits patients with unresectable or metastatic/recurrent HER2-positive GC, and HER2 testing is used to predict potential therapy response)
- 2) **Biomarkers partly established and/or under development such as:**
 - a. Receptor tyrosine kinases;
 - b. MSI *status* and EBV infection;
 - c. Biomarkers for cancer immunotherapy (Tumour mutational load, density of intratumoural CD8+ T cell infiltrates and PD-L1 expression);
 - d. Two molecular subtypes (MSI-high and EBV + might be potential good candidates for immunotherapy targeting of the PD-L1/PD-1 axis).

Upfront molecular testing. Is it time yet?

Integrated Molecular Pathology



- Better understanding
- Translation to clinics

- Lloyd M et al: Pathology to enhance precision medicine in oncology: Lessons from landscape ecology. *Adv Anat Pathol* 22: 267, 2015
- Salto-Tellez M & Kennedy M: Integrated molecular pathology: the Belfast model. *Drug Discovery Today* 20: 1451, 2015



Thanks for your attention