he image cannot be displayed. Your computer may not have enough memory to open the image, or the image may have been corrupted. Restart your computer, and then open the file again. If the red x still appears, you may have to delete the image and then insert it again.

Transposable elements in humans

Scott E. Devine, Ph.D.

Associate Professor Institute for Genome Sciences, Department of Medicine, and Greenebaum Cancer Center, University of Maryland School of Medicine



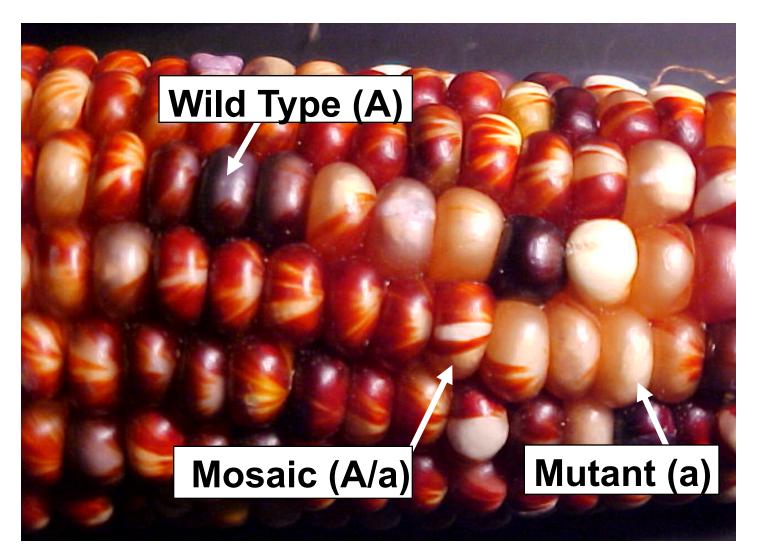
McClintock's transposons caused phenotypic changes

