



# Vector NTI Advance® 11.5 Quick Start Guide

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**User Manual**

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# Introduction

This Quick Start Guide is designed to get you started using Vector NTI Advance® 11.5. It provides brief descriptions of the Vector NTI Advance® 11.5 graphical user interface, including **Vector NTI Explorer** and the **Molecule Viewer**, and step-by-step instructions for using the most common features and functions of the software. The topics covered include locating the desired tools, displaying molecules, designing PCR primers, cloning two fragments, gene synthesis, aligning molecules, performing a restriction analysis, and assembling contigs.

This guide assumes that you have a working knowledge of basic Microsoft® Windows® and Mac OS® features and functions (how to open and save files, how to use your mouse, and so on) and that Vector NTI Advance® 11.5 is installed on your computer.

# Opening Vector NTI Advance® 11.5

The **QuickStart Page** is a single page that consolidates most commonly used modules, tools, and utilities that Vector NTI provides.

To launch the QuickStart Page, select **Start > All Programs > Invitrogen > Vector NTI Advance 11 > Quick Start**.



Figure 1. QuickStart Page


You can configure the software to open both the **Molecule Viewer** and **Vector NTI Explorer** when you select **Vector NTI** from the **Start** menu.

1. In the **Molecule Viewer** window, go to the **View** menu and select **Options**.
2. In the **General** tab of the dialog, select the **Open Local Database at Startup** checkbox.
3. Click **OK** to make the change.

# Local Database

Vector NTI Explorer is the main tool for accessing the information in your local Vector NTI Advance® database. Using the Explorer, you can import, open, export, and organize molecules and other database items, and launch other Vector NTI Advance® modules (Figure 2).

To launch Vector NTI Explorer:

- On **QuickStart Page**, click on **Launch Local Database**.
- In the Molecule Viewer, click on the Local Database icon ()
- From the Windows® Start menu, select **Programs > Invitrogen > Vector NTI Advance 11 > Vector NTI Explorer**.
- The local database in Vector NTI Advance® contains records for different types of molecular biology objects. Each database record includes all the information for that object (e.g., a DNA molecule record includes the DNA sequence, defined features of the molecule, and other information). Objects in the database can include molecules, analysis results, BLAST search results, citations, and other types of information.

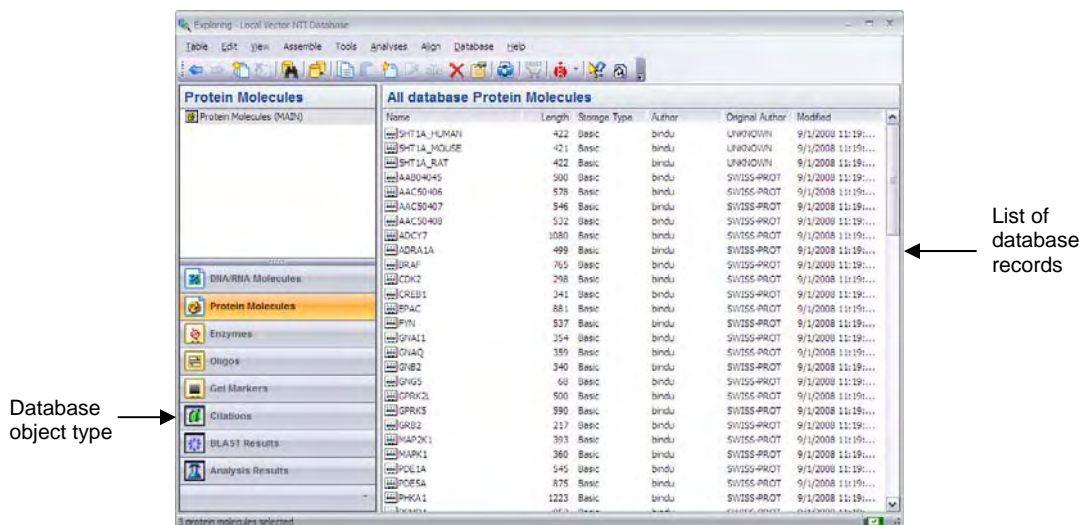


Figure 2. Vector NTI Explorer (Local Database) window

Database objects in Vector NTI Advance® are categorized by type (DNA molecules, protein molecules, and so on). Some molecules are installed with the software. When you first open the software, **DNA/RNA Molecules** is the selected object type. Click on the tab in the lower left corner of the **Vector NTI Explorer** to select from the other available database objects.

To open an object from the local database, double-click on the object name in the right-hand pane of the **Explorer**. Depending on the object type, information about that object may be displayed in a dialog box, or the object may be loaded into a viewer. For example, DNA, RNA, and protein molecules are displayed in the **Molecule Viewer**.



When you install Vector NTI Advance®, the default local database is created in a folder called **VNTI Database** in the root directory of your computer (e.g., C:\VNTI Database).

# Database Backup/Restore

It is strongly recommended that the local database be backed up routinely. You may launch the Database Backup manually, or use the Database Backup Reminder to trigger the task automatically as configured (Figure 3).

To manually perform Database Backup:

- From the Vector NTI Explorer menu, select **Database > Backup Database Now**.

To configure the Database Backup Reminder:

- From the Vector NTI Explorer menu, select **Database > Database Backup Reminder**.
- To set a specific date or interval (e.g., backup every 15 days), click on **Set Reminder**.