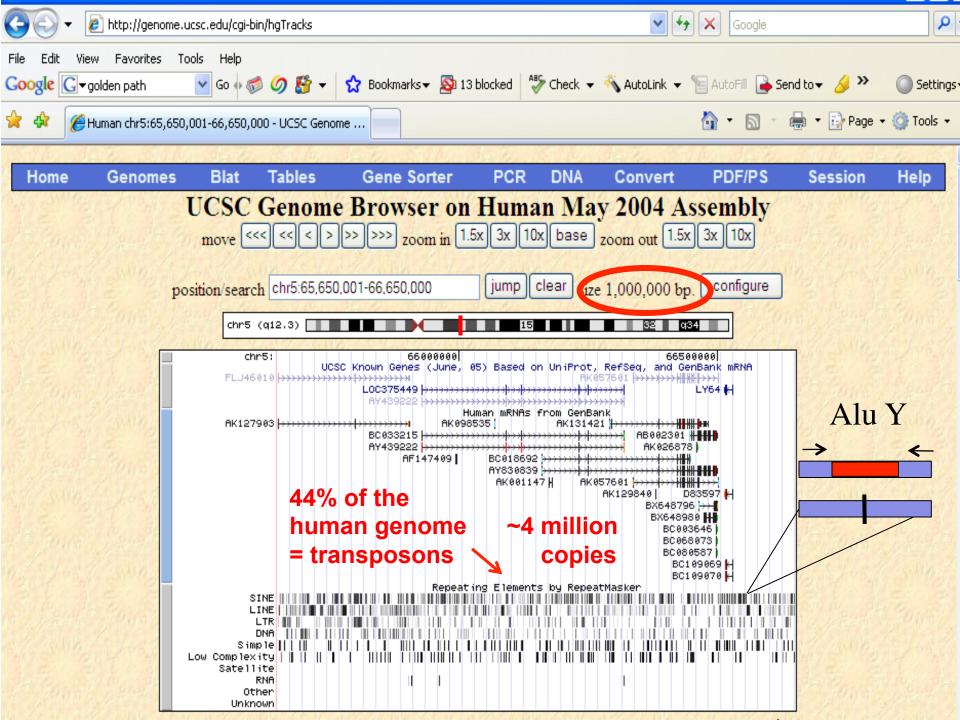


Born 1902, Hartford, CT. B.A. 1923, Cornell University Ph.D. 1927, Cornell University, Botany 1927-1931, Instructor in Botany, Cornell University 1931-1933, Fellow, National Research Council 1933-1934, Fellow, Guggenheim Foundation 1934-1936, Research Associate, Cornell University 1936-1941, Assistant Professor, University of Missouri 1942-1967, Staff member, Carnegie Institution of Washington's Department of Genetics, Cold Spring Harbor, NY 1967-1992, Distinguished Service Member, CIW Department of Genetics, Cold Spring Harbor 1944, Member, National Academy of Sciences 1945, President, Genetics Society of America 1967, Kimber Medal 1970, National Medal of Science 1981, Lasker Award 1983, Nobel Prize in Physiology or Medicine

Source: Cold Spring Harbor Laboratory (http://www.cshl.org/History/mcclintock.html)

McClintock's transposons caused phenotypes

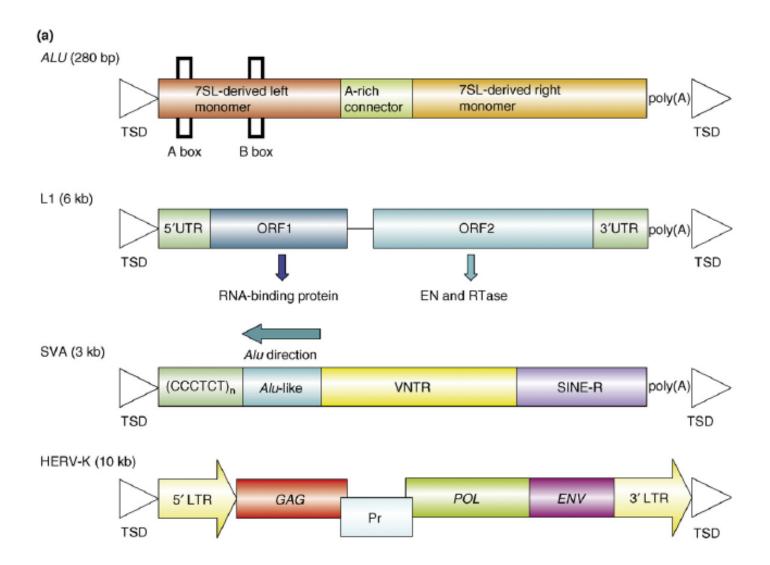




Human transposons cause phenotypes (diseases) too

	Gene	Disorder	Element	<u>Mechanism</u>
Alu	NF1	Neurofibromatosis	Alu Ya5	Intron/skipping
	BCHE	Acholinesterasemia	Alu Yb8	Exon insertion
	F9	Hemophilia B	Alu Ya5	Exon insertion
	CASR	Familial hypocalciuric hypercalemia	Alu Ya4	Exon insertion
	ADD1	Huntington disease	Alu	Intron
L1	Factor VIII	Hemophilia A	L1	Exon insertions
	APC	FAP	L1	Exon insertion
	Dystrophin	Muscular Dystrophy	L1	Exon insertions
	Globin	beta thalassemia	L1	Intron
	RP2	Retinitis Pigmentosis	L1	Intron
	Fukutin	Muscular Dystrophy	L1	Intron/skipping

Mills, R.E., Bennett, E.A., Iskow, R.C., and Devine, S.E. (2007) Trends Genet. 23:183-91.



Mills, R.E., Bennett, E.A., Iskow, R.C., and Devine, S.E. (2007) *Trends Genet.* 23:183-91.

Human TEs produce a large amount of genetic variation

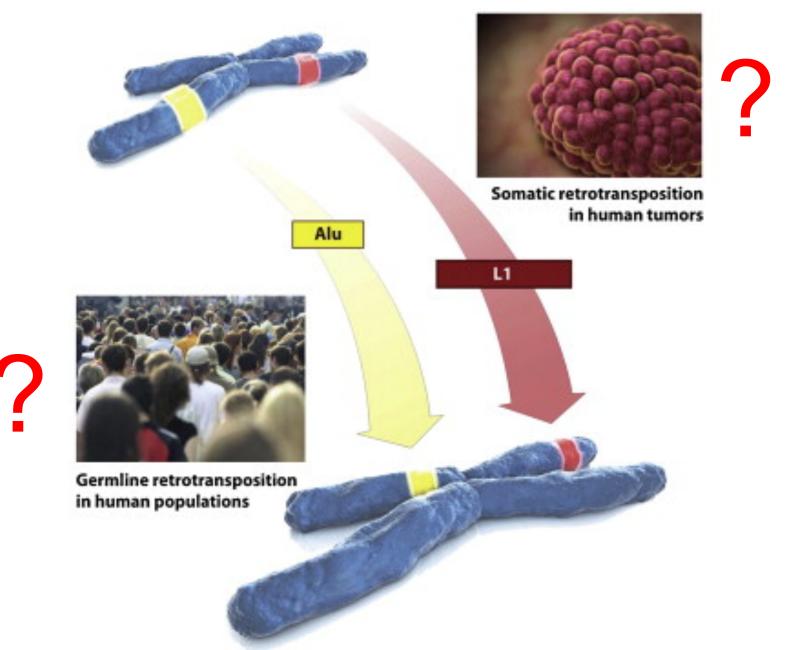
~1 in 20 to 200 live births is predicted to have a new germline TE insertion

Kazazian, H.H. (1999) *Nat. Genet.* **22**, 130. Li, X. et al. (2001) *Hum. Mutat.* **34**, 511-519. Cordaux, R. et al. (2006) *Gene* **373**, 134-137.

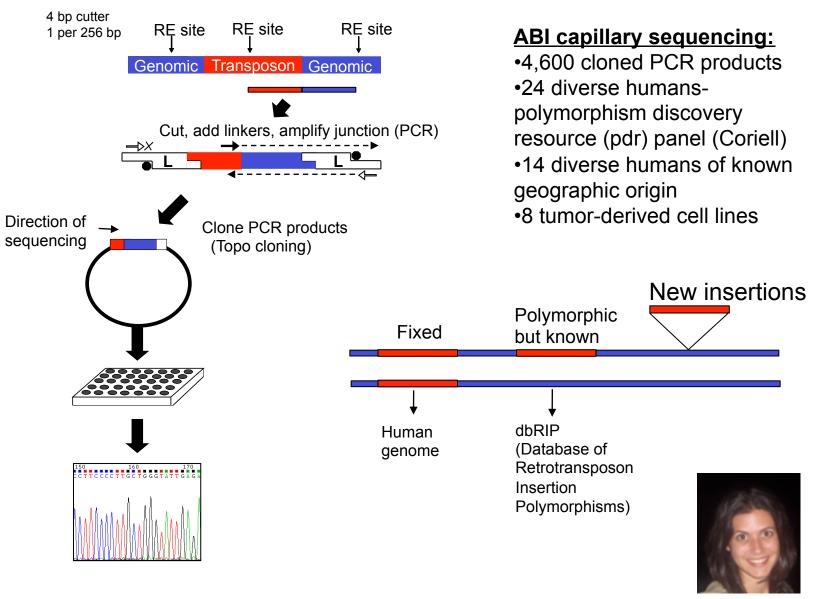
6 billion people X 0.05 = 300 million germline TE insertions

Equivalent to 1 new TE insertion for every 10 bp of genome (an impressive mutagenesis experiment!!)

Needed a better method to find new insertions



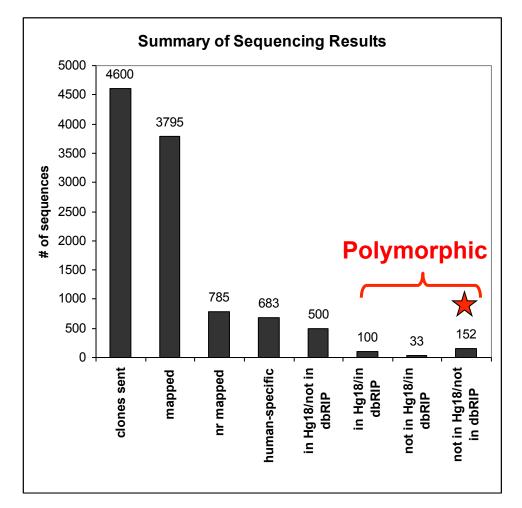
"Transposon-seq" technologies to identify new L1 and Alu insertions