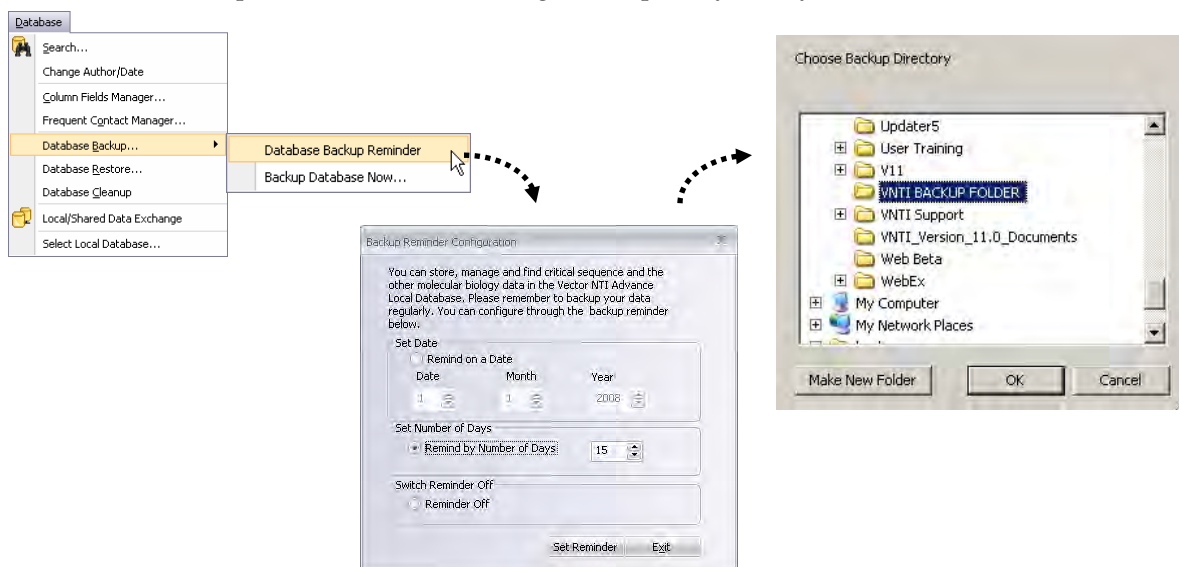


Figure 3. Database Backup

To restore a database:

- From Vector NTI Explorer menu, select **Database > Database Restore**.

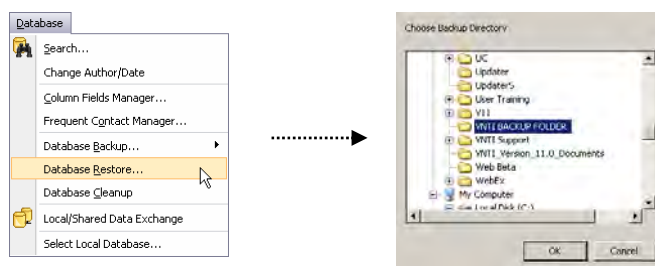



Figure 4. Database Restore

# Molecule Viewer

The Molecule Viewer displays information about DNA, RNA, and protein molecules. To launch the Molecule Viewer:

- Click on **Launch Molecule Viewer** on the **QuickStart** page, or
- From the Windows® Start menu, select **Programs > Invitrogen > Vector NTI Advance 11 > Vector NTI**, or
- Double-click on a molecule name in the **Vector NTI Explorer**.

To open a molecule from within the Molecule Viewer, click on the Open button (  ) on the main toolbar and select the molecule name from the dialog box.

The molecule will be loaded into the Molecule Viewer.

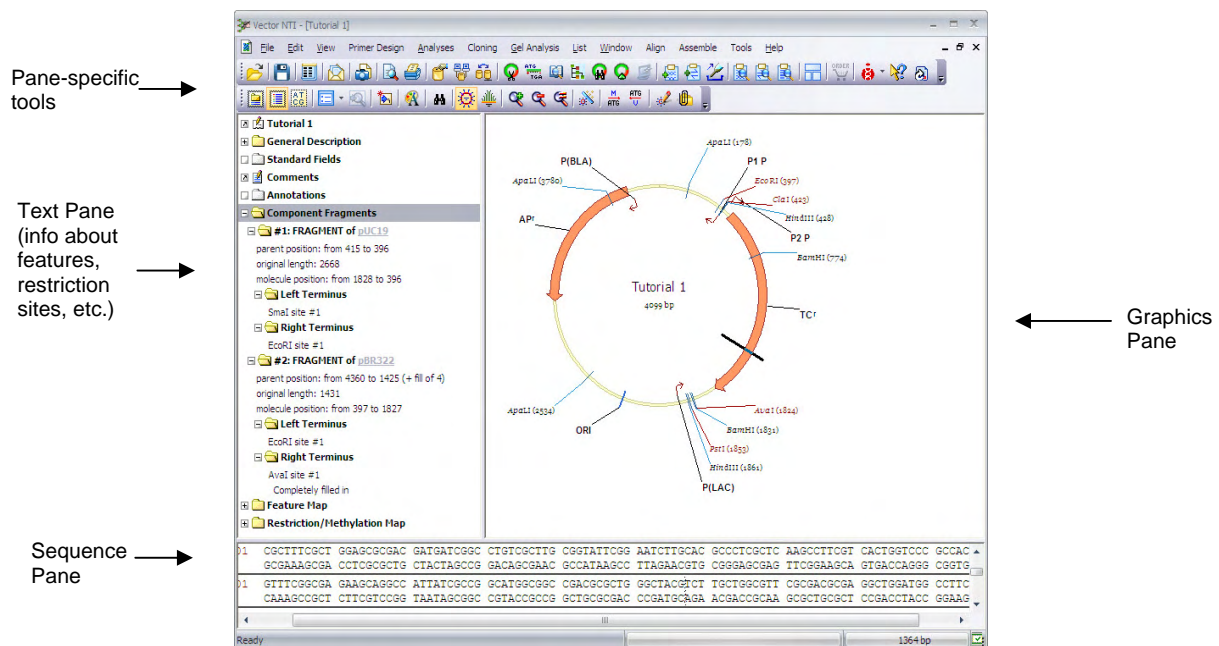


Figure 5. Molecule Viewer window for a DNA molecule

The Molecule Viewer window has different panes for displaying different types of information about the molecule, as shown in Figure 5. Click inside a pane to make it the active pane. The available tools and right-click menu options will change depending on which pane is active.