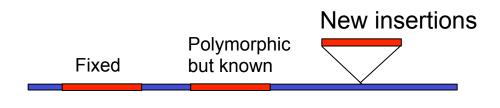


Iskow et al. (2010) Cell 141, 1253-1261

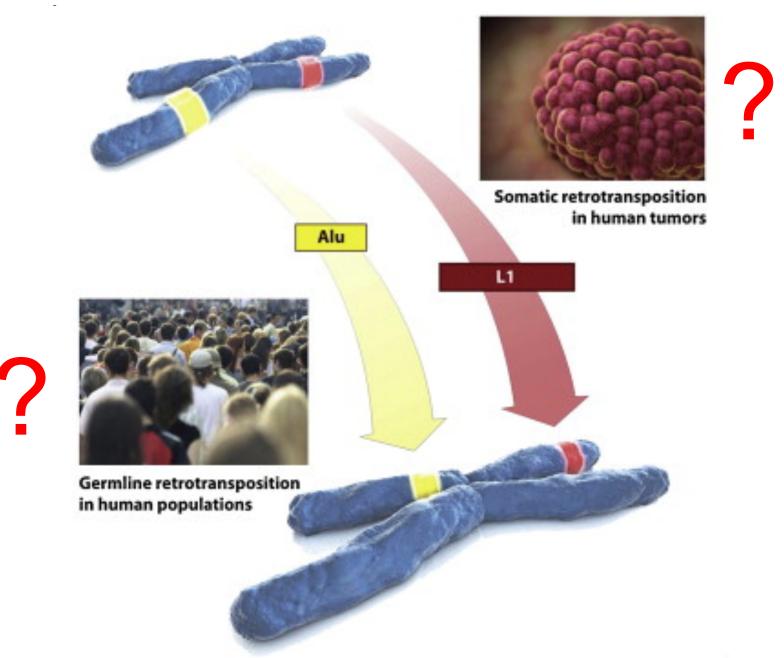
Rebecca Iskow

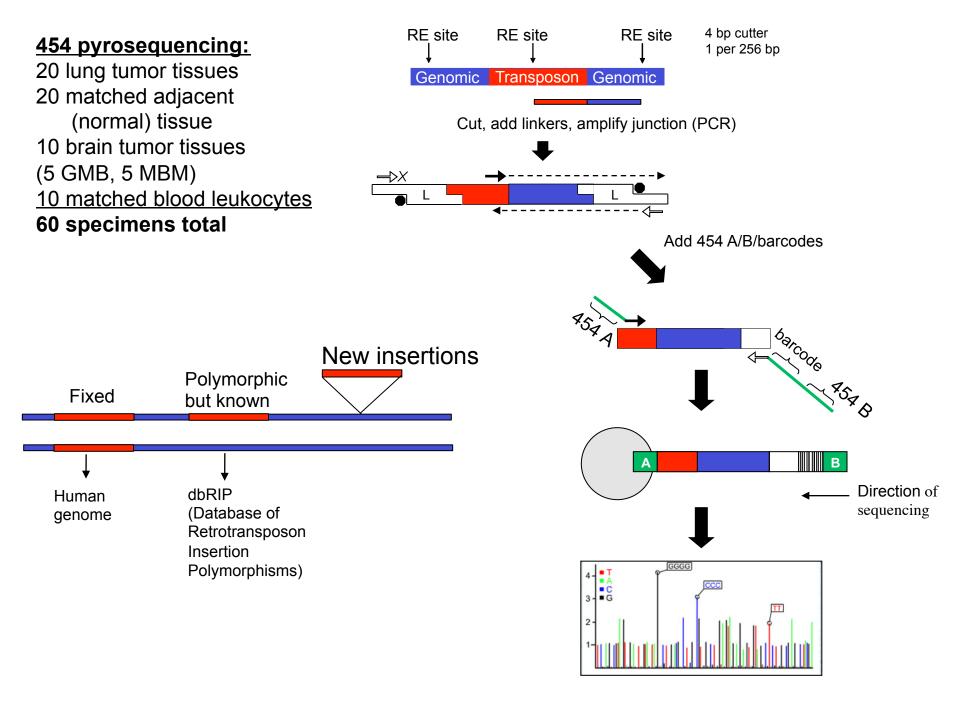
We found 152 New L1 insertions





Can we detect new germline and somatic TE insertions in cancer patients?