Use tools on the dropdown menus and toolbars to add information about the molecule and perform various analysis functions, as described in the step-by-step instructions on the following pages.

Multiple molecules can be displayed in separate windows of the Molecule Viewer.

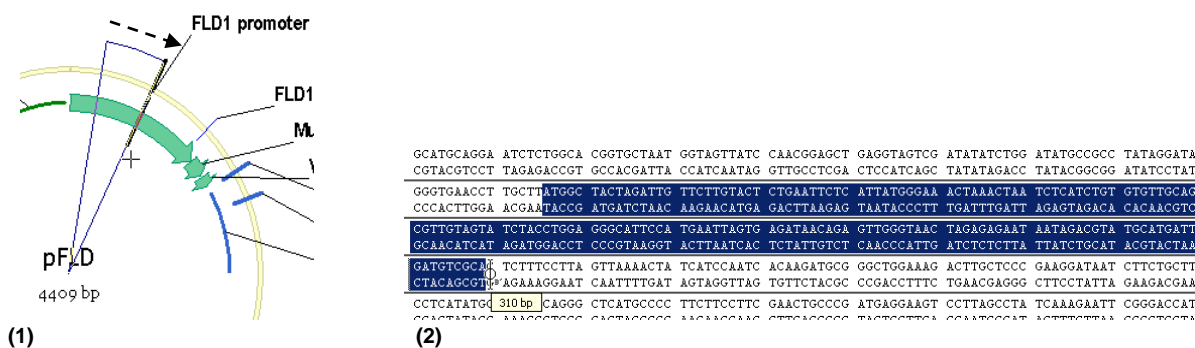


# Selecting and Editing Molecule Sequences

In the Molecule Viewer, you can select part of a molecule sequence in several different ways:

- Hold down the mouse button and drag the cursor across the sequence in the Sequence Pane or Graphics Pane (Figure 6).
- Go to the *Edit menu*, select **Set Selection**, and enter the sequence base-pair range in the dialog box.
- Click on a defined feature in the Graphics Pane.
- Click on a defined feature in the Text Pane, and click on **Find** (  ) on the main toolbar.

The selected sequence will appear highlighted in both the Graphics Pane and the Sequence Pane.



**Figure 6. Selecting a DNA sequence by (1) dragging in Graphics Pane or (2) dragging in Sequence Pane**

To copy a molecule sequence:

1. Select it as described above.
2. To copy it to the Windows® clipboard, use the CTRL + C keyboard command, or

To copy the sequence as a text file, go to the *Edit menu* and select **Copy to > File**. You will be prompted to select a format and enter a name for the file.

To delete a molecule sequence:

1. Select it as described above.
2. Click on the DELETE key on your keyboard.

To paste a molecule sequence:

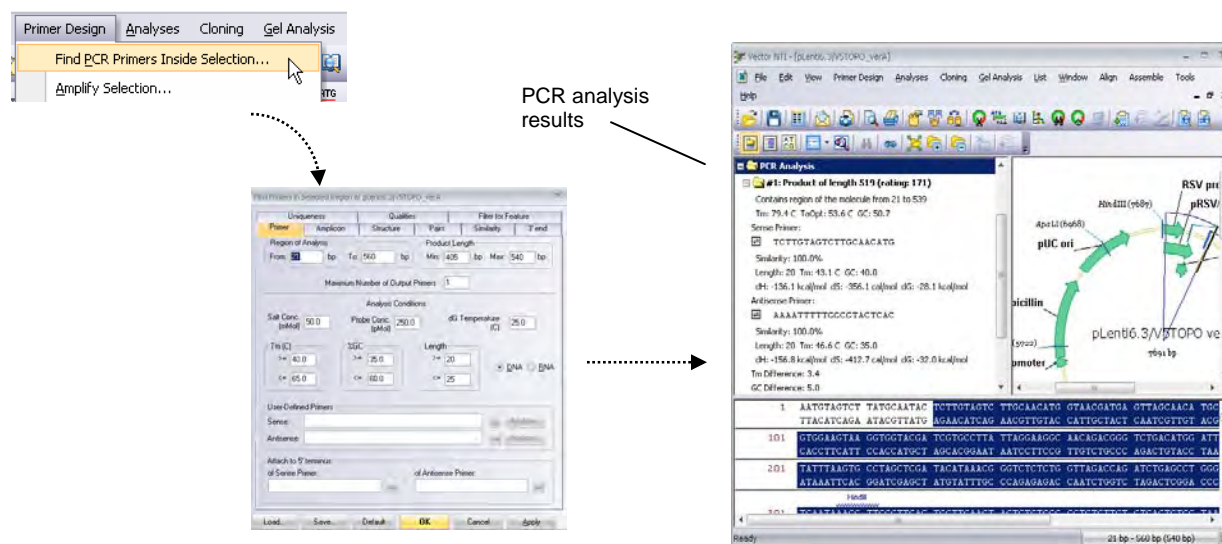
1. With the sequence in text format on the Windows® clipboard, click on the point in the Sequence Pane where you want to add the insert.
2. Click on CTRL + V on your keyboard.
3. The **Insert Sequence dialog** will open, displaying the sequence to be inserted.
4. Click on **OK** to complete the insertion.

# Designing PCR Primers from a Sequence

Vector NTI Advance® 11.5 can analyze a selected sequence and design PCR primers for it, based on parameters such as desired melting temperature (T<sub>m</sub>), GC content, and amplicon length.

With a DNA or RNA molecule open in the **Molecule Viewer**:

1. Select the part of the sequence for which you want to design primers, as described on the previous page.
2. Go to the **Primer Design** menu.
3. Select **Find PCR Primers Inside Selection** to find primers within the sequence (Figure 7), or **Select Amplify Selection** to find primers in the regions before and after the sequence (other amplification selections are available; see the Vector NTI Advance® 11.5 User's Manual for more information).
4. In the dialog box, select the desired primer-design parameters. Note that most of these parameters have default values based on typical PCR primers.
5. Click on **OK**. The results will appear under **PCR Analysis** in the Text Pane.

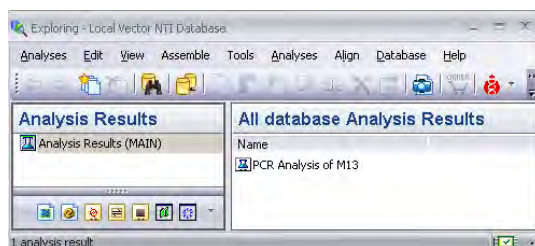


**Figure 7. Designing PCR primers within a selected region**

To save the PCR analysis results as a separate object in the database:

1. Right-click on the **PCR Analysis** folder in the Text Pane.
2. Select **Save as Analysis Result**.



The saved results will be listed under the Analysis Results object type in the Vector NTI Explorer (Figure 8).



**Figure 8. PCR analysis results listed in Vector NTI Explorer**

# Cloning Two Fragments with Clone2Seq

You can use **Clone2Seq** to easily clone two fragments in Vector NTI Advance® 11.5.

1. In the Molecule Viewer, go to **Cloning > Clone2Seq**, or click on the **Clone2Seq** button (  ) on the main toolbar.
2. To load a molecule (e.g., an insert or vector), click on **Open Molecule** in one of the window panes.
3. With the desired restriction enzyme sites displayed (click on the  button to select), select the first restriction site by clicking on it, then hold down the SHIFT key and click on the second site.

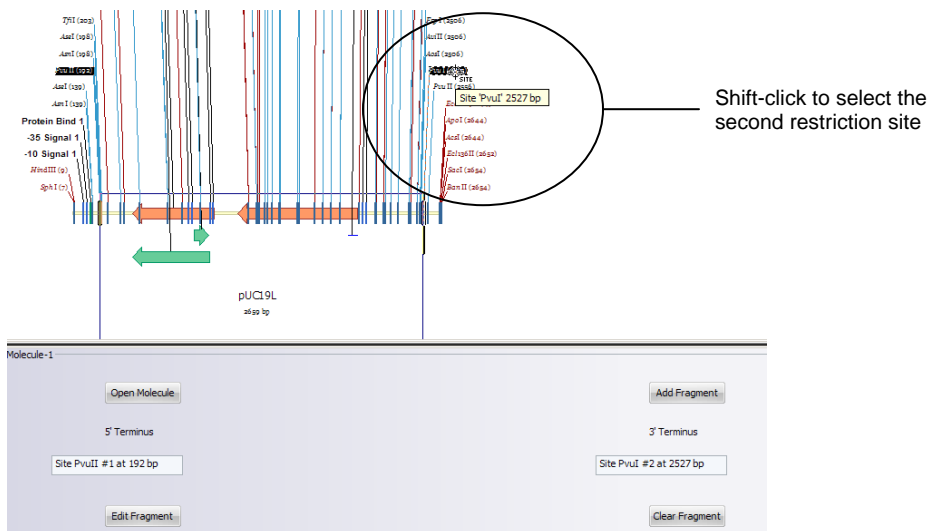


Figure 9. Selecting restriction sites in the Clone2Seq viewer

4. Add the selected fragment by clicking on **Add Fragment**.
5. Repeat these steps with the second molecule. Make sure the left terminus of the first fragment is compatible with the right terminus of the second fragment and vice versa.
6. When you have selected both fragments, click on **Clone**.