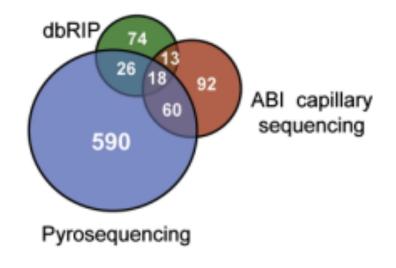


Summary of transposon-seq data

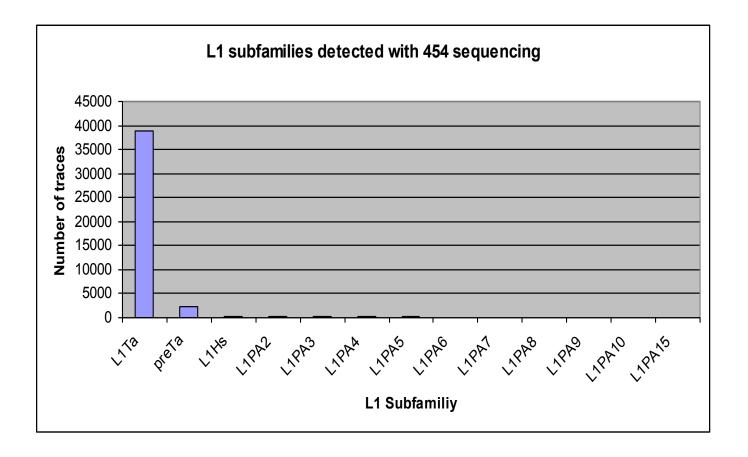
<u>Sequencing</u> <u>Strategy</u>	Sample Description	<u>Retrotransposon</u>	<u>Reads</u>	<u>Mapped</u>	<u>Distinct</u> <u>Retrotransposons</u>	<u>Previously</u> <u>Unknown</u>	<u>PCR</u> Validated
ABI capillary sequencing	Pools of diverse human DNA and tumor-derived cell line DNA.	L1	4,600	3,795	785	152	64/66 (97%)
Pyrosequencing	Lung tumor and adjacent normal lung DNA. Brain tumor and matched normal blood DNA.	L1	266,126	50,532	1,389	650	162/182 (89%)
Pyrosequencing	Brain tumor and matched normal blood DNA.	Alu	35,022	22,338	3,799	403	53/56 (95%)

We identified 742 novel L1's and 403 novel Alus (1145 total)

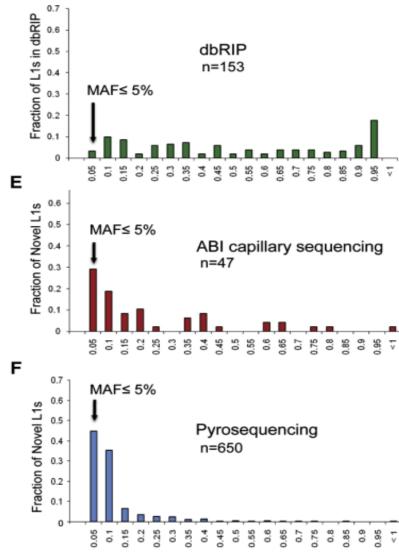
L1 elements beyond the reference sequence



Most of new insertions detected belong to the youngest, most active subfamilies in the genome

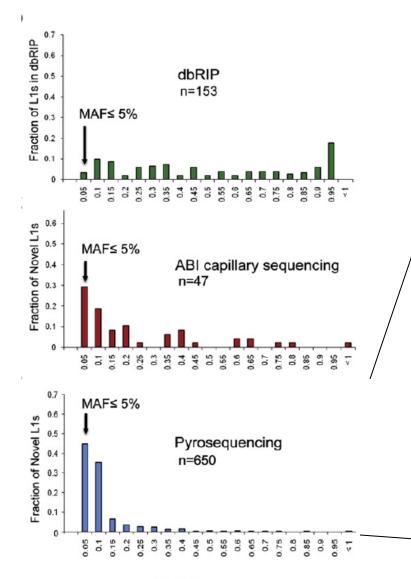


Most of the novel insertions also had low allelic frequencies, further indicating that they were inserted recently



Allelic Frequency

Singletons (insertions found only in a single specimen) Include some of the most recently-inserted L1's



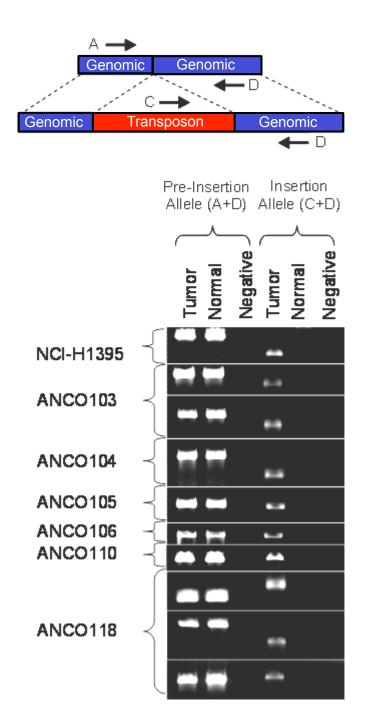
<u>339 "singleton" L1's</u>

Glioblastoma (GBM):	57
Normal leukocytes, GBM:	56
Medulloblastoma (MBM):	18
Normal leukocytes, MBM:	32
Lung tumor:	101
Normal lung:	75

Follow up PCRs on lung tumor singletons: 9 were somatic insertions; Most of the rest were in matched normal too, so rare germline insertions

~10% no PCR (some likely subclonal)

Allelic Frequency



9 somatic insertions In lung cancers validated by PCR

Cloned and sequenced Insertion junctions to Verify—flanked by tsd's

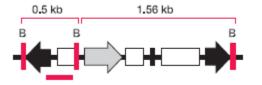
6/20 (30%) of lung tumors positive for at least one new L1 Insertion

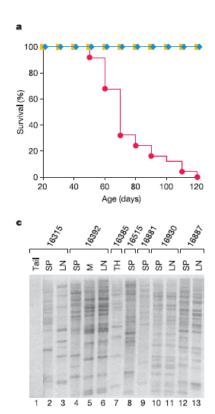
ARTICLES

b

Mammalian mutagenesis using a highly mobile somatic Sleeping Beauty transposon system

Adam J. Dupuy¹, Keiko Akagi¹, David A. Largaespada², Neal G. Copeland¹ & Nancy A. Jenkins¹