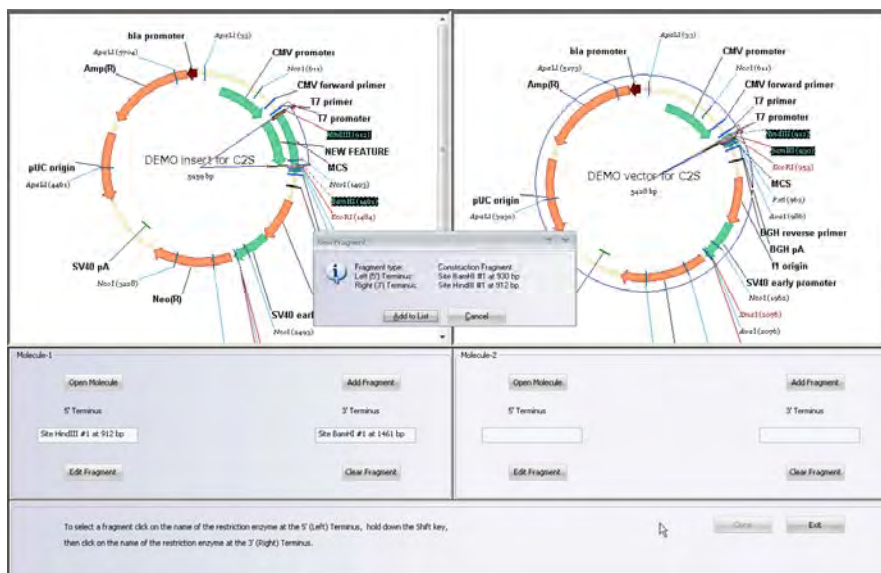


Figure 10. Clone 2 fragments with Clone2Seq

# In-Silico Gene Synthesis with ReGENERator

ReGENERator offers the fastest way to construct a DNA sequence based on the final protein molecule that you want to express. You can introduce amino acid mutations and specify the flanking sequences you need for expression, purification, or detection. When you are finished, you can send the DNA sequence directly to the GENEART® secure website ([www.geneart.com/vectormti](http://www.geneart.com/vectormti)) for rapid gene synthesis.

With a protein molecule loaded in the Molecule Viewer, select **Cloning > ReGENERator** or click on the



icon on the main toolbar to open ReGENERator. Using the tool, you can:

- Mutate the protein molecule by selecting amino acids in the Protein to Be Expressed window and inserting, deleting, or replacing single or multiple amino acids.
- Click on the **Change** button to select the desired Codon Usage Table for the organism.
- Select the desired Genetic Code from the dropdown menu.
- Add specific binding sites or adaptors to the ends of the DNA sequence by selecting the desired terminus (5' or 3') and clicking on the appropriate button (e.g., **Restriction Sites**, **Gateway Sites**, or **User Defined**).

To view the back-translated DNA, click **View In Silico DNA**.

To send the DNA sequence to GENEART® for synthesis, click **Send for Synthesis**.

The figure illustrates the ReGENERator workflow in four stages:

- Protein to be Expressed:** A list of amino acid sequences is shown. The first sequence is:
 

```
1  MPKKKPTPIQ LNFAPDGGVA VNGTSSAETN LEALQKKL
51  LEAFLTQKQK VGELKDDDFE KISELGAGWG GVVFVKVSH
101 HLEIKPAIRN QIIRELQVLH ECNSPIYVGF YGAFYSDG
151 SLDQVLKAG RIPEQLQKV SIAVIRGLTY LREKHKIM
201 SRGEIKLCDF GVSQQLIDSM ANSFVGTIRSY HSPERLQG
251 GLSLVEMAVG RYPIPPDDAK ELELMFGQV EGDAAETP
```
- Codon Usage Table:** A dialog box shows a list of organisms and their codon usage tables. The 'Standard' table is selected.
- Add Attachments to In Silico DNA Sequence:** A dialog box allows selecting the attachment type (5' terminus or 3' terminus) and the attachment type (Restriction Sites, Gateway Sites, or User Defined). The 5' terminus and Restriction Sites options are selected.
- VectorNTI - [DNA/RNA Molecule from document \_seq.txt]:** The final DNA sequence is displayed with various restriction sites and primers highlighted. The sequence is:
 

```
ATGCCCAAGA AGAAGCCAC CCCCATCCAG CTGAACCCCG CCCCCTCCGA CGGCAGCGCC GTGAACGGCA CCAGCAGCGC CGAGACCAA
TACGGGTTCCT TCTTCGGGTG GGGGTAGGTC GACTTGGGGG GGGGGGGGCT CGCGTCGGCG CACTTGCCTG GTCGTGGCG GCTCTGGTTT
PstI
TGCAGAAGAA GCTGGAGGAG CTGGAGCTGG ACGAGCAGCA CGGGAAGCGG CTGGAGCCCT TCCTGACCCA GAAGCAGAAG GTGGGGCAG
ACGCTTCTTT CGACTCTCTC GACTCTGACC TCCTGCTGCT CGCCTTCGCC GACTTCGGGA AGGACTGGGT CTTCGCTTC CACCOCCTCC
CGACTTCGAG AAGATCAGCG AGCTGGGGCC CGGCAACGGC GCGGTGGTGT TCAAGGTGAG CCACAAGCCC AGGGCCCTGG TGATGGCCG
CGTGAAGCTC TTCTAGTCGC TCGACCCGGC GCGGTGGCG CCGCACCACA AGTTCACACT GGTGTTGGGG TCGCCGGACC ACTACCGGG
PstI
CACTTGAGGA TCAAGCCCGC CATCCGGAAC CAGATCATCC GGGAGCTGCA GGTGTGCAC GAGTGAACA GCCCTACAT GCTGGGCTT
GTGGACCTCT AGTTCGGGGC GTAGGCCCTG GTCTAGTAGG CCCTCGACGT CCACGAGGTG CTCACGTTGT CGGGATGTA GCACCCGAA
```

Figure 11. In-Silico Gene Synthesis with ReGENERator



# Identifying Open Reading Frames (ORFs)

Vector NTI Advance® 11.5 can analyze a DNA/RNA molecule and identify the open reading frames (ORFs) in it, based on start and stop codons within the molecule.

With a DNA or RNA molecule open in the Molecule Viewer:

1. Go to the *Analyses menu* and select **ORF** (Figure 12).
2. In the dialog box, select the parameters for identifying and marking ORFs in the molecule.
3. When you click on **OK**, the sequences identified as ORFs will be marked with directional arrows in the Graphics Pane and Sequence Pane, and the ORFs will be listed in the Text Pane.
4. To identify an ORF in the different panes:
  - Click on a directional ORF arrow in the Graphics Pane to highlight its sequence in the Sequence Pane, or
  - Open a folder under **Open Reading Frames** in the Text Pane, right-click on the ORF name, and select **Find ORF** to highlight it in the Graphics and Sequence Panes.
5. To save an ORF to the feature map of the molecule, right-click on the ORF arrow in the Graphics Pane or the ORF folder in the Text Pane, and select **Add ORF to FMap**.