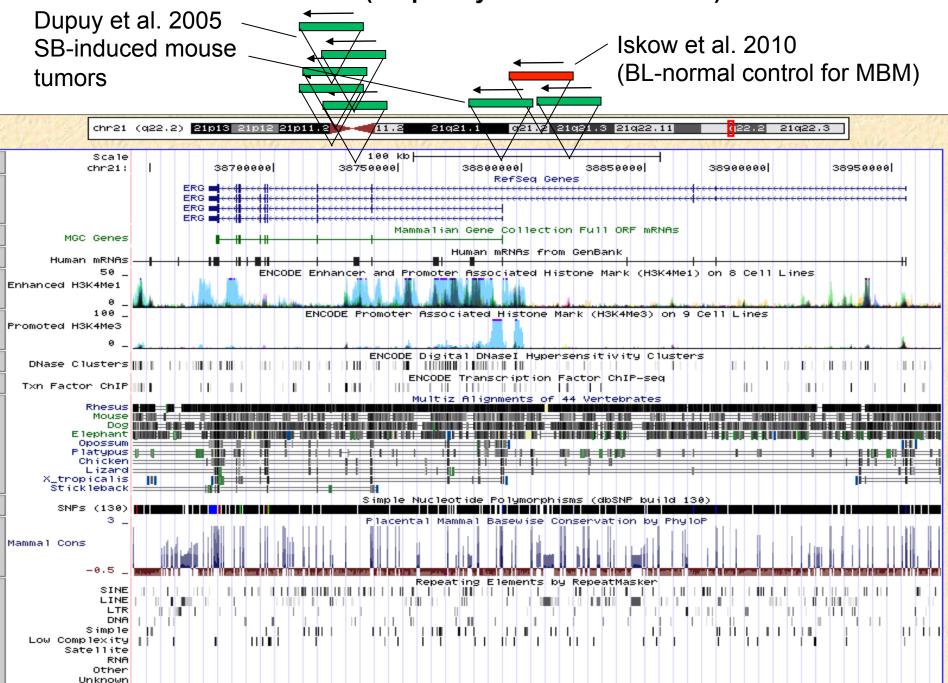


Mouse D	Age (days)	Tumour type
TG6057-16315	114	T-cel
TG6057-17106	101	Medulloblastoma
TG6057-17306	84	T-cel, non-T/non-B
TG6057-18137	45	B-ce
TG6070-16392	57	B-ce
TG6070-16515	65	T-cell (thymic), non-T/
		non-B (disseminated)
TG6070-16881	58	Non-T/non-B
TG6070-16887	62	T-cell
TG6070-17221	60	T-ce
TG6070-17223	87	B-cell, hyperplasia
		in duodenum
TG6070-17503	102	Non-T/non-B
TG6070-17900	68	T-cell (thymic), non-T/
		non-B (disseminated)
TG6070-17902	68	T-ce
TG6070-18533	73	T-ce
TG6070-18665	53	T-cell
TG6070-18732	47	T-cell, hyperplasia
		in duodenum
TG6070-18804	58	Non-T/non-B,
		medulloblastoma,
		intestina neoplasia
TG6113-16835	67	T-ce
TG6113-16927	96	T-cel,
		pituitary hyperp las ia
TG6113-16930	71	T-ce
TG6113-17548	52	T-cell, non-T/non-B
TG6113-18019	59	Non-T/non-B
TG6113-18022	62	T-cel, non-T/non-B
TG6113-18521	62	T-cel

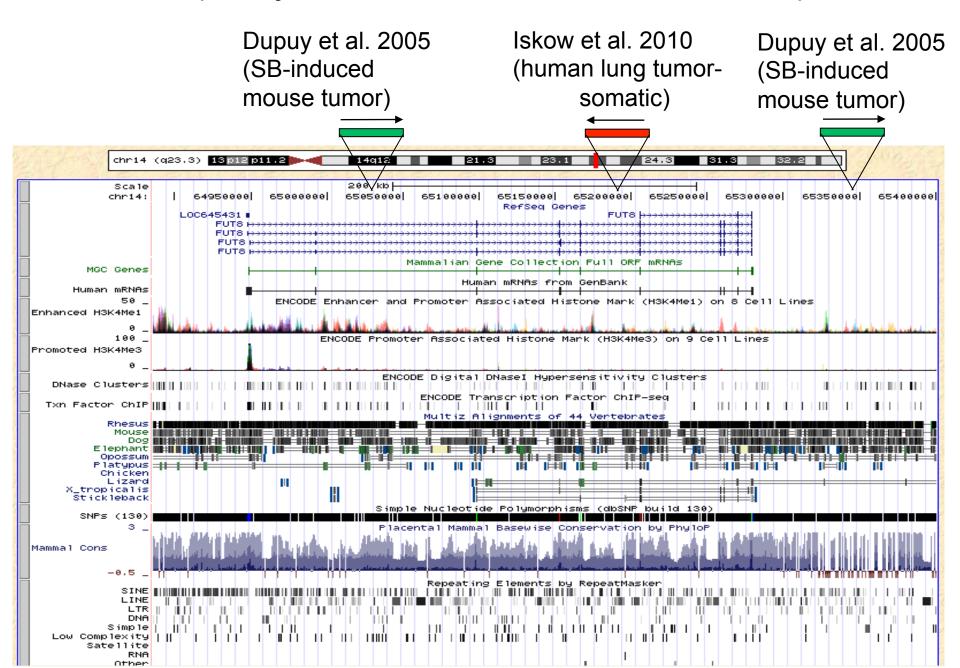
Figure 2 | Adult double-transgenic mice die from cancer. a, Survival curves showing decreased viability of double-transgenic mice: yellow, RosaSB; blue, T2/Onc2; red, double-transgenic. b, Age at death and tumour type of

double-transgenic mice. c, Southern analysis of BamHI-digested tumour DNA. Each band represents a separate SB transposon integration. LN, lymph node; M, mass; SP, spleen; TH, thymus.