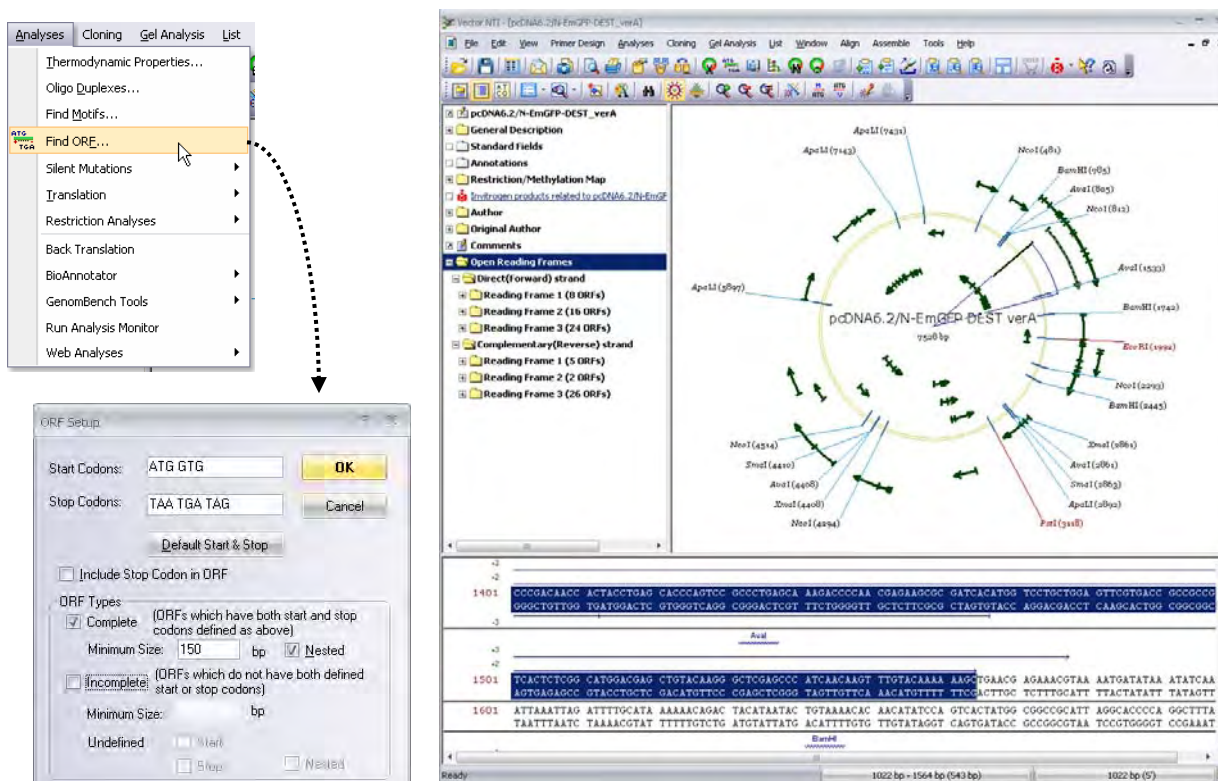


Figure 12. Identifying ORFs

# Creating a Restriction Map

Vector NTI Advance® 11.5 can analyze a DNA/RNA molecule and identify the restriction sites in it, using the software's comprehensive library of restriction enzymes.

With a DNA or RNA molecule open in the Molecule Viewer:

1. Go to the *Analyses menu* and select **Restriction Analyses > Restriction Sites** (Figure 13).
2. In the **Restriction Map Setup dialog**, review the list of restriction enzymes in the *Use Enzymes:* field. These are the enzymes that will be used to identify the restriction sites. Click on the **< Add, > Remove, and >> Remove All** buttons to add and remove enzymes from the list.

**Note:** If you click on **< Add**, the **Choose Database Enzymes dialog** will open, listing all the enzymes in the database. Select enzymes in the list by clicking on them or click on the **Select All** button, and then click on the **OK** button to add them to the **Restriction Map Setup dialog**.

3. Click on **OK** in the **Restriction Map Setup dialog**. The restriction enzymes and their binding sites will be shown in the Graphics Pane and Sequence Pane. The specific cut site of each enzyme will be listed under **Restriction/Methylation Map** in the Text Pane.

The figure illustrates the workflow for creating a restriction map in Vector NTI. It shows the **Analyses** menu with **Restriction Sites...** selected. The **Restriction Map Setup** dialog is shown with various options for enzyme selection and site sorting. The **Choose Database Enzymes** dialog lists various enzymes with their recognition strings and terminus types. The main software interface displays a circular **pTarget-DEST** map with various restriction sites marked, such as **EcoRI (5552)**, **BamHI (4)**, and **SmaI (11)**. The **Restriction/Methylation Map** in the Text Pane lists the enzymes and their recognition sites. The **Sequence Pane** shows the DNA sequence with a **BamHI** site circled.

**Restr. site shown in Graphics Pane**

**Rest. site shown in Sequence Pane**


| Name   | Recognition String | Terminus Type |
|--------|--------------------|---------------|
| AarI   | caactgc            | 5' Overhang   |
| AakII  | gacgtc             | 3' Overhang   |
| Acc65I | ggatcc             | 5' Overhang   |
| AccI   | gtmkac             | 5' Overhang   |
| AceIII | caagtc             | 5' Overhang   |
| AcII   | ccgc               | 5' Overhang   |
| AcII   | aacgct             | 5' Overhang   |
| AcuI   | ctgaag             | 3' Overhang   |
| AfeI   | agcgc              | Blunt         |
| AflIII | cttaag             | 5' Overhang   |
| AflIII | acrygt             | 5' Overhang   |
| AgeI   | accggt             | 5' Overhang   |

Figure 13. Creating a Restriction Map

# Aligning Molecules

Vector NTI Advance® 11.5 can align the sequences of two or more DNA/RNA molecules. The tool for doing this is called AlignX. This tool can be launched from either the Molecule Viewer or Vector NTI Explorer.

To align sequences using Vector NTI Explorer:

1. In the **Explorer**, select the molecules that you want to align using **SHIFT + CLICK** or **CTRL + CLICK** key commands (Figure 14).
2. Go the *Align* menu and select **AlignX—Align Selected Molecules**. The **AlignX Window** will open, with the molecules you selected listed in the upper left Text Pane.
3. In the **AlignX Window**, use **SHIFT + CLICK** or **CTRL + CLICK** key commands to select two or more molecules in the Text Pane list to align.
4. To begin the alignment, click on the **Align button** (  ) on the toolbar. The alignment may take several minutes, depending on the length and number of the molecules selected.
5. When the alignment is complete, the results are displayed in the **AlignX Window**, as shown in Figure 14. The **AlignX Window** has panes showing different similarity graphs and the points at which the sequences align.

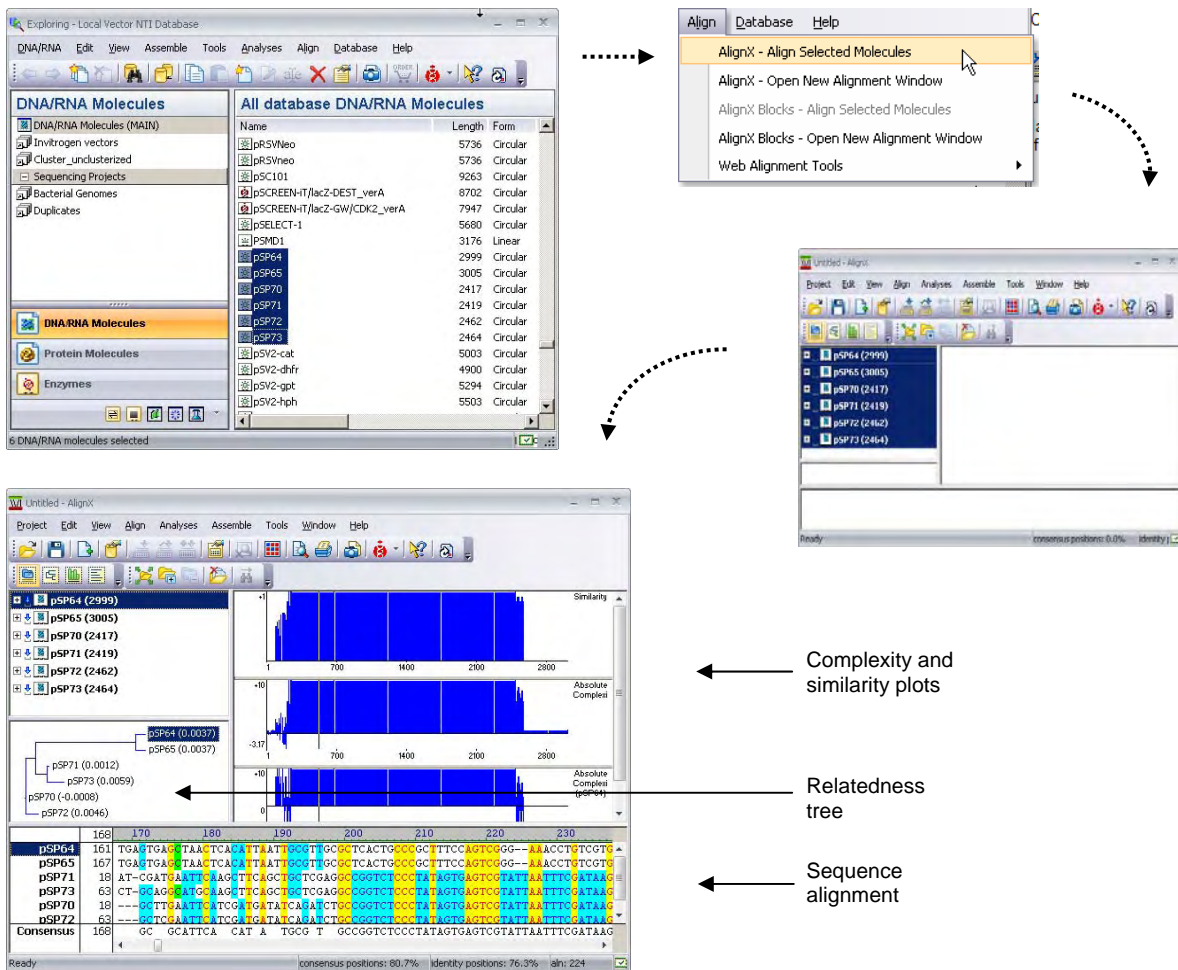



Figure 14. Aligning molecules

# Contig Assembly

Vector NTI Advance® 11.5 can be used to assemble DNA fragments—both text sequences and chromatograms from automated sequencers—into longer contiguous sequences or “contigs.” The tool for doing this is called ContigExpress.