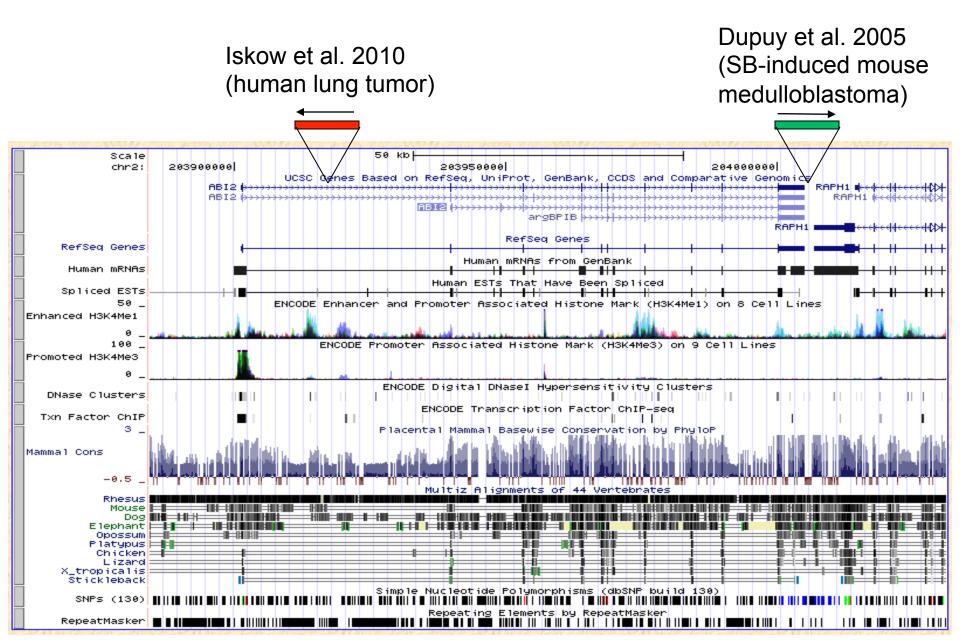
ERG (frequently mutated in cancers)

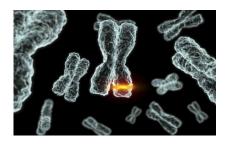


FUT8 (Fucosyltransferase 8—linked to tumor invasiveness)

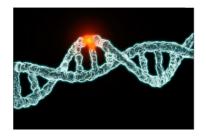


ABI-2 (ABL-Interacting-2; antagonizes oncogenic potential of ABL)





Conclusions



- 1. Transposon-seq revealed that rare Alu, L1 germline insertions are highly abundant in human genomes. Likely to have greatest Impact on humans because selection has not yet fully acted on them.
- 2. Somatic L1 insertions also occur at high frequencies in lung tumor genomes
- 3. Both germline and somatic insertions may work together to initiate and then drive tumorigenesis by hitting specific genes
- 4. Provides a new mechanism for mutagenizing cancer genomes in addition to point mutations, DNA repair, large-scale rearrangements
- 5. Once integrated, L1 might also promote large scale chromosomal rearrangements that are commonly found in human tumors

Emory University





Steve Pittard

Ryan Mills

R01HG002898 2T32GM00849 SUN Microsystems Paula Vertino Erwin Van Meir Mike McCabe

University of Maryland Institute for Genome Sciences

Julienne Mullaney Luke Tallon Jerry Liu Ankit Maroo Brandi Cantarel Ellen McRae Andy Neuwald Steve Pittard Research Associate Genomics Resource Center Senior bioinformatics engineer Bioinformatics analyst Research associate Postdoctoral fellow IGS faculty member Senior informatics engineer

R01HG002898-07, R01CA166661-01