In Vector NTI Explorer or the Molecule Viewer:

1. Go to the *Assemble menu* and select **ContigExpress—Open New Assembly Project** (Figure 154).
2. In the **ContigExpress Project Explorer**, go to the *Project menu* and select **Add Fragments >**. Select your fragment file type from the submenu list. The **Import Sequence dialog** will open.
3. In the **Import Sequence dialog**, navigate to the directory containing your fragment sequence files. Select the files and click on **Open**.
4. Depending on the file type, you may be prompted to list the fragments by their Windows® file names or by their internal fragment names. Select the desired option. The fragments will be loaded in the **ContigExpress Project Explorer**.
5. To view a particular fragment, double-click on it in the **Project Explorer** list. It will be loaded into the **Fragment Viewer**.
6. When you are ready to perform contig assembly, select the fragments in the **ContigExpress Project Explorer**.
7. Click the **Assemble Selected Fragments icon** () on the main toolbar. Fragments will be analyzed and assembled into one or more contigs, which will be listed in the **Project Viewer** along with the fragments in each contig.
8. Double-click on a contig in the list. It will be displayed in the **Contig Viewer**. The Sequence Pane at the bottom shows the sequence of the assembly. The Graphics Pane on the right shows the orientations of the fragments in the assembly. The Text Pane on the left lists the fragments in the assembly.
9. If you wish to edit the contig, enable the Enhanced Edit Mode by clicking the icon **Use Enhanced Edit Mode** (far left on the toolbar in the Contig window) before making any reasonable changes.
10. There are three trimming options in **ContigExpress**. Fragments can be trimmed for ambiguities, Phred quality scores, and vector contamination. Refer to the Vector NTI Advance® 11.5 User’s Manual for details.

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*Continued on the following page*



# Contig Assembly, continued

The process of adding fragments to a contig assembly project is shown in four stages:

- Menu Selection:** The 'Add Fragments' menu is open, and 'From ABI file...' is selected.
- File Selection:** A file explorer window shows the selection of 'ONE17R.abi' from a directory of 'Demo Projects'.
- History View:** The 'History View' shows 12 fragments in the project. The table below summarizes the data:

| Name      | Length | Original length | 5'Trimmed bases |
|-----------|--------|-----------------|-----------------|
| ONE17KANR | 759    | 759             | 0               |
| ONE2KANR  | 747    | 747             | 0               |
| ONE3KANR  | 755    | 755             | 0               |
| ONE4KANR  | 756    | 756             | 0               |
| ONE6KANR  | 770    | 770             | 0               |
| ONE8KANR  | 764    | 764             | 0               |
| ONE9KANR  | 758    | 758             | 0               |
| ONE10KANR | 758    | 758             | 0               |
| ONE11KANR | 755    | 755             | 0               |
| ONE13KANR | 819    | 819             | 0               |
| ONE15KANR | 784    | 784             | 0               |
| ONE16KANR | 767    | 767             | 0               |

The 'Fragment Viewer' shows the sequence and chromatogram for fragment ONE17KANR:

```

1 CAATT NTCGA TGATG GTTGA GATGT GTATA AGAGA CAGC
51 TCCTG CATTG GGAAG CAGCC CAGTA GTAGG TTGAG GCCC
101 CGCCG CAAGG AATGG TGCAT GCCAG GAGAT GGCTG CCCJ
151 GCCAC GGGGC CTGCC ACCAT ACCCA CNGCC GAAAC AAG
201 CGAAG TGGCG AGCCC GATCT TCCCC ATCGG TGATG TCG
251 CCAGC AACCG CACTG GTGGC GCGCG TGATG CCGGC CAC
301 GTAGA GGATC TGGCT AGCGA TGACC CTGCT GATTG GTT
351 NCGGG TCGCG GACGG CGTTA CCAGA AACCT AGAAG GTT
401 CCGAC TCTGA CCGCA GTTTA CGAGA GAGAT GATAG GGT
  
```

The 'History View' after assembly shows 14 items in 'Assembly 1':

| Name      | Length | Original length | 5'Trimmed bases |
|-----------|--------|-----------------|-----------------|
| Contig 1  | 3675   |                 |                 |
| ONE17KANR | 737    | 759             | 0               |
| ONE13KANR | 723    | 819             | 96              |
| ONE15KANR | 567    | 784             | 76              |
| ONE10KANR | 688    | 758             | 70              |
| ONE11KANR | 698    | 755             | 38              |
| ONE16KANR | 599    | 767             | 90              |
| ONE9KANR  | 719    | 758             | 38              |
| ONE6KANR  | 714    | 770             | 39              |
| ONE3KANR  | 704    | 755             | 38              |
| ONE2KANR  | 705    | 747             | 42              |
| Contig 2  | 764    |                 |                 |
| ONE8KANR  | 762    | 764             | 0               |
| ONE4KANR  | 719    | 756             | 37              |

The 'Fragment Viewer' also shows the sequence and chromatogram for Contig 1:

```

3290 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3300 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3310 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3320 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3330 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3340 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3350 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3360 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3370 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3380 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3390 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3400 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3410 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3420 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3430 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3440 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3450 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3460 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3470 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3480 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3490 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
3500 TACCGTATATCCACAGAAATCAGGGGATATACCAGGAAAGAAACATGTGTAGCCTAAAGGCCAGCA
  
```

Figure 15. Assembling a Contig

# Technical Support

Free technical support for Vector NTI Advance® 11.5 is available exclusively through the web. For more information, check out the Software Support section at [www.invitrogen.com/VectorNTI](http://www.invitrogen.com/VectorNTI).

To obtain personalized technical support by telephone or email, you must have an annual support contract. Users may purchase an Advanced Support Contract by contacting Invitrogen at [bioinfosales@lifetech.com](mailto:bioinfosales@lifetech.com).

For paid support, use the following contacts:

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